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RIPARIAN CORRIDOR VEGETATION STRUCTURE AND SOIL FUNCTION
ALONG URBAN, SUBURBAN, AND RURAL STREAMS IN LOUISVILLE, KY,
USA

By

R. Jonathan White
B. S., Western Carolina University, 1996
M. S., Western Carolina University, 2001

A Dissertation
Submitted to the Faculty of the
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in Partial Fulfillment of the Requirements
for the Degree of

Doctor of Philosophy

Department of Biology
University of Louisville
Louisville, KY

May 2011

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ABSTRACT

RIPARIAN CORRIDOR VEGETATION STRUCTURE AND SOIL FUNCTION ALONG URBAN, SUBURBAN, AND RURAL STREAMS IN LOUISVILLE, KY, USA

R. Jonathan White

May 13, 2011

Stream riparian zones are ecotones between terrestrial and aquatic environments.

Studying these areas in urban environments is important since they lie adjacent to stream water supplies. I conducted a study of riparian woody and groundcover vegetation along urban, suburban, and rural streams (land-use designations based on % impervious surface) to assess how cities are affecting plant community structure. I also studied riparian soil gas flux (carbon dioxide, methane, and nitrous oxide) in relation to water table depth and groundwater nutrient concentrations spanning a 10-month period (January to October, 2008). I found distinct woody and groundcover communities associated with proportion of impervious surface surrounding the research sites. These communities differed regarding diversity and proportion of native, exotic, and wetland species distributions. The most urban communities generally had lower species richness, more exotic species, and fewer wetland species when compared to rural areas. Urban areas also exhibited the highest streambanks and lowest water tables. Carbon dioxide gas flux rates were higher in urban areas, but methane and nitrous oxide fluxes did not respond uniformly to site differences as classified by proportion of impervious surface.

Methane and nitrous oxide differences, in addition to seasonal variability, were more greatly affected by local site level differences in substrate and nutrient ratios as well as soil moisture.

TABLE OF CONTENTS

ABSTRACT.....	iv
LIST OF TABLES.....	viii
LIST OF FIGURES.....	x
Chapter 1: Introduction.....	1
Chapter 2: Variation in woody streambank plant communities along urban, suburban, and rural streams in metro Louisville, Kentucky.....	14
Introduction.....	14
Methods.....	17
Results.....	33
Discussion.....	45
Conclusions.....	51
Chapter 3: Variation in riparian groundcover plant communities surrounded by urban, suburban, and rural land-use in Louisville, Kentucky.....	90
Introduction.....	90
Methods.....	94
Results.....	109
Discussion.....	120
Conclusions.....	127
Chapter 4: Seasonal greenhouse gas flux in riparian soils along urban, suburban, and rural riparian zones in metro Louisville, Kentucky.....	155
Introduction.....	155
Methods.....	159
Results.....	172
Discussion.....	178
Conclusions.....	187
Chapter 5: Conclusions.....	215
REFERENCES.....	219
APPENDICES	
Appendix 1.....	229
Appendix 2.....	232

Appendix 3.....	233
Appendix 4.....	234
Appendix 5.....	237
Appendix 6.....	243
Appendix 7.....	248
Appendix 8.....	250
Appendix 9.....	252
Appendix 10.....	255
Appendix 11.....	258
Appendix 12.....	261
Appendix 13.....	276
Appendix 14.....	279
Appendix 15.....	280
Appendix 16.....	281
 CURRICULUM VITAE.....	 282

LIST OF TABLES

Table 2-1: Area of research catchments and the number of study sites classified as urban, suburban and rural at the subcatchment scale, and within a 1-km and 500-m radius from each study site.....	53
Table 2-2: Distribution of riparian width intervals by stream order and land-use categories at the subcatchment scale.....	54
Table 2-3: Relative importance value of dominant (>5%) woody species by strata within land-use classes.....	55-57
Table 2-4: Tree frequency, mean density and basal area (\pm S.E.) in land-use categories at the subcatchment scale.....	58-60
Table 2-5: Sapling frequency and mean density (+ S.E.) in land-use categories at the subcatchment scale.....	61
Table 2-6: Tree seedling frequency and mean density (+ S.E.) in land-use categories at the subcatchment scale.....	62
Table 2-7: Shrub frequency and mean density (+ S.E.) in land-use categories at the subcatchment scale.....	63
Table 2-8: Tree diameter distribution in riparian zones by land-use category at the subcatchment scale.....	64-65
Table 2-9: ANOVA of woody plant richness by plant type among land use categories at the subcatchment scale.....	66
Table 2-10: Tukeys HSD pairwise comparisons of tree seedling richness by land-use category at the subcatchment scale.....	67
Table 2-11: Woody plant species composition for the most common species and significant indicator values for a 3-cluster configuration using optimal partitioning.....	68
Table 2-12: Woody plant species composition and significant indicator values for 5-cluster configuration using optimal partitioning.....	69

Table 2-13: Environmental and National Land Cover Database (NLCD) metrics used for non-metric multidimensional scaling.....	70
Table 3-1: Area of research catchments and the number of study sites classified as urban, suburban and rural at the subcatchment scale, and within a 1-km and 500-m radius from each study site	129
Table 3-2: Distribution of riparian width intervals in this study by stream order and land use categories at the subcatchment scale.....	130
Table 3-3: Mean % cover (+S.E.) for the top ten most abundant herb and woody vine species in urban, suburban and rural land-use categories....	131
Table 3-4: Riparian groundcover mean species richness by distance from the stream bank and land use categories determined at the subcatchment scale.....	132
Table 3-5: Frequently occurring and significant indicator species (using indicator species analysis) for 3-cluster configuration using complete clustering.	133
Table 3-6: Environmental and National Land Cover Database (NLCD) metrics used for non-metric multidimensional scaling.....	134
Table 4-1: Gravimetric soil water content (g water/g oven dry soil) in the upper 15-cm horizons within two meters of streambanks in urban, suburban, and rural riparian sites.	190
Table 4-2: Site mean vegetation data surrounding the static chambers at riparian sites.....	191
Table 4-3: Riparian groundwater nitrate+nitrite-N monthly means (\pm S.E.) for subcatchment land-use categories and total monthly rainfall from November 2007 to October 2008.....	192
Table 4-4: Riparian groundwater dissolved organic carbon (DOC) monthly means (\pm S.E.) for subcatchment land-use categories and total monthly rainfall.....	193
Table 4-5: Relationships between landscape-level soil factors and CO ₂ and CH ₄ fluxes in riparian areas across an urban-to-rural land-use gradient.....	194
Table 4-6: Relationships between plot-level soil factors and CO ₂ and CH ₄ fluxes in riparian areas across an urban-to-rural land-use gradient.....	195

LIST OF FIGURES

Figure 1-1: Conceptual model of variables that affect and are affected by vegetation within a riparian area.....	13
Figure 2-1: Schematic map of the three watersheds in Jefferson and Oldham counties in Kentucky.....	71
Figure 2-2: Plot and site design for sampling adult trees (≥ 2.54 cm DBH), saplings, tree seedlings, and shrubs.....	72
Figure 2-3: Silhouette plot of a cluster analysis resulting in three groupings of research sites.....	73
Figure 2-4: Bank height measurements for land-use categories determined at the subcatchment scale.....	74
Figure 2-5: Tree diameter distributions by land-use category in riparian zones.....	75
Figure 2-6a-b: Tree seedling richness/ $\log(\text{Area sampled})$ for (a) near-stream plots only ($n = 41$ nearstream plots), and (b) sites ($n = 41$ sites).....	76
Figure 2-7: Square root of the tree density averaged across all nearstream plots in each land-use category.....	77
Figure 2-8: Shannon's equitability index (E_h) for all woody species using nearstream plots only for comparisons.....	78
Figure 2-9a-b: Exotic shrub species richness (a) and density (b) proportions with increasing impervious surface (subcatchment scale).....	79
Figure 2-10: Partana ratios and overall silhouette widths for 2 to 10 clusters using optimal partitioning.....	80
Figure 2-11: Silhouette plot of cluster analysis using optimal partitioning and Bray-Curtis distance resulting in five groupings of research sites.....	81

Figure 2-12: 3-dimensional configuration (only 2 dimensions shown) of woody plant data using non-metric multidimensional scaling (NMS) with sites (points) assigned to their respective groupings from the 3-cluster configuration from cluster analysis.....82

Figure 2-13: 3-dimensional configuration (only 2 dimensions shown) of woody plant data using non-metric multidimensional scaling coupled with cluster assignments from the 5-cluster configuration from cluster analysis.....83

Figure 2-14a-b: Non-metric multidimensional scaling of woody plant data coupled with cluster assignments from the 3-cluster configuration using cluster analysis, shown as colored shapes..... 84

Figure 2-15a-b: Non-metric multidimensional scaling of woody plant data coupled with cluster assignments from the 5-cluster configuration using cluster analysis..... 85

Figure 2-16a-b: Non-metric multidimensional scaling of research sites arranged according to woody species composition.....86

Figure 2-17: Linear regression of *L. maackii* relative importance value (IV) against proportion of impervious surface within a radius of 500-m of site centers..... 87

Figure 2-18a-b: Regressions of tree seedling and sapling densities compared to *Lonicera maackii* density at each site.....88

Figure 2-19: New conceptual model of variables that affect and are affected by vegetation within a riparian area modified from Figure 1-1.....89

Figure 3-1: Schematic map of the three watersheds in Jefferson and Oldham counties in Kentucky.....135

Figure 3-2a-b: Plot and site structure for sampling adult trees (≥ 2.54 cm DBH), saplings, tree seedlings, and shrubs..... 136

Figure 3-3: Silhouette plot of a cluster analysis (complete clustering using Bray-Curtis distance) resulting in three groupings of research sites.....137

Figure 3-4: Groundcover species richness variation in nearstream plots with % impervious surface at the subcatchment scale..... 138

Figure 3-5a-c: Box-and-whisker diagrams of diversity indices computed for groundcover species in urban, suburban, and rural riparian zones..... 139

Figure 3-6a-c: Relationship between diversity measures of groundcover species at riparian plots along first order streams and % impervious surface cover within 1000-m of each plot.....140-141

Figure 3-7: Mean groundcover riparian species richness at indicated distance intervals from the streambank in urban, suburban, and rural riparian zones..... 142

Figure 3-8: Linear regression model of cumulative catchment area upstream of the subcatchment for each suburban nearstream plot against normalized groundcover species richness..... 143

Figure 3-9: Non-metric multidimensional scaling (NMS) 3-dimensional configuration of research sites arranged by groundcover species composition..... 144

Figure 3-10a-d: Non-metric multidimensional scaling (NMS) 3-dimensional configuration of research sites arranged by groundcover species composition and color-coded by cluster assignments from the 3-cluster configuration using cluster analysis..... 145-147

Figure 3-11: Non-metric multidimensional scaling of research sites arranged according to groundcover species composition..... 148

Figure 3-12a-b: Scatterplots of *Euonymus fortunei* percent cover in response to *Lonicera maackii* density..... 149

Figure 3-13a-b: Species richness (a) and Shannon’s diversity (b) of groundcover species in nearstream plots in response to *Lonicera maackii* density..... 150

Figure 3-14: Shannon’s equitability index of groundcover species in nearstream plots in response to *Euonymus fortunei* percent cover.....151

Figure 3-15a-c: Shannon’s Index (a), Simpson’s Index (b), and species richness (c) of groundcover species in nearstream plots in response to *Euonymus fortunei* percent cover.....152-153

Figure 3-16: New conceptual model of variables that affect and are affected by vegetation within a riparian area modified from Figure 1-1..... 154

Figure 4-1a-b: Air temperature and soil temperature taken during twice-monthly gas sampling visits in urban, suburban, and rural riparian sites in 2008..... 196

Figure 4-2: Gravimetric soil moisture in the upper 15-cm soil horizon in urban, suburban, and rural riparian sites in 2008.....	197
Figure 4-3a-b: Nearstream depth to groundwater (water table) in relation to bank height in years 2007 (a) and 2008 (b).....	198
Figure 4-4: Nearstream depth to groundwater (water table) from November 2007 to October 2008.....	199
Figure 4-5: Mean nitrate+nitrite-N nearstream riparian groundwater concentrations in urban, suburban, and rural subcatchments.....	200
Figure 4-6: Mean (\pm SE) riparian CO ₂ flux between January and October 2008 averaged for sites in rural, suburban, and urban subcatchments	201
Figure 4-7: Principal components analysis (PCA) of sites and monthly CO ₂ fluxes.....	202
Figure 4-8: Relationship between riparian winter CO ₂ flux and % soil organic matter (OM).....	203
Figure 4-9: Relationship between riparian spring CO ₂ flux and soil bulk density (BD).....	204
Figure 4-10: Increasing mean fall CO ₂ flux rates with increasing mean soil temperature among riparian sites.....	205
Figure 4-11: Mean (\pm SE) riparian CH ₄ fluxes from January to October 2008 averaged for sites in rural, suburban, and urban subcatchments.....	206
Figure 4-12: Principal components analysis (PCA) of sites and mean monthly CH ₄ fluxes from January to October 2008.....	207
Figure 4-13a-b: Mean monthly methane flux rates at each site from 2008 in relation to PCA row 1 scores from Fig. 13 for the months October (a) and September (b).....	208
Figure 4-14: Changes in mean summer CH ₄ flux in relation to subcatchment %IS.	209
Figure 4-15: Changes in mean spring CH ₄ flux with increasing mean soil temperature among sites averaged over April and May 2008.....	210
Figure 4-16: Summer mean net CH ₄ uptake (values less than zero) increases (values from left to right) with decreasing soil % carbon.....	211

Figure 4-17: Variation in site CH₄ flux with soil moisture.....212

Figure 4-18a-c: Relationships between mean fall 2008 CH₄ flux and soil molar C:N ratios (a) and gravimetric soil moisture (b).....213

Figure 4-19: Nitrous oxide flux from static chambers along riparian sites in three watersheds spanning an urban-to-rural gradient.....214

CHAPTER 1: INTRODUCTION: THE ECOLOGICAL STRUCTURE AND FUNCTION OF RIPARIAN ZONES

Riparian zone vegetation

Stream riparian zones are ecotones at the interface between terrestrial and aquatic systems. Therefore, width of the riparian zone, its plant communities and ecosystem processes are influenced by stream size, geomorphology, watershed drainage patterns and conditions in the adjacent stream channel as well as those in the upland catchment and upstream tributaries, (Fig. 1-1) (Dunne and Leopold 1978, Kalliola and Puhakka 1988, Naiman and Decamps 1997, National Research Council 2002). Riparian plant species survival and establishment can also be affected by biological factors that occur within the riparian zone, such as herbivory and competition for limiting resources (Naiman and Decamps 1997). However, flooding and sedimentation processes have been found to be the major determinants of the distribution of plant species in riparian areas (Robertson et al. 1978, Nilsson 1981, Hupp 1982, Kalliola and Puhakka 1988, Naiman and Decamps 1997).

Floodwaters not only disturb vegetation, but also deposit seed and other reproductive plant parts to downstream locations (Bendix and Hupp 2000). Nilsson et al. (1991) demonstrated the importance of water regimes in colonizing sites with seeds, suggesting that rivers can be significant corridors for some species of plants. For example, a regulated river exhibited the same riparian plant species richness as a non-

regulated river, but differed in plant species frequencies. Varied hydrological regimes can also alter success of seed dispersal and plant establishment. Greater flooding frequency may allow deposition of a greater number of water-vectored propagules. Studies within riparian wetlands in southeastern Alaska have shown that areas with intermediate flood disturbance supported more species than either permanently or rarely flooded sites (Pollock et al. 1998). As stream size increases, the areal extent of its influence on the land and vegetation also increases.

Riparian zones: role in the environment

Riparian systems and their vegetation can also affect the adjacent stream, influencing hydrological dynamics and chemistry, stream temperature, sediment load, and organic matter inputs (Fig. 1-1) (Naiman and Decamps 1997). Riparian zones filter nutrients and sediment from surface and ground-water flowing through them towards streams (Brenner et al. 1991, Gilliam 1994), thereby mitigating eutrophication of stream systems. Eutrophication of water bodies can have detrimental effects on water quality, which riparian vegetation buffers may act to mitigate. Vegetation in the riparian zone reduces the velocity and erosion potential of floodwater moving over the floodplain and prevents sediment from entering the stream (Wenger 1999, Easson and Yarbrough 2002, Geyer et al. 2003, Boothroyd et al. 2004). Plant litter, debris dams, and uprooted trees provide nutrients and habitat for stream organisms (Naiman and Decamps 1997, Wenger 1999). Trees and shrubs provide shade, benefiting light sensitive and shade-tolerant riparian plants as well as stream organisms that depend on relatively cool stream temperatures for survival (Kiffney et al. 2004). Therefore, riparian environments provide

many ecosystem services for society (Gregory et al. 1991) by improving quality of water entering streams, maintaining stream secondary productivity and therefore providing many recreational and human health benefits.

Riparian zones can act as effective sediment filters and nutrient sources or filters (Jacobs and Gilliam 1985a, Cooper et al. 1987, Lowrance et al. 1988) (Cooper et al. 1987, Puckett 2004). Riparian soils, since they lie adjacent to streams, have a water table relatively close to the soil surface. Coupled with periodic flooding, soil microorganismal processes can change in response to these moisture conditions based on the presence or absence of oxygen (Steudler et al. 1989). In addition to common soil functions that include nitrification, N-mineralization, and soil respiration, riparian soil functions can shift towards lesser or greater soil nitrate (NO_3^-) and nitrite (NO_2^-) consumption, or methane production or consumption based on soil moisture. Greater soil saturation would result in increases in the anaerobic processes of denitrification and methanogenesis. Consumption of soil NO_3^- and NO_2^- and production of N_2O and N_2 gases would result in removal of soil nutrients, but potentially contribute to atmospheric greenhouse gas concentrations if N_2O to N_2 production ratios increase. Under aerobic conditions and when nitrification processes are high, however, N_2O efflux from soils can be correspondingly high due to the "leaky pipe" phenomenon described by Davidson et al. (1993). Wet soils would also result in greater rates of methanogenesis, an anaerobic process producing the greenhouse gas, methane (CH_4). However, overall rates would depend on substrate availability and gaseous diffusion rates into and through the soil (Dorr et al. 1993, Schimel et al. 1993). The opposing process of soil CH_4 uptake by methanotrophic bacteria could dominate net CH_4 fluxes when soils are drier.

Riparian zones: effects of impervious surfaces

Urban and suburban land cover and land uses, both upstream and adjacent to a particular riparian stretch, can influence stream characteristics, riparian vegetation cover, and plant species composition (Fig. 1-1). Impervious surfaces in the catchment are largely responsible for the environmental effects of urbanization on streams and their riparian areas (Arnold and Gibbons 1996). Urbanization results in less water percolation into the soil and greater overland runoff and conduit flow that bypasses many of the water quality and water absorption services provided by riparian soils and vegetation (Paul and Meyer 2001, Zipperer 2002, Meyer et al. 2005). As compared to rural lands, urban and suburban riparian areas may contain higher species diversity, since overland flow is more frequent and intense (Walsh et al. 2005) due to greater amounts of runoff from impervious surfaces. Since increasing impervious cover causes an increase in the amount and proportion of overland vs. belowground water flow and increases water entry through storm drains, streams in urban and suburban areas become characterized by higher amplitudes in stream flow volume during flooding (Walsh 2000). In non-mountainous and non-coastal urban areas, frequent and smaller high stream flow events can cause deeper incision of stream banks and scouring of the streambeds than larger, infrequent events (Neller 1989, Walsh 2000). As incision increases, the water table in the riparian zone drops to coincide with the new lowered level of the stream surface (Groffman et al. 2002, Groffman et al. 2003). Urbanized watersheds with highly incised streams have the potential to severely lower the riparian water table, which changes the dynamics of processes within the upper layers of the soil environment. Exacerbating this are local-scale edge effects created by urbanization, which also contributes to soil drying

(Forman 1995, Zipperer 2002). As the amount of vegetation decreases within a riparian buffer, the amount of light that reaches the soil surface increases, which can alter the microclimate at the soil/atmosphere interface (Belnap et al. 2003). When this occurs, these once anaerobic, wet layers can more rapidly become aerobic, xeric layers, allowing organisms adapted to aerobic conditions to dominate the upper soil layers. This will therefore change nutrient cycling within riparian soils and, consequently, the chemical inputs from the riparian zone to the stream. Both lowered water tables and altered nutrient dynamics (i.e. carbon and nitrogen mineralization) can potentially change the types of plant species (e.g. wetland vs. upland species; proportion of nitrophilic species) that can persist within the altered riparian habitat. As the water table falls below the root zone, the ability of vegetation to sequester excess nutrients becomes more limited, thereby reducing the ecosystem services that they can provide (Groffman et al. 2002, Groffman et al. 2003, Groffman et al. 2004). Therefore, riparian vegetation and soil processes in urban and suburban areas are particularly important to study as they are key regulators of the aquatic-terrestrial linkage (Naiman and Decamps 1997).

Urban influences on riparian vegetation

Although impervious surfaces account for many of the environmental effects of urbanization on riparian vegetation, other urban influences on species diversity and exotic species composition are also important. These include increased fragmentation and edge effects, and shifts in abiotic and biotic influences from a highly altered matrix, including diverse and exotic seed sources from cultivated lots. Fragmented landscapes with a greater proportion of forest edges favor light-loving species over shade-tolerant species

that tend to grow within forest interiors (MacDougall and Kellman 1992). For example, the abundance of exotic plant species in forests has been directly linked to increased road density and use, and subsequent increased light availability. Parendes and Jones (2000) found light availability to be strongly correlated to habitat type within the H. J. Andrews Experimental Forest, where sites along high-use roads had the greatest amount of light penetration. Consequently, the greatest numbers of exotic species occurred along these stretches of road. Canadian riparian areas studied along an urban-to-rural gradient showed a trend towards greater numbers of opportunistic species, which tended to be exotic annuals, within the urban riparian areas (Moffatt and McLachlan 2004). Vulnerable species, classified as perennial natives, occurred primarily at the rural end of the urbanization gradient in the Canadian study. Urban effects associated with density of humans can dramatically alter riparian vegetation dynamics. Species diversity and composition can be impacted by surrounding land use via seed dispersal from residential yards and other landscaped open spaces. Forest remnants and corridors in areas with higher densities of people could receive more plant propagules from landscaped areas, which tend to be exotics (Kendle and Forbes 1997, McKinney 2004). Areas with lower human density and fewer landscaped areas should be less vulnerable to these external plant inputs. Exotic, opportunistic species, therefore, may exhibit greater success within riparian areas that have been disturbed by such factors in the surrounding urban environment (Vidra et al. 2006).

Influence of proximate land use and riparian soil moisture on plant communities and soils processes

Less understood factors related to riparian plant species dispersal and establishment include spheres of influence of different land-use types nearby (Fig. 1-1, Arrow 1), as well as differences in water table depths that differentially affect establishment (Fig. 1-1, Arrow 5a). Propagule dispersal and selection pressures on plant establishment differ for riparian and upland species, and depend on various spatial and environmental factors. Woody canopy species are more greatly influenced by macro-scale environmental factors, while understory herbaceous and groundcover species are more greatly influenced by micro-scale environmental factors (Guillaume 2002). Also, with increasing forest fragmentation in urban areas (Zipperer 2002), corridors promoting propagule dispersal for native forest species may become limited, thereby reducing the native species pool. Increased forest edge density and lower riparian water tables in cities and suburbs could also differentially affect plant species establishment by changing light availability, nutrient cycling and surface soil moisture. These conditions could favor upland woody species that are better adapted to drier soils, or to soil moisture regimes that vary greatly intra- and inter-annually.

The interaction of urbanization and physical properties of the riparian zone can also result in different attributes of the plant community. Plant diversity has been shown to vary with location within riparian zones. Lateral movement away from and longitudinal movement downstream have both been shown to relate to species diversity (Nilsson et al. 1994, Lite et al. 2005). A study of riparian plant species richness in arid environments in Arizona revealed decreasing herbaceous species richness moving

laterally from the stream channel during a dry year (Lite et al. 2005). This finding was attributed to lower water availability farther from the stream, as the water table was found to drop with lateral distance from the channel. Furthermore, the opposite was true following monsoon rains and flooding, as herbaceous species richness increased with distance from the stream channel following increases in the water table. A difference in plant species diversity with lateral distance from the stream between riparian areas with varying land-uses is less well understood.

In addition, catchment area and longitudinal location along a stream from upstream to downstream has been shown to affect plant diversity. Nilsson et al. (1994) showed that a river in Sweden had a higher mean species richness along the main channel than the tributaries draining in to the channel. Explanations offered for differences in species richness were increases in mean annual discharge, and amount of peat and silt cover when comparing the main channel and its tributaries. In contrast, Lite et al. (2005) found a pattern of increasing mean plant species richness in an upstream direction following summer monsoon and flooding events in an arid environment as herbaceous annual species rapidly responded to limiting resource availability. Whether these patterns exist across a land-use gradient with changing resource availabilities and disturbances has yet to be determined.

Riparian functional responses to urban and suburban land-use contexts that require further study include gas flux rates and soil processes in relation to urbanization and water table depths. These functional responses are important, because they determine whether riparian zones are sources or sinks for particular nutrients and greenhouse gases. As more people move into growing urban areas, these processes will grow ever more

important to climate modelers and land managers to formulate appropriate water management plans and atmospheric greenhouse gas predictions.

Riparian projects within this dissertation

I conducted three projects to determine riparian vegetation structure and gas flux dynamics along an urban-to-rural land-use gradient within three adjacent watersheds. The first project involved characterizing woody riparian plant communities and statistical explorations of factors likely to influence species composition of those communities. I did the same with the second project using herbaceous and groundcover species (woody vines) to determine the differing effects of site environmental variables on their abundance and diversity. In addition, the influences of impervious surface and National Land Cover Dataset land-cover types were explored at varying buffer distances around the sites. I also wanted to determine the correlations between defining species of the groundcover layer and those within the woody vegetation communities. The final project was aimed at evaluating the effect of urbanization on riparian functions by assessing soil gas flux responses to land-cover variables, soil nutrients, water table depth, and soil moisture differences.

The overarching goal of the first study was to determine how woody plant communities along streams in Louisville, Kentucky varied with degree of urban and suburban development in three watersheds. I accomplished this through a series of four objectives. The first objective was to determine differences in woody vegetation assemblages and investigate whether relationships with impervious surface cover and National Land Cover Database (NLCD) land-cover types could be detected at different

spatial scales. The second objective was to determine whether differences in diversity and identity of plant species could be related to this urban-to-rural land-use gradient. The third objective was to determine the extent to which native and non-native woody vegetation was associated with the various land-use and -cover types in these catchments. The final objective was to determine the extent to which wetland and upland species were associated with various land-use and -cover types.

The overarching goal of the second study was to determine how groundcover (herbs and vines) plant communities along streams vary with degree of urban and suburban development in three watersheds in Jefferson and Oldham counties in Kentucky. I proposed a series of five objectives to accomplish this goal. The first objective was to determine whether potential relationships existed between riparian groundcover vegetation and variation in impervious surface cover and NLCD land-cover types at different spatial scales. The second objective was to determine whether differences in plant species diversity could be related to changes in this urban-to-rural land-use, to increasing catchment area and to increasing lateral distance from the streambank. The third objective was to determine the extent to which native and non-native groundcover vegetation was associated with varying land-use and -cover types. The fourth objective was to determine the extent to which wetland and upland species were associated with various land-use and -cover types. The final objective was to determine whether groundcover species associations and diversity differed with the degree of colonization by the exotic shrub, *Lonicera maackii*.

The primary goal of the third study was to assess variation in soil gas fluxes in riparian soils in relation to soil nutrients, moisture, groundwater depth, and temperature.

Since depth to water table and soil moisture content is a known determinant of soil gas flux, the first objective, was to determine whether differences in nearstream water table depth existed in urban, suburban, and rural riparian zones. I predicted that urban riparian water tables would be lower when compared to suburban and rural reference riparian sites because of higher streambanks (see Dissertation Chapter 2). My second objective was to measure flux rates of three greenhouse gases (carbon dioxide (CO_2), methane (CH_4), nitrous oxide (N_2O)) from urban, suburban, and rural riparian zones. Assuming that plant root density and organic matter content and lability in riparian soils varied similarly along the land-use gradient, I predicted that CO_2 flux could be higher in urban areas due to the “urban heat island” effect whereby higher atmospheric temperatures would raise soil temperatures, thereby increasing microbial activity and root respiration. In addition drier soils should favor aerobic microbial activity over that of anaerobes, with the consequence that CO_2 efflux should dominate over CH_4 efflux. I also predicted that net methane uptake (methanotrophy) would occur within urban riparian sites, as urban soils were predicted to be driest among the three land-use types and methanotrophs are obligate aerobes. Conversely, I predicted a net methane efflux (methanogenesis greater than methanotrophy) from soils in rural reference sites, because shallow water tables and subsequent anoxic soil should favor methanogens. Since denitrification is also an anaerobic process, I predicted the greatest N_2O flux rates would occur in rural reference sites due to the shallower water table and higher soil moisture. However, because N_2O efflux from soil can also occur under aerobic conditions when nitrification rates are high, the direction and magnitude of the gradient effect on this process may not be as predictable. My third objective was to evaluate the influence of groundwater nitrate and

dissolved organic carbon (DOC) concentrations on N_2O flux rates, because denitrifying bacteria depend on DOC as an energy source and NO_3^- as a final electron acceptor. Therefore, I predicted higher concentrations of groundwater DOC and nitrates would result in greater nitrous oxide flux rates from the soil under anaerobic conditions. The fourth objective was to evaluate the influence of soil properties on greenhouse gas flux rates. I predicted % clay and soil bulk density would be negatively correlated with CH_4 uptake due to their impact on gas diffusion rates into and through the soil from both deeper soil horizons and from CH_4 in the atmosphere. I also predicted flux of CO_2 and N_2O would follow patterns others have found with regards to bulk density (Xu and Qi 2001), with decreasing flux rates at higher bulk densities due to fewer soil pore spaces for microbial activity.

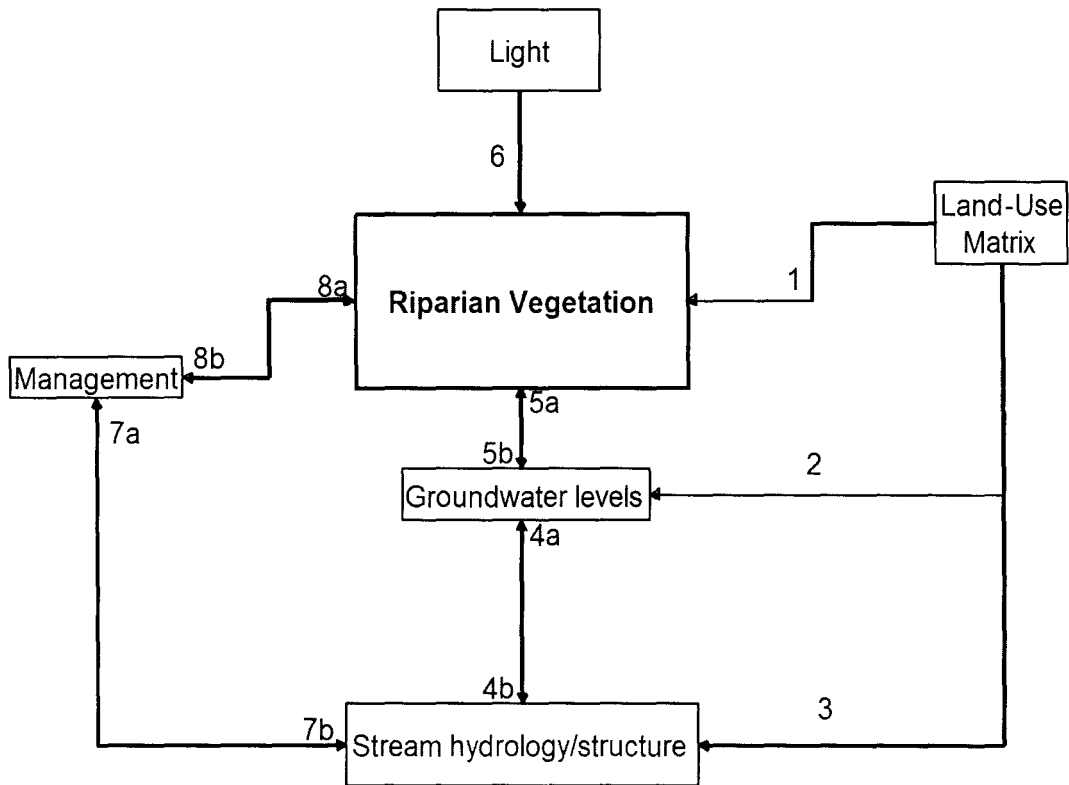


Figure 1-1: Conceptual model of variables that affect and are affected by vegetation within a riparian area. In this study, I will focus on the arrow labeled 1. The land-use matrix can affect riparian vegetation both directly (arrow 1) and indirectly (arrows 2 and 3), as illustrated. Stream hydrology and structure affect groundwater levels (arrow 4a), thereby indirectly affecting vegetation. Groundwater levels feed back on stream hydrology by determining stream depths and determining whether particular reaches are gaining or losing water (arrow 4b). Groundwater level can be altered through processes such as transpiration and degree of overland water flow and can determine which vegetation types can survive, depending on their root tolerance for wet, microaerophilic conditions (arrows 5a and 5b). The vegetation structure can affect the amount of light that permeates to the ground, and light affects the type of subcanopy vegetation that can grow based on light tolerance (arrow 6). Management decisions may be based on structure and/or water levels within the stream (arrow 7a), and some management decisions can affect stream levels and structure (arrow 7b). Highly incised streams have lowered water tables within the riparian zone. Management decisions or lack of management can determine dominant vegetation types (arrow 8a). Riparian species may be preferred over drier-adapted species when attempting to restore a riparian zone to a more natural vegetative state. Restoration management decisions are based on existing vegetation (arrow 8b).

CHAPTER 2: VARIATION IN WOODY STREAMBANK PLANT COMMUNITIES ALONG URBAN, SUBURBAN, AND RURAL STREAMS IN METRO LOUISVILLE, KENTUCKY

Introduction

Riparian zones adjacent to streams perform a number of valuable ecosystem services. As ecotones between terrestrial and aquatic systems, riparian areas act as sediment and nutrient filters (Brenner et al. 1991, Gilliam 1994), provide habitat and nutrients to stream and riparian organisms (Naiman and Decamps 1997, Wenger 1999), and reduce floodwater velocity and erosion potential (Wenger 1999, Easson and Yarbrough 2002, Geyer et al. 2003, Boothroyd et al. 2004). Riparian woody vegetation plays a significant role in providing these services (Brenner et al. 1991, Gilliam 1994, Naiman and Decamps 1997). Since riparian zones occur adjacent to streams and rivers, reductions of or alterations to these services would affect water quality, resulting in reduced recreation potential for swimming and fishing, and enhanced water treatment services and costs to towns and cities using these water sources for drinking. Because urban areas are growing and becoming more populous (Cordell and Macie 2002), the need to protect water resources from the effects of development will increase. Since woody vegetation within riparian zones provides such valuable services, it deserves particular attention to determine in what ways species structure will change as the land becomes more developed. In addition, depending on their sensitivity to land-use change,

riparian vegetation may also serve as an indicator of environmental change (Naiman and Decamps 1997).

Riparian woody species are more greatly influenced by macro-scale anthropogenic disturbances due to forest and river management rather than micro-scale influences such as site-level soil nutrient and moisture differences (Guillaume 2002). Along a river system in Sweden riparian sites along the main channel had higher mean abundances of species than along smaller tributaries (Nilsson et al. 1994). Furthermore, the middle reaches of the main channel exhibited the highest species richness, which may have been related to flooding differences. Differential effects of flooding dynamics on woody species composition and diversity have also been observed in other systems. For example, studies within riparian wetlands in southeastern Alaska have shown that areas with intermediate flood disturbance supported more species than either permanently or rarely flooded sites (Pollock et al. 1998).

Some of the ways urbanization can alter the environment that could then affect riparian woody species composition include lowering of the water table via stream incision (Groffman et al. 2003), increasing the proportion of forested edges (Forman 1995, Zipperer 2002), and producing more intense, flashy floods (Paul and Meyer 2001, Walsh et al. 2005). Water table depth changes are important since woody species vary in their ability to tolerate high moisture, low oxygen soil conditions (Naiman and Decamps 1997). Conversely, chronically low water tables can stress those species requiring higher moisture levels and favor species able to tolerate dryer conditions (Groffman et al. 2003). Forest edges, while contributing to species diversity, also attract non-native species (Fraver 1994, Zipperer 2002) due

to factors that include greater seed capture potential along forested edges (Cadenasso and Pickett 2001), higher light intensity and wind turbulence, greater temperature fluctuations, and lower soil moisture conditions (Gehlhausen et al. 2000, Zipperer 2002). Changes in flooding regimes can affect riparian vegetation communities due to changes in minimum and maximum flows (Auble et al. 1994, Naiman and Decamps 1997). These changes can result in reduced establishment and resource conditions that favor specific woody species over others (Bren 1992, Johnson 1993, Naiman and Decamps 1997). All of these changes can occur in predictable ways along an urban-to-rural gradient, resulting in distinct woody plant communities.

The overarching goal of this study was to determine how woody plant communities along streams in Louisville, Kentucky vary with degree of urban and suburban development in three watersheds. The first objective was to describe the woody vegetation assemblages and investigate whether relationships with impervious surface cover and National Land Cover Database (NLCD) land-cover types could be detected at different spatial scales. Specific predictions developed under this objective were that distinct woody vegetation communities existed that could be correlated with impervious surface cover, as an index of urbanization. The second prediction was that the sphere of influence of different land-use and land-cover types on woody vegetation communities would be scale-dependent and correlate more strongly at larger scales due to the relatively low magnitude of woody species response to microclimatic conditions (Guillaume 2002). The second objective was to determine whether predictable differences in diversity and identity of plant species existed along this urban-to-rural

land-use gradient. I predicted that woody plant diversity would be higher in suburban riparian areas than in either urban or rural riparian ones, because of the presumed availability of more diverse seed sources from low-density residential areas. The third objective was to determine the extent to which native and non-native woody vegetation was associated with the various land-use and -cover types in these catchments. Since one consequence of urbanization is creation of edges along vegetation patches, I predicted that native vegetation would dominate riparian assemblages associated with forested rural land-cover, and that exotic species would take on greater importance in riparian assemblages associated with urban land-use types. The final objective was to determine the extent to which wetland and upland species were associated with various land-use and -cover types. Because of greater impervious runoff and stream incision observed in cities, I proposed that higher streambanks would be found in urban areas, and that streambank height would be positively correlated with species adapted to drier soil conditions, and negatively correlated with wetland species. I specifically predicted that obligate and facultative wetland species would assume greater dominance in riparian assemblages associated with forested rural land-cover and that facultative upland and upland species would exhibit greater dominance in riparian assemblages associated with urban land-cover types.

Methods

Study Area

The study area consisted of three watersheds located within and adjacent to Louisville, Kentucky, USA (Latitude 38°15'N, Longitude 85°46'W), which has a

population of approximately 700,000 with a mean density of 695 people km⁻² (U.S. Census Bureau 2008). These were the Beargrass Creek (Middle Fork), Goose Creek, and Harrods Creek (South Fork) watersheds. The watersheds lie adjacent to each other with all streams flowing in a primarily westerly direction and eventually emptying into the Ohio River (Fig. 2-1).

The three watersheds are of approximately equivalent sizes, containing varying proportions of impervious surface cover (ISC; Table 2-1). Beargrass Creek watershed, located in the north-northeast portion of Jefferson County, KY, is approximately 65 km² (Metropolitan Sewer District 1999b) and contains the greatest proportion of impervious surface cover (33% ISC). This Middle Fork of Beargrass Creek joins the South Fork of Beargrass Creek before flowing into the Ohio River. Several stretches of this stream are featured on Kentucky's Division of Water 303d list, which list streams with impairment issues as well as the pollutant(s) causing the impairment (Kentucky Environmental and Public Protection Cabinet 2008). Since combined sewer overflows (CSOs) commonly occur in this stream, the presence of fecal coliform bacteria and sewage are stated as the problem pollutants within Beargrass Creek.

Goose Creek is a 50-km² watershed also in Jefferson County, in-between and adjacent to Beargrass and Harrods Creek watersheds (Metropolitan Sewer District 1999a). Of the three watersheds in this study, impervious surface cover is intermediate (20% ISC). The creek is composed of a pair of forks that meet before emptying into the Ohio River. Goose Creek is also listed on the 303d list, with cadmium and fecal coliform levels stated as impairment issues.

South Fork Harrods Creek watershed (60-km²), which has the lowest impervious surface cover of the three (10% ISC), is located primarily in neighboring Oldham County, but extends into northeast Jefferson County. The South Fork of Harrods Creek eventually joins the North Fork of Harrods Creek to become Harrods Creek before flowing into the Ohio River. Sections of the larger Harrods Creek network were added to the 303d list in years past, specifically the portion passing through Jefferson County, which is a more developed area. As in the other two watersheds, fecal coliform levels have been a persistent problem (Metropolitan Sewer District 1999c). The current 303d list contains Harrods Creek with fecal coliforms as an impairment issue, although this information is for the stream section downstream of where the North Fork and the South Fork Harrods Creek join.

Watershed classification

Watersheds were chosen from digitized catchment layers downloaded from the Louisville/Jefferson County Information Consortium (LOJIC) database (LOJIC 2010). LOJIC is a multi-agency institution that maintains a geographic information systems (GIS) database to serve primarily Jefferson County, KY, although limited information is available for a portion of Oldham County, KY. After checking for topographical accuracy using topographical maps of the watersheds and ArcGIS 9.0 software, I divided watersheds into subcatchments based on stream order and topography from the original LOJIC layers. I further divided subcatchments of higher stream orders with longer reaches into two or three constituent subcatchments based on topographic boundaries.

Beargrass Creek watershed contained 13, Goose Creek watershed 12, and Harrods Creek watershed 14 subcatchments.

Impervious land-cover classification was determined using three data layers from the LOJIC database digitized in 2001: buildings, miscellaneous transportation, and roads. The buildings-layer included all built structures, whether commercial or residential. The roads-layer included all paved road surfaces, excluding parking lots. The miscellaneous transportation-layer included secondary transportation features such as driveways, parking lots, and sidewalks. The area of land covered by impervious surfaces for each subcatchment was determined using ArcGIS 9 (ESRI, 2010) and was divided by the total land area for each subcatchment to determine the proportion of impervious surfaces.

Categorization of subcatchments into urban, suburban, and rural land-use types was modified from Schuler (1994). For this study, urban subcatchments were defined as containing $\geq 30\%$ impervious surface cover. Rural subcatchments contained $\leq 10\%$ and suburban subcatchments between 10% and 30% impervious surface cover. Research sites were classified by land use based on impervious surface cover at three scales. The first and largest areal scale was the subcatchment level, where sites were classified as urban, suburban, or rural based on the % impervious cover for subcatchment in which they were located. Impervious surface cover was calculated at two additional smaller scales surrounding the sites. Site buffers were created using ArcGIS 9 at a radius of 1-km and 500-m from the site center. The proportion of impervious surface cover was determined for these buffers in the same way as for the subcatchments. Sites were then classified into one of the three land-use categories in the same way as for the

subcatchment scale of analysis. Therefore, urban, suburban, and rural categorization of sites was changed depending on the scale at which impervious cover was calculated.

Site selection and plot establishment

Sites for plot establishment were chosen among riparian zones within the aforementioned subcatchments. A vertical line was drawn the length of the stream section within the subcatchment using ArcGIS 9.0 software and a digital map of the watersheds. A random number multiplied by 100 meters was used to find the location of the site along the vertical measurement line, starting at the downstream end. Sites were visited and chosen if they were at least partially covered with unmanaged vegetation and at least 100 meters from the nearest fork to avoid sampling riparian zones from separate subcatchments. Riparian zones were defined by topography and structural confinements, such as roads. Areas of level ground between the streambank and either an upland slope, structural barrier (roads, parking lots, buildings), or other obstruction (grazed pasture, ephemeral stream channel) were considered as being within the riparian zone. Riparian zones located completely within regularly used pastures and those found to be mown to the edge of the streambank were not used for this study. Only one side of the stream was sampled at each site. If both sides contained vegetation, the larger side was chosen, if it fit the criteria above, if it was reachable, and if the proper permissions could be attained. If only one side of the stream contained vegetation then that side was chosen. Locations where at least one full plot could not be placed (riparian width < 12-m) were sampled with a partial plot due to the difficulty in acquiring entry permission at many of the sites, and to avoid potential biases of purposefully choosing a site of a particular width. The

distribution of riparian widths by stream order and land-use category is provided in Table 2-2.

After visiting the locations identified by ArcGIS selection, the location of the actual study plot or plots was chosen by taking a randomly determined number of steps parallel to the stream to establish the center of a transect. A compass was used to extend a transect tape perpendicular to and away from the stream channel up to 91 meters, if possible. Circular plot centers (plot radius = 5.64-m, plot area = 100-m²) were established beginning at 6, 46, and 86 meters perpendicularly away from the stream channel. Three belt transects measuring 10 x 2 meters were nested within the circular plots, with the long edge perpendicular to the stream. The distance between midlines of adjacent belt transects was four meters. Twelve 1 x 1 meter quadrats were nested within the belt transects (four quadrats per belt transect). Quadrats were placed every two meters down the midline of each belt transect, so that the centers of adjacent quadrats within a belt transect were three meters apart (Fig. 2-2). Trees > 1 m high and ≥ 2.54 cm DBH were measured within the circular plots, living stem counts of shrubs and saplings (> 1-m high, ≤ 2.54 -cm DBH) were measured in the belt transects, and tree seedlings were measured in the 1-m² quadrats.

For each 100-m² plot, I determined the diameter at breast height (DBH) and number of trees by species of all trees > 2.54-cm DBH. Trees were determined to be inside or outside of the circular plot using a ny-pole (Nyland and Remele 1973). This was a wooden pole wide enough to be seen clearly from 6 meters. Two horizontal lines were positioned at eye level using different colored tape so that they overlapped flush when viewed through a prism (10 BAF) held at 5.64 meters from the pole. Standing next to

each tree, one edge of the prism was used to view the ny-pole (positioned in the plot center). If the lines on the ny-pole did not overlap or overlapped slightly, then the tree was counted within the boundary. If the lines overlapped flush with each other, then the tree was considered at the boundary of the plot, and every other tree in that category was counted. If the lines extended past each other, so that the top line appeared on the bottom and the bottom line appeared on the top, then the tree was outside the boundary of the plot and was not counted. For each 20-m² belt transect, shrub stem number (>1-m high) and sapling (> 1 meter high, < 2.54-cm DBH) number was recorded for each species using a meter stick held perpendicular to the transect tape with one end held directly over the tape. Any stem occurring beyond the far end of the meter stick was not counted. For tree and shrub seedling (<1-m high) counts and their respective % cover, four 1-m² quadrats were nested two meters apart within each of the three 20-m² belt transects (12 quadrats per circular plot). For all vegetation data, only living stems were considered. Tree, sapling, and shrub sampling was conducted in 2005 (June through August) and in 2006 (May to October). Tree seedling samples were made in 2006 (May – October).

Plant identification and classification

Plant identification was conducted in the field whenever possible. The primary source used for field identification was Wharton and Barbour (1973). Plants that could not be identified in the field were collected for later keying using Jones (2005).

Collections included as many parts of the plant as possible, including stem, leaf, and flowering parts. However, the authority for all species names used here was the USDA PLANTS Database (2010). Native status at the continental level was determined using

Jones (2005) and the USDA PLANTS Database (2010). Three categories of species provenance at the continental scale were used: native, exotic, and naturalized. Term usage is as defined in the USDA Natural Resources Conservation Service (2009). Native plants are those that have developed prior to European settlement in an area (in this instance, the continental U.S.). Exotic plants are those introduced with human assistance to a continent on which they were not previously found during the pre-European era. Naturalized species are exotic species that do not need human assistance to successfully reproduce and establish, thereby maintaining populations over an unspecified period of time. Inclusion of the naturalized species category was to distinguish exotic species in urban and/or suburban areas that were ornamental species likely coming from yards or cultivated landscapes.

Importance value and diversity

Relative importance value (IV) for each tree species (> 2.54-cm DBH) was calculated at each site (Appendix 1) and for each land-use category (Table 2-3). For analytical purposes, the term 'site' refers to the combination of all plots at a particular research location, to distinguish from those instances when only the plot nearest the stream was used. The IV at the site level was calculated as the sum of relative density ((Stem density for a single species (stems/ha)/Total stem density for all species (stems/ha)*100) and relative dominance ((Basal area for single species (m²/ha)/Total basal area for all species (m²/ha))*100).

Eq.1

$$\text{Tree (> 2.54-cm DBH) species \% IV} = \left(\frac{\text{species stems ha}^{-1}}{\text{total stems ha}^{-1}} + \frac{\text{species m}^2 \text{ ha}^{-1}}{\text{total m}^2 \text{ ha}^{-1}} \right) * 100$$

Sapling (> 1-m high, < 2.54-cm DBH), tree seedling (< 1-m high) and shrub IV at the site level was calculated from relative density ((species stems ha⁻¹)/(total stems/ha⁻¹)) and relative frequency (species occurrence per sampling unit [quadrats or belt transects]/total occurrences). For analytical purposes, only shrubs greater than one meter high were used to compute the IV's for each site, unless no shrubs of that species were present, in which case shrub seedlings (< 1-m high) were used. This was done to reduce the sampling bias caused by collecting stem counts at different scales (quadrat vs. belt transect) due to the clumping nature of shrub growth.

Eq. 2

$$\text{Sapling or Tree seedling or Shrub \% IV} = \left(\frac{\text{species stems ha}^{-1}}{\text{total stems/ha}^{-1}} + \frac{\text{species occurrence per sampling unit}}{\text{total occurrences}} \right) * 100$$

Species diversity was expressed using species richness, Shannon's (H') Index (Shannon 1948, Magurran 2004) and Simpson's Reciprocal (1/D) Index (Simpson 1949, Magurran 2004).

Eq. 3

$$H' = \sum_{i=1}^S (p_i \ln p_i)$$

Eq. 4

$$D = 1 - \frac{\sum_{i=1}^S n_i(n_i - 1)}{N(N - 1)}$$

Where S = species richness, p_i = proportion of an individual species, n_i = number of stems for a species, and N is the total number of stems. Shannon's equitability index (E_h) was also computed as a measure of community evenness:

Eq. 5

$$E_h = H' / \ln(S), \text{ where } H \text{ is Shannon's index and } S \text{ is species richness.}$$

Because of different riparian widths and plot sizes, richness was calculated for all sites as well as solely for nearstream plots as the number of species divided by the log of the area sampled (Conner and McCoy 1979, Nilsson et al. 1997). Species richness was calculated by totaling the species across all plots for a site, and in some other comparisons by considering richness in only the plot nearest the stream.

EstimateS (Colwell 2009) was used for computing diversity indices. Densities of woody species (trees, saplings, tree seedlings, and shrubs) were used to compute Simpson's reciprocal and Shannon's indexes for plots nearest the stream. Tree, sapling, and tree seedling information was collected at different spatial scales, so densities were computed separately for each life stage and added together where shared species existed

across life stages. In EstimateS, diversity index estimators were not used, so indices were computed using the observed species order within the input file.

Metrics for wetland indicator status (WIS) and species provenance (NEZ; N=native, E=exotic, Z=naturalized) were determined for each site to assess correlations with various land-cover variables. Species were labeled with their WIS from the USDA PLANTS database as well as NEZ as described previously. WIS and NEZ codes and definitions are shown in Appendix 2. A numerical value was assigned to each site for each category of WIS and NEZ using the relative IV's calculated for each species in each site. Values were calculated by summing the IV's of the species in each category and adding a weighting factor to account for the number of species within that category. The weighting factor was the proportion of species within each category divided by the total number of species within the site. Being based upon relative IV's, the maximum value for any category, all others being zero, was 900 (200 each for trees, saplings, tree seedlings, and shrubs + 100 for the weighting factor). For example, the relative IV for *Acer negundo*, a native tree, would be 200 if it were the only tree species in a site. If it was also the only sapling and seedling species located at that site then those IV's would also be 200. Since *A. negundo* is a native species the computed metric for the category "native" would be:

$$\text{Tree IV} + \text{Sapling IV} + \text{Seedling IV} + \text{Shrub IV} + \text{Weighting factor} \left[\frac{\# \text{ native spp.}}{\text{Total \# spp.}} * 100 \right] = \text{Native metric}$$

200 (Tree IV) + 200 (Sapling IV) + 200 (Seedling IV) + 0 (Shrub IV) + 1 native spp./2
total spp. *100 = 650

If a naturalized species, such as *Lonicera maackii*, were the only shrub, it would have an
IV of 200. The computed metric for the category “naturalized” in that same plot would
be:

Tree IV + Sapling IV + Seedling IV + Shrub IV + Weighting factor (# nat. spp./Total #
spp.*100) = Naturalized metric

0 (Tree IV) + 0 (Sapling IV) + 0 (Seedling IV) + 200 (Shrub IV) + (1 nat. spp./2 total
spp.)*100 = 250.

Data analysis

Potential relationships between plant species composition, degree of
imperviousness as well as other land-cover attributes, and other environmental
parameters listed below were explored using non-parametric ordinations (PC-Ord v. 4.41)
(McCune and Mefford 1999) and R v.2.11 software (R Development Core Team 2009),
indicator species analysis, and cluster analyses (R v.2.11) (R Development Core Team
2009). Scatterplots were used to explore relationships between diversity and abundance
measures of the woody vegetation strata with % IS at the subcatchment scale. Non-metric
multidimensional scaling (NMS) in PC-Ord was executed according to the recommended
procedure outlined in McCune & Grace (2002). For each analysis, a separate starting
configuration was supplied based on an initial analysis with the following configuration:

Sorensen distances, 50 runs with real and random data, a random starting configuration, and 200 iterations to evaluate stability of the final solution (stability criterion ≤ 0.0005). Dimensionality was assessed using a Monte Carlo test for significance, with a p-value indicating the probability of finding the computed stress value for a specific number of dimensions by chance (based on 1000 permutations). Final solutions were completed using one run with real data. Ordinations run using R were given an optimal geometric starting configuration based on principal coordinates analysis.

The goal of cluster analysis was to obtain groupings of study sites based on commonalities in species composition. To do this, a site dissimilarity matrix was constructed based on woody species composition. Several dissimilarity measures and clustering techniques were applied to the data in an attempt to achieve the best cluster configuration for the sites. A clustering technique known as optimal partitioning was decided upon as it produced the highest within-to-among cluster similarity value (Partana ratio). To accomplish this, the Bray-Curtis distance measure was used to create a dissimilarity matrix of the sites prior to cluster analysis. To decide on the number of groupings and assess their overall quality, I used the Partana ratio, as mentioned, as well as silhouette plots. Figure 2-3 shows an example of a silhouette plot for three groupings. Silhouette plots assign numerical values termed “silhouette widths” (S_i) to each site (i) with a value between -1 and 1 within a cluster. The silhouette width is obtained by comparing a site’s mean similarity to other sites within the cluster, followed by comparison to its mean similarity to sites within the nearest cluster. A silhouette width of 1 means the within-cluster similarity is much higher than between-cluster similarity, indicating a good fit of that site to the cluster. A value of -1 means the between-cluster

similarity is much higher than within-cluster similarity, indicating a poor fit of that site to the cluster. An average silhouette width is computed for each cluster to compare quality of the groupings. An overall silhouette width is computed for the whole graph to compare different numbers of groupings, where the goal is to obtain a value as high as possible without compromising the quality of the individual clusters.

Indicator species analysis (ISA) is a useful method for evaluating different species as indicators of environmental conditions. ISA is a statistical technique for species comparisons across two or more groups of sample units (sites, quadrats), taking into account abundance and frequency of each species within a group. An indicator value is assigned to each species in each group, and the value is tested for significance using a randomization procedure (Monte Carlo). Indicator values range from 0 to 1, where '0' means the species is not present in the group and '1' means it is always present within and exclusive to that group (Dufrene and Legendre 1997).

To determine whether measures of environmental degradation (i.e., bank height) and measures of abundance and species diversity (stem density, diversity indices) varied with the proportion of land cover, a single factor model (ANOVA, R v.2.11) was used for partitioning variance among land-use categories (urban, suburban, rural). Tukeys HSD pairwise comparisons were used for determining statistical significance of the variance between land-use categories. Data were assessed for homoscedasticity and homogeneity of variance prior to analysis and were transformed to better meet those assumptions, if necessary. In addition, any potential outliers were identified using quantile by quantile (Q-Q) plots and graphs of leverage vs. standardized residuals plotted with Cook's distance. Points appearing to assert a disproportional influence on the data were

sequentially eliminated from the analysis to assess effects on the statistical assumptions of homoscedasticity and homogeneity of variance. Data points were permanently removed from the analyses if their elimination improved these statistical assumptions.

Environmental parameters

Several environmental and socio-economic variables external to the plots were used to explore their ability to explain variation in plant species composition among plots. These were % impervious surface cover (at three scales: the subcatchment, within 1-km radius and within a 0.5-km radius of site center), canopy openness, bank height, plot size and distance from stream, stream order, cumulative upstream catchment area, and monetary value of residential property. Impervious surface cover was determined as described previously and the three scales were used to explore which scale better predicted plant community composition. Since canopy openness and hence light can also be a determinant of species presence and dominance, a spherical densiometer (Lemmon 1956, 1957), held over the middle of each 1-m² quadrat at each plot, was used to determine this measure. These values were averaged to obtain percent canopy openness for each plot. Cumulative catchment area was determined by adding subcatchment areas upstream of the site to the area of the subcatchment in which the site was located. This was done because catchment size is believed to affect stream conditions including flood potential, and therefore, it could be important for riparian plant composition. Bank height was measured from the first terrace out of the channel and referenced to the depth at the middle of the stream bed. Where the center of the channel was not reachable from the bank or by wading, bank height was measured at arm's reach. Bank height was measured

once at the midline of each plot nearest the streambank. These measurements were used as a proxy for depth to groundwater to determine correlations with species composition at the sites. Additional measurements were taken 50 meters to either side of the plot midline at 13 of the 41 sites where studies of depth to ground water and soil trace gas fluxes were conducted. These 13 additional measurements were averaged with the 41 site measurements to assess differences in overall bank height by land-use category.

To explore relationships between species identity, percent exotic species, and the measures of community diversity with a measure of socio-economic status of land owners, property value determinations were made from information at the Property Valuation Administrator offices in Jefferson and Oldham counties. The value of the parcel of land in which the site was located was taken as the property value. In instances where the site was located on the edge of a park abutting residential housing units or businesses, the average values of surrounding privately owned parcels was taken as the property value. Where sites were located in the interior of a park or natural area, the property was assigned a value of \$1 to indicate no private monetary value.

Finer-grained land-cover categorizations were also obtained using the U.S. Geological Survey's National Land Cover Database from 2001 (NLCD 2001; Appendix 3) acquired through LOJIC. NLCD 2001 is a database developed from a consortium initiated by the U.S. Geological Survey to interpret land-cover attribute categories from Landsat 5 and 7 images (Homer et al. 2004). Pixels from these images were assigned digitized land-cover attribute categories with a spatial resolution of 30-m. This digitized layer was used to determine the areal proportion of land represented by each land-cover category surrounding each study site within two circular areas of different sizes (1-km

and 500-m radius buffers). The proportions of these land-cover categories were entered as environmental variables into an ordination analysis for species composition at each site to determine which landscape scale correlated most strongly with species composition.

Results

Subcatchment delineation & classification

Forty subcatchments were delineated based on topography within and near the three main catchments. Sites were classified into land-use categories based on % impervious surface cover (ISC) at three scales: subcatchment, 1-km and 500-m radius site buffers (Table 2-1). Twelve subcatchments were classified as urban, with one in Goose Creek watershed and eleven in Beargrass Creek watershed. Rural subcatchments included eleven in Harrods and three in Goose Creek. Suburban subcatchments spanned all three catchments, with eight in Goose and three each in Beargrass and Harrods Creek watersheds (Table 2-1). Impervious surfaces within 500-m and 1-km of each site generally decreased compared with values at the subcatchment scale. Therefore, as the scale of the buffer distance around each site decreased, the number of sites classified as urban fell, while sites classified as suburban and rural increased (Table 2-1). One site in Beargrass Creek watershed on a privately owned farm was reclassified as rural when analyzed using the 500-m radius site buffer. The site was kept primarily in mown grass, with a thin strip of woody vegetation allowed to persist adjacent to the creek. All sites with $\geq 30\%$ ISC within 1-km and 500-m radii of a site were located in Beargrass Creek watershed.

Riparian bank height and zone width variation

Bank height at each site ranged from 15- to 230-cm. Average bank height was found to vary among the three land uses ($p = 0.08$; Fig. 2-4). Pairwise comparisons revealed urban streambanks (mean = 122-cm) were higher than suburban streambanks (mean = 86-cm; $p = 0.068$). Mean rural streambank height was 96-cm. Bank height followed a general upstream-to-downstream pattern of lower-to-higher banks among all land-use categories, with banks getting higher as upstream cumulative catchment area increased (data not shown).

Riparian width was measured at each site to determine the number of plots to establish per site. Suburban sites exhibited the smallest overall width, while rural sites were widest, thus permitting the greatest number of multi-plot sites (Table 2-2). Six of fourteen rural sites, four of fifteen suburban and three of twelve urban sites were less than 12 meters wide, resulting in smaller plot sizes in these sites. Only two urban sites contained more than one plot. One of the sites, located in a city park (Cherokee Park), contained two plots. The other site accommodated three plots, since it extended approximately 100-m before meeting a highway. All suburban sites contained only 1 plot as all vegetated riparian sites were less than 30 meters wide.

The distribution of riparian widths at urban and rural multi-plot sites was similar (Table 2-2). In rural subcatchments, two of the multi-plot sites were along 1st order reaches, one occurred along a 2nd order reach, and two lay adjacent to 3rd order streams. One of the rural multi-plot sites along a 1st order reach appeared to have an altered stream flowpath, as it was located along a residential street and directly abutted an adjacent

upland slope. The two multi-plot sites in the urban subcatchments occurred along 2nd and 3rd order reaches.

The distribution of narrow riparian zones with respect to stream order showed more variation between land-use categories (Table 2-2). Seven of the sites with riparian widths less than 12 meters were found along first and second order streams. In suburban subcatchments, three of the smaller plots were located along 1st order streams, and one was located along a 3rd order stream. Two of the urban riparian sites with less than 12-meter riparian widths were located along 1st order streams, and one along a 3rd order stream. Both of the narrow sites along 3rd order streams (one along urban and one along a suburban stream) appeared to have been dramatically altered, possibly to contain floodwaters. In the urban area, the first terrace out of the stream channel contained a narrow riparian zone (< 12-m wide) which ended at a vertical incline leading to a second terrace approximately 6 feet above the first terrace. The opposite bank was practically identical.

Plant Communities In Relation To Land-cover

The distribution of vegetative cover was uneven among the three land-use categories. Three rural sites and four rural nearstream plots had no trees. Two suburban sites had no trees, and all suburban sites were only wide enough for one plot. All urban sites and nearstream plots contained trees. Four rural, six suburban and four urban sites contained no saplings. When only nearstream plots were considered, the pattern remained the same. Two rural, two suburban and three urban sites contained no tree seedlings. The

distribution of tree seedlings was again the same for nearstream plots. Shrubs were absent from three suburban sites, but all other sites contained shrubs.

Urban, suburban, and rural woody species composition showed distinct patterns, but with common themes across categories. Each land-use category contained unique dominant woody species (Table 2-3). In addition, the most dominant species within land-use categories changed among woody vegetation strata, indicating some measure of succession is occurring within the three categories. The most abundant species also tended to be the more ubiquitous species across land-use categories, and woody species that were unique to a particular land-use category occurred in low frequencies (one or two sites; Tables 2-4 to 2-7). Shared species across all land-use categories included *Acer negundo* and *Celtis occidentalis* (Table 2-3 and 2-4). Both species had the lowest tree densities in rural riparian areas. *A. negundo* was also present in all woody vegetation strata across land-use categories (Tables 2-3 to 2-6). The most frequently occurring shrub species was the invasive exotic, *Lonicera maackii* (Table 2-7). It was located in a majority of all sampled sites and occurred in all urban locations. The mean density of *L. maackii* in urban sites was three times greater than in either suburban or rural areas. The only facultative wetland (FACW) shrub species, *Lindera benzoin*, occurred in all land-use categories, but it occurred most frequently and with the greatest density in rural areas. An obligate wetland (OBL) species, *Rosa palustris*, occurred in only one site, which was urban.

Tree diameter distributions also showed some patterns across land-use categories. Most trees ranged between 2.5- and 25-cm in diameter at breast height (DBH) (Table 2-8, Fig. 2-5). Rural riparian areas contained nearly half the number of trees as urban or

suburban areas in the 5-10 and the 10-25-cm diameter categories. Suburban areas contained the fewest trees greater than 35-cm DBH. In urban and rural subcatchments, *Platanus occidentalis* (FACW) only occurred in the higher diameter classes (>30-cm DBH) and only in the smaller diameter classes in suburban areas (5-10-cm DBH).

Species richness of woody vegetation can differ dramatically along an urban-to-rural gradient (Burton et al. 2005, Burton and Samuelson 2008). As such, species richness for woody plant communities was computed as one measure of species diversity and was computed individually for trees, saplings, tree seedlings, and shrubs. As area sampled differed among nearstream plots and among sites due to variation in number of plots per site, species richness was transformed as in Nilsson (1997) (species richness/log(area sampled)). Analysis at the nearstream scale was conducted to control for any differences in species diversity caused by varying riparian widths (site scale) across land-use categories, and also to more fully capture riparian plant diversity across the width of the riparian zone (site scale). Regressions of species richness of the various woody vegetation strata against %IS at the subcatchment scale revealed no strong trends. However, species richness patterns within limited ranges of impervious surface were evident. To explore these patterns I grouped site and nearstream plots into land-use categories based on proportion of impervious surface at the subcatchment scale. No significant differences in richness were evident among land-use categories for trees \geq 2.54 DBH, saplings, or shrubs (Table 2-9). Seedling richness, however, differed significantly among land-use types at both the nearstream and site scales of analysis (Fig. 2-6, Table 2-9). Tukey's HSD pairwise comparisons revealed suburban sites contained nearly twice the seedling richness (site and nearstream mean = 2.8 species) of urban sites

(site mean = 1.5 species; nearstream mean = 1.3 species) at both the site and nearstream scales (Table 2-10). *Acer negundo* and *Celtis occidentalis* were the most frequently occurring tree seedling species in urban (42% and 33% site occurrence, respectively), suburban (60% site occurrence for both species), and rural (43% and 36% site occurrence, respectively) riparian zones. These two species were also the densest in urban riparian zones (*A. negundo* – 3935 stems/ha; *C. occidentalis* – 1389 stems/ha). *C. occidentalis* (2556 stem/ha) and *Asimina triloba* (1444 stems/ha) were the densest in suburban riparian areas. In rural riparian zones, *F. americana* (1647 stems/ha) and *A. negundo* (1210 stems/ha) were the densest (Table 2-6).

Additional structural and community characteristics for woody species used for exploring relationships with land-cover attributes were stem density, tree diameter, and diversity indices (Shannon's, Simpson's reciprocal, Shannon's equitability). As with species richness, I used regression analysis to explore relationships with %IS at the subcatchment scale. Again, no strong correlations were found, so I grouped nearstream plots into the three land-use categories (urban, suburban, and rural) to further explore these relationships. Tree, sapling, shrub, and tree seedling densities were statistically analyzed at both site and nearstream scales within urban, suburban, and rural land-use categories (subcatchment scale). No statistically significant differences between land-use types (mean site-scale densities reported) were revealed in sapling (U = 521, S = 2092, R = 577 stems/ha), tree seedling (U = 5729, S = 11879, R = 5595 stems/ha), or shrub densities (U = 20219, S = 10276, R = 15920 stems/ha) even after eliminating differences in plot size by considering only full-size nearstream plots (n = 27 plots). I tested whether tree density and DBH varied by land-use category at the nearstream and site levels by

first using all sites and nearstream plots, and second using only treed sites and nearstream plots. Significant differences at the $\alpha \leq 0.1$ level were found among tree (≥ 2.54 -cm DBH) densities between land-use categories at the nearstream scale when all nearstream plots were used. Mean tree density (477 stems/ha) in rural plots was about half that in urban plots (mean = 954 stems/ha, $p = 0.073$; Fig. 2-7), with suburban plots being intermediate (705 stems/ha). When only treed plots were used no statistically significant differences were evident in tree densities among land-use categories. All urban nearstream plots contained trees ≥ 2.54 cm DBH. Two non-treed nearstream plots occurred in rural subcatchments (one was a 100-m² full size plot and the other was 50-m²). Four non-treed nearstream plots occurred in suburban subcatchments (two were in 100-m² full size plots and two were in plots smaller than 100-m²). All woody species (trees, saplings, seedlings, and shrubs) from nearstream plots were used to compute Shannon's diversity and Simpson's reciprocal indices, as well as Shannon's equitability index. No significant differences were observed among land-use categories for Shannon's and Simpson's diversity indices. Shannon's equitability index did, however, differ significantly among land-use types ($p = 0.047$), as urban sites demonstrated lower evenness than suburban sites (mean = 0.54 and 0.77, respectively) as determined by Tukey's HSD pairwise comparisons ($p = 0.037$; Fig. 2-8).

Most of the exotic species found were shrub species (Appendix 4). Scatterplots revealed an increasing proportion of exotic shrub species ($p = 0.0003$, Pearson's $r = 0.53$) and abundances ($p = 0.0003$, Pearson's $r = 0.54$) with increasing % ISC (subcatchment scale; Fig. 2-9). Above 10% ISC, at least half of the shrub species occurring within the sites were exotic (Fig. 2-9a). Above 30% ISC, at least 40% of all shrub stems were from

exotic species, with most sites containing 80% or greater exotic stem densities (Fig. 2-9b).

To control for differences in vegetation that could be related with stream order, I chose sites along first order streams to explore whether patterns in diversity and abundance could be detected by land use (Appendix 5). The strongest trend was a negative relationship between tree seedling density and %IS (subcatchment scale; $p = 0.07$, Pearson's $r = 0.39$). A similar negative correlation was observed between tree species richness and bank height ($p = 0.055$, Pearson's $r = 0.39$); however, this relationship was caused by a single influential point with the greatest proportion of impervious surface.

NMS & Cluster analysis

I conducted a cluster analysis to obtain groupings of research sites based on commonalities in species composition. I computed Partana ratios and silhouette widths for two to ten clusters to determine the number of clusters to use (Fig. 2-10). The 2-cluster configuration gave the highest Partana ratio, but when applied, all but one of the research sites were placed into a single cluster. Partana ratios were similar for three to ten cluster configurations, but overall silhouette widths differed more dramatically. I decided to use cluster regimes with three and five site groupings, since they exhibited the highest overall silhouette widths (0.18 and 0.16, respectively). The quality of the groupings can be seen in silhouette plots for the 3-cluster (Fig. 2-3) and 5-cluster configuration (Fig. 2-11). The 3-cluster configuration contained one cluster with 26 sites, a second cluster with 12 and a third cluster with 3 sites. The 5-cluster configuration resulted in more evenly

sized clusters (Fig. 2-11). Three of the clusters had fewer than ten sites each, with the largest cluster containing 20 sites. One of the clusters (Cluster-4) contained two sites and had a negative average silhouette width, indicating a poor grouping likely containing outliers.

Clusters were analyzed for differences in species composition using frequency of occurrence and indicator species analysis. Species composition of the 3-cluster configuration was relatively unique for each cluster (Table 2-11). The dominant species in cluster-1 was *Symphoricarpos orbiculata*, a native shrub. Cluster-2 was dominated by *Lindera benzoin* (native shrub) and *Asimina triloba* (native tree). Major species in cluster-3 were *Lonicera maackii* (exotic, invasive shrub) and *Celtis occidentalis* (native tree). Those species with significant frequencies of occurrence in each of the clusters ($p \leq 0.05$), determined using indicator species analysis, are shown in Tables 7 and 8. The shrub layer was found to be important in defining groupings, as each cluster contained a significant indicator shrub species, but not necessarily a tree, sapling, or tree seedling species.

The 5-cluster configuration showed similar species assemblages, but with more overlapping species assignments (Table 2-12). The *L. benzoin/A. triloba* cluster remained intact as cluster-1, with other notable species including *Aesculus glabra* and *Ulmus rubra* (native trees). Cluster-3 was the *L. maackii*-dominated cluster, also containing the nearly ubiquitous species, *A. negundo* and *C. occidentalis*. This was also the largest cluster in the configuration, containing 20 sites. Clusters -2 and -5 shared a common dominant species, *A. negundo*, although the form of the species differed among the groups, with trees dominating cluster-2 and seedlings dominating cluster-5. Both cluster-2 and cluster-

5 shared species with cluster-1, with *L. benzoin* in cluster-2 and the less frequent *Ligustrum sinense* (exotic shrub) in cluster-5.

Non-metric multidimensional scaling (NMS) was used to determine cluster associations with environmental variables and species attributes (e.g., native status, wetland indicator status). NMS was run using the 41 sites organized into a distance matrix using Sorensen's index based on woody species Importance Values (IV's). A three-dimensional configuration was chosen, achieving a final stress of 17.7 and instability of 0.00048 for the final configuration. No site overlap occurred in 3-cluster or 5-cluster groupings in three dimensions, indicating satisfactory agreement between the cluster analysis and the NMS solution (Fig. 2-12 & 2-13).

Clusters were analyzed in relation to environmental variables that might correlate with and potentially help explain the observed variation in species composition among clusters (Table 2-13). Environmental variables calculated for each site were entered into a site-by-variable matrix and included as a secondary matrix in NMS using PC-Ord. The biplot feature in PC-Ord was used to generate a line for each variable with an $r^2 \geq 0.15$ for one or more axes, with the direction of the line corresponding to the direction of positive correlation of the variable, and the length of the line indicating the strength of the correlation along one or more axes. Implied in the ordination plot but not visible, a vector of equal length but opposing sign points in the opposite direction to indicate a negative correlation.

Percent impervious surface and NLCD categories were included in the NMS as environmental variables (Fig. 2-14a-b) to evaluate cluster identity correlations with coarse and fine scale land-cover variables (Table 2-13; Fig. 2-14b). Cluster-3, having the

exotic shrub *Lonicera maackii* as its indicator species in the 3-cluster configuration, was the most “urban” of the clusters, being most strongly correlated with % IS within a 500-m radius of each plot, as indicated by the length of this vector (Fig. 2-14a). At this smallest scale, low-intensity developed land most strongly correlated with this cluster (Fig. 2-14b). This type of land-cover is typically composed of single-family residential housing, and was the most frequently observed impervious land-cover type found in this study. To a lesser extent, medium-intensity and open-space developed land also positively correlated with the species composition of cluster-3, followed by high-intensity developed land, which occurred around a few of the most urban sites. Cluster-2, having the native shrub *Lindera benzoin* as its indicator species, was most positively correlated with the proportion of deciduous forest cover. Cluster-1 with another native shrub *Symphoricarpos orbiculata* as its indicator species, was most visibly associated with the proportion of pasture covering the land within 500-m, with two of the sites containing the greatest proportion of pasture than any of the other sites (not shown).

The NMS configuration was the same for the 5-cluster assignment even as site cluster identities changed the perceived layout. Cluster-3 was the most “urban” cluster in the 5-cluster configuration (Fig. 2-15a). Clusters -1 and -2 were the “deciduous forest” clusters, and cluster-5 contained the two sites with the highest proportion of pasture (Fig. 2-15b.). Figure 2-12 also demonstrates that %ISC at the 500-m buffer most strongly explained the variation among clusters. Clusters-1 and -2 contained 15% or less ISC within 500-m of each site. Cluster-3 communities were surrounded by 15% or more ISC at each site. Sites in Cluster-5 were intermediately placed in ordination space and were associated with between 10 and 20% ISC within a 500-m radius. Association of %ISC

with Cluster-4 was not considered, as it was thought to be a poor cluster with two outlier sites.

Unique indicator species among the deciduous forest and urban clusters in the 3-cluster and 5-cluster configurations include the native species, *L. benzoin* and *A. triloba*, in the deciduous forest clusters (Cluster-2 and Cluster-1, respectively) and the exotic shrub, *L. maackii*, in the urban clusters (Cluster-3 in both configurations; Tables 7 & 8). The tree species, *C. occidentalis*, also occurred most frequently in the urban clusters. *L. maackii* is an exotic invasive shrub growing ubiquitously across the landscape in both riparian and upland habitats. It is considered a naturalized species as it successfully propagates itself independently of human interference. The other three species are native to the area. *L. benzoin* is also a facultative wetland species, occurring most frequently in wetland habitats (Appendices 2 & 4).

Wetland indicator and native status for each species (Appendix 2) was included in the analyses as environmental variables to assess which categories were associated with the same site clusters as the land-cover variables. In this way an indirect correlation could be drawn between these species attributes and land-cover variables. Figure 2-16a-b shows the direction of correlation of these attributes in relation to the various site clusters. Facultative wetland (FACW) and native species attributes most strongly correlated with the deciduous forest clusters (Clusters-2 and -1 in the 3- and 5-cluster configuration). Facultative upland species did not positively associate with any particular grouping of sites. *L. maackii*, while contributing to the arrangement of sites in the NMS configuration, was not included in the calculation of species attribute values due to its overwhelming dominance in the urban sites, which would have masked any associations

of urban clusters with species attributes of other species. Had it been included, the urban groupings (Cluster-3) would have shown a strong correlation with exotic species as it was the sole significant indicator species of those groupings (Table 2-11 & 2-12).

Since *L. maackii* was such an important species in urban clusters, I decided to compare its relative importance within sites with proportion of impervious surface at the subcatchment scale and within 500-m of the sites. *L. maackii* IVs significantly increased with increasing proportions of impervious surface within 500-m of site centers ($p = 0.000057$, $r^2 = 0.34$; Fig. 2-17). In addition, tree seedling densities and, to a lesser extent, tree sapling densities showed a markedly decreasing trend with increasing *L. maackii* site densities (Fig. 2-18). The highest seedling and sapling densities occur at the lowest densities (<5000 to 10000 stems/ha) of *L. maackii*, and the lowest seedling and sapling densities at high *L. maackii* stem densities >10000 stems/ha. These results show *L. maackii* to be an urban species with potentially negative impacts on the regeneration layer for other woody plants.

Discussion

This research revealed that some components of riparian woody species structure and composition, as well as density and diversity, correlated with land-cover metrics associated with urban land use. Distinct plant assemblages were found based partially on an urbanization gradient. The 3-cluster configuration (Fig. 2-12) highlights the primary species differences between the riparian sites (Table 2-11), essentially splitting them into “deciduous forest” (cluster-2) and “urban” (cluster-3) with a small “pastoral” cluster (cluster-1). All three clusters were distinguished by different indicator shrub species. *S.*

orbiculata, *L. benzoin*, and *L. maackii*, which respectively defined clusters -1, -2, and -3 (Table 2-11) and suggests that those species have differing tolerances for disturbance, such as flooding frequency and intensity, or other anthropogenically altered landscape level factors. The tree, *A. triloba*, was also an indicator species for cluster-2, and is listed together with *L. benzoin* as a riparian species by Samuelson and Hogan (2006). Cluster 2 being the least-urban, most-forested cluster (Fig. 2-14) indicates those particular species are likely sensitive to factors such as disturbance associated with pastured (free-roaming) livestock and those associated with urban development within a 500-m radius. *L. benzoin* was the most prominent facultative wetland species in this study, occurring almost exclusively in rural riparian habitats (Fig. 2-16, Table 2-11 & 2-12) at all scales. *L. maackii* has been positively correlated to % IS and urban areas in other studies (Luken and Thieret 1996, Hutchinson and Vankat 1997, Borgmann and Rodewald 2005). For example, Borgmann (2005) also found a positive relationship between the degree of urban land cover within 1-km of riparian areas and percent cover of *L. maackii*. Exotic species invasions in urban riparian zones has been correlated with greater natural (flooding) and anthropogenic (trampling) disturbances (Moffatt and McLachlan 2004, Burton et al. 2005, Burton and Samuelson 2008). These findings indicate that as areas become more urbanized, exotic shrubs, such as *L. maackii*, will assume greater occurrences and densities within riparian zones.

More discrete plant communities were found when the 5-cluster configuration (Fig. 2-13) was used for grouping communities than when the 3-cluster grouping was used. The “deciduous forest” cluster from the 3-cluster configuration was separated into two distinct clusters (Fig. 2-15). Cluster-1, containing both *L. benzoin* and *A. triloba* was

most similar to the original three-cluster assemblage. Cluster-2 also contained *L. benzoin*, with *A. negundo* assuming the greatest importance within the assemblage (Table 2-12). The relative absence of *A. triloba* from this cluster could indicate environmental factors non-conducive to the retention, establishment and growth of this species over the more ubiquitous *A. negundo* (Samuelson and Hogan 2006), which occurred within all clusters. The degree of sensitivity of *A. triloba* and *L. benzoin* to % ISC is shown in Figure 2-13. Cluster-1, containing both species, ends at 10% ISC when calculated within a 500-m radius of a site. Cluster-2 communities are found when ISC increases to 15%, suggesting that *A. triloba* is more tolerant to increasing urban conditions than *L. benzoin*.

Some of the urban factors that might affect riparian plant assemblages include human manipulation of riparian areas (mowing, selective cutting of brush), human choices for plantings and manipulation of land close to naturalized riparian areas (yard plantings, pesticide and fertilizer use, pets), urban wildlife effects (deer and small mammal herbivory and seed predation and dispersal), habitat fragmentation, edge effects, the urban heat island, air pollution and urban hydrological drought. In this study, I attempted to explore the possible effect of urban hydrologic drought (Groffman et al. 2003) on riparian species composition, since it has been implicated as a possible determinant of riparian species composition (Tickner et al. 2001, Burton et al. 2005). I used bank height as an indicator of this phenomenon, where the highest banks would indicate the possibility of urban hydrologic drought. I found no direct relationship between species composition or diversity and bank height, despite the fact that sites classified as urban at the subcatchment level were found to contain significantly higher streambanks than suburban areas. However, depth to groundwater does not necessarily

equate to soil dryness in the upper soil horizons where seedlings and shallow rooted species would be most affected. Other factors that may contribute to soil dryness include soil type and compaction, the urban heat island effect, proportion of impervious surface and frequency of storm drains, stream flooding frequency, riparian width and proportion covered by vegetation. The rarity of wetland species in urban riparian zones does indicate that soil dryness may contribute to their loss of dominance, but the absence of a correlation with bank height suggests that other factors may also explain their rarity. With respect to determining the potential effects of depth to groundwater as a control on riparian plant community composition, additional research should be focused on more directly eliciting the effects that soil dryness and interactions with other soil characteristics in upper horizons have on species composition via manipulated experiments.

A new conceptual model is presented in Fig. 2-19 to reflect the findings of this study and offer a potential area of focus for future studies. In these watersheds, depth to groundwater (as measured by bank height) does not appear to provide major selective pressures on species establishment and growth compared to some other factors. Degree of soil moisture as determined by precipitation patterns, as well as other environmental conditions (light penetration, proportion of watershed that is forested) may be stronger determinants of the species that successfully establish in urban and suburban environments.

Tree seedling richness was significantly lower in urban areas than in suburban areas (Table 2-8, Fig. 2-6), but tree seedling densities did not differ. Equally high seedling density indicates that the lower richness in urban areas is not due to fewer

individuals surviving current conditions, and instead suggests that a) the supply of propagules to urban areas is less diverse than in suburban areas, b) urban conditions only allow a very limited number of species to propagate, and/or c) biotic factors like selective herbivory, are affecting successful species establishment.

Both sapling and seedling densities exhibited a decreasing trend with greater *L. maackii* densities (Fig. 2-18a-b). Reductions in seedling and sapling densities under high proportions of exotic shrubs have been shown in other studies (Merriam and Feil 2002, Loewenstein and Loewenstein 2005, Burton and Samuelson 2008). Exotic invasive shrubs pose a growing threat to overstory species regeneration. Rural tree densities were also significantly lower than that in the other two land-use types determined at the subcatchment scale. Large tree diameters have been shown to contribute to greater spacing between trees (Porter et al. 2001), yet I found no significant differences existed in mean tree DBH between land-use types at the subcatchment scale. Differences in tree densities were attributed to the absence of trees at several rural and suburban sites, since elimination of those plots without trees resulted in no significant differences between land-use types in tree density. Plot size was not indicated as a major contributing factor to these differences in tree densities among land uses either, since a majority of the smaller plots for all sites contained trees, and half of the non-treed nearstream plots were full size (100-m²).

Woody plant diversity as measured by species richness, Shannon's and Simpson's diversity indices, and Shannon's evenness index did not significantly differ among land-use categories in this study. This trend was in contrast to that found by Burton et al. (2005) in the Atlanta, Georgia area, where Shannon diversity for woody species in

riparian zones increased with distance from the city center. In my study, woody plant community evenness was lower in urban areas than in suburban areas, indicating that, if these communities are not directly managed, natural succession in urban sites favors success of a few exotic shrubs. Restoration managers attempting to reclaim an urban riparian area should be selective when choosing species to include along streams. Also, some form of management plan may need to be formed to control for specific exotic plants, such as *L. maackii*, that limit understory growth.

Exotic, naturalized vegetation was more highly associated with the urban riparian communities due to the high prevalence of *L. maackii*. This shrub occurred in all land-use categories, but dominated urban sites, correlating most strongly with proportion of impervious surface within a 500 m site radius (Fig. 2-17). Brown and Peet (2003) showed higher species richness of both native and exotic species with flooding frequency in mountainous riparian areas. This was attributed to immigration-driven selection pressures due to propagule deposition and frequent disturbance. Similar findings were reported by Burton et al. (2005) and Burton and Samuelson (2008). These same pressures may be at work in urban riparian areas, coupled with conditions unique to urban environments (heat island, potentially drier soils, more edge habitat) that allow exotic generalist species to gain a foothold over native riparian species once they arrive.

Two of the more ubiquitous tree species across land-use types were *A. negundo* and *C. occidentalis*. Both species are tolerant of a wide range of soil and moisture conditions. *A. negundo* is the most widely distributed species within its genus in North America, extending from coast to coast and from Canada south to Texas and Florida. *C. occidentalis* is widely distributed within the eastern continental U.S., although it does not

extend as far north as *A. negundo*. Both species are equally drought tolerant, and able to withstand prolonged periods of inundation exceeding 100 days (Burns and Honkala 1990).

The findings regarding riparian widths have implications for restoration efforts. Width of the riparian zone has been shown to be important for maintaining amphibian and bird species (Rudolph and Dickson 1990, Kinley and Newhouse 1997) and for the ability of the riparian area to buffer undesirable nutrient inputs into streams from adjacent land. Reestablishing riparian zones with appropriate widths will require reverse engineering to counter the efforts that created the narrow riparian widths along high order streams in urban and suburban locations. This, along with hydrological characteristics of riparian zones as they relate to species composition and ecosystem services, should continue to be studied to determine species responses to development and the resulting alteration to riparian functionality.

Conclusions

Woody plant assemblages are influenced by the proportion of impervious surface within a 500-m to 1-km radius. Exotic shrubs, such as *L. maackii*, showed a high affinity for urban sites. Wetland species exhibited a low affinity for these urban sites, and instead occurred in those sites with a greater proportion of deciduous forest.

In addition to influencing species composition, the various growth stages of woody species displayed different patterns along this urban-to-rural gradient of riparian sites. Tree seedling richness declined in urban compared to suburban riparian areas. However, tree seedling densities did not differ among sites within these two land-use

categories. Tree, sapling, and shrub species richness did not significantly differ among sites across the urban-to-rural land use gradient. However, tree densities were significantly lower in rural riparian zones than in suburban or urban sites due to the higher number of plots with no trees in rural areas.

I expected certain environmental conditions, such as bank height, within the associated riparian area to influence plant species assemblages and diversity. While mean bank height was highest adjacent to urban riparian zones, it did not explain the variation in woody species composition across sites. Instead, the high prevalence of *L. maackii* in sites with relatively high proportions of impervious surface surrounding them appeared to exhibit the greatest associations with plant abundances. Sapling and tree seedling densities both declined with increasing densities of this exotic, invasive shrub.

Land managers and riparian restoration teams will need to consider these findings when considering woody species to plant in riparian areas. For example, woody species survivorship in different land-use contexts will vary, making the choice of species for introducing or re-introducing into the riparian zone more risky from a resource perspective.

Table 2-1: Area of research catchments and the number of study sites classified as urban (U), suburban (S) and rural (R) at the subcatchment (SC) scale, and within a 1-km and 500-m radius from each study site. Classification as urban (U), suburban (S) and rural (R) land cover was based on proportion of impervious surface with $\geq 30\%$ being urban, $\leq 10\%$ being rural, and between 10 and 30% being suburban.

Catchment	Area (km ²)	SC			1 km			500 m		
		U	S	R	U	S	R	U	S	R
Beargrass Creek	65	11	3	0	8	6	0	6	7	1
Goose Creek	50	1	8	3	0	8	4	0	6	6
Harrods Creek	60	0	3	11	0	1	13	0	2	12

Table 2-2: Distribution of riparian width intervals by stream order and land-use category at the subcatchment scale. Riparian zone width was measured along areas of level ground beginning at the streambank edge and extended until reaching a structural barrier (such as a road, parking lot, or building), upland slope, or other obstruction (grazed pasture, ephemeral stream channel). Classification as urban, suburban and rural land cover was based on proportion of impervious surface with $\geq 30\%$ being urban, $\leq 10\%$ being rural, and between 10 and 30% being suburban.

Land use	Stream Order	Riparian width (m)			
		<12	12-45	46-85	>85
Urban	1	2	4	0	0
	2	0	1	0	1
	3	1	2	1	0
Total		3	7	1	1
Suburban	1	3	7	0	0
	2	0	4	0	0
	3	1	0	0	0
Total		4	11	0	0
Rural	1	4	2	2	0
	2	2	0	1	0
	3	0	1	2	0
Total		6	3	5	0

Table 2-3: Relative importance value of dominant (>5%) woody species by strata within land-use categories at the subcatchment scale.

Land Use ₁	Strata	Species	Relative IV	
Rural	Seedling	<i>Fraxinus americana</i> L.	36	
		<i>Acer negundo</i> L.	28	
		<i>Acer saccharum</i> Marsh.	25	
		<i>Celtis occidentalis</i> L.	23	
		<i>Asimina triloba</i> (L.) Dunal	16	
	Saplings	<i>Asimina triloba</i> (L.) Dunal	54	
		<i>Robinia pseudoacacia</i> L.	35	
		<i>Celtis occidentalis</i> L.	25	
		<i>Acer negundo</i> L.	22	
		<i>Aesculus glabra</i> Willd.	20	
		<i>Ulmus rubra</i> Muhl.	20	
		<i>Fraxinus americana</i> L.	16	
	Trees	<i>Juniperus virginiana</i> L.	46	
		<i>Juglans nigra</i> L.	33	
		<i>Maclura pomifera</i> (Raf.) Schneid.	19	
		<i>Acer saccharum</i> Marsh.	16	
		<i>Acer negundo</i> L.	15	
		<i>Asimina triloba</i> (L.) Dunal	14	
		<i>Fraxinus americana</i> L.	13	
	Shrubs	<i>Symphoricarpos orbiculatus</i> Moench	58	
		<i>Lonicera maackii</i> (Rupr.) Herder*	45	
		<i>Lindera benzoin</i> (L.) Blume	32	
		<i>Ligustrum sinense</i> Lour.*	25	
		<i>Rubus</i> spp.	22	
	Suburban	Seedling	<i>Fraxinus pennsylvanica</i> Marsh.	30
			<i>Celtis occidentalis</i> L.	27
<i>Acer negundo</i> L.			24	
<i>Asimina triloba</i> (L.) Dunal			18	
<i>Acer saccharum</i> Marsh.			14	
Saplings		<i>Prunus serotina</i> Ehrh.	12	
		<i>Ulmus rubra</i> Muhl.	12	
		<i>Staphylea trifolia</i> L.	49	
		<i>Asimina triloba</i> (L.) Dunal	26	
		<i>Acer negundo</i> L.	14	
	<i>Fraxinus pennsylvanica</i> Marsh.	13		

Table 2-3 (continued)			
Suburban		<i>Fraxinus americana</i> L.	13
		<i>Celtis occidentalis</i> L.	13
		<i>Cornus</i> spp.	12
		<i>Ulmus rubra</i> Muhl.	11
	Trees	<i>Acer negundo</i> L.	46
		<i>Fraxinus americana</i> L.	21
		<i>Acer saccharum</i> Marsh.	20
		<i>Ulmus rubra</i> Muhl.	19
		<i>Staphylea trifolia</i> L.	17
		<i>Acer saccharinum</i> L.	16
		<i>Cornus alternifolia</i> L. f.	14
		<i>Tilia americana</i> L.	13
		<i>Morus alba</i> L.*	11
	Shrubs	<i>Lonicera maackii</i> (Rupr.) Herder*	56
		<i>Ligustrum sinense</i> Lour.*	49
<i>Lindera benzoin</i> (L.) Blume		26	
<i>Euonymus alatus</i> (Thunb.) Sieb.*		23	
<i>Rubus</i> spp.		18	
Seedling	<i>Acer negundo</i> L.	72	
	<i>Celtis occidentalis</i> L.	41	
	<i>Fraxinus americana</i> L.	21	
	<i>Aesculus glabra</i> Willd.	13	
	<i>Gleditsia tricanthos</i> L.	13	
	<i>Morus rubra</i> L.	13	
Urban	Saplings	<i>Celtis occidentalis</i> L.	57
		<i>Acer saccharum</i> Marsh.	27
		<i>Acer negundo</i> L.	26
		<i>Morus rubra</i> L.	15
		<i>Fraxinus quadrangulata</i> Michx.	15
		<i>Fraxinus americana</i> L.	13
		<i>Fraxinus pennsylvanica</i> Marsh.	12
Trees	<i>Platanus occidentalis</i> L.	37	
	<i>Morus alba</i> L.*	35	
	<i>Acer negundo</i> L.	29	
	<i>Celtis occidentalis</i> L.	27	
	<i>Fraxinus pennsylvanica</i> Marsh.	12	
	<i>Carya cordiformis</i> (Wangenh.) K. Koch	11	

Table 2-3 (continued)			
Urban	Shrubs	<i>Lonicera maackii</i> (Rupr.) Herder*	73
		<i>Cornus</i> spp.	29
		<i>Hibiscus syriacus</i> L.*	25
		<i>Lindera benzoin</i> (L.) Blume	18
		<i>Lagerstroemia indica</i> L.*	17
		<i>Ligustrum sinense</i> Lour.*	16

Land use category based upon % impervious surface at the subcatchment scale.
 Urban \geq 30%, Rural \leq 10%, Suburban between 10 and 30% IS

*Non-native species

Relative IV calculation is the sum of relative density and relative frequency for seedlings, saplings, and shrubs. Tree relative IV is the sum of relative density and relative basal area. Maximum value is 200. Only species with IV > 10 (5%) are shown.

Table 2-4: Tree frequency, mean density and basal area (\pm S.E.) in land use categories at the subcatchment scale.

Species	Urban			Suburban			Rural					
	Freq	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)	Freq	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)	Freq	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)
<i>Acer negundo</i> (FAC)	0.33	192 (97)	5 (3)	16 (8)	0.53	196 (75)	4 (2)	8 (2.6)	0.43	68 (29)	1.5 (0.8)	3.6 (1.6)
<i>Acer saccharum</i> (FACU)	0.17	42 (34)	0.72 (0.66)	4.3 (3.6)	0.27	86 (39)	1.4 (0.8)	5 (2)	0.21	42 (26)	1.6 (1.3)	7.3 (5.4)
<i>Aesculus glabra</i> (FACU)	0.08	8 (8)	0.55 (0.55)	6.7	0.07	7 (7)	0.01 (0.01)	0.2	0.14	29 (22)	0.1 (0.08)	0.8 (0.2)
<i>Carya cordiformis</i> (FACU)	0.08	25 (25)	2 (2)	17.5	0	0	0	0	0	0	0	0
<i>Celtis occidentalis</i> (FACU)	0.25	108 (58)	6 (3)	22 (6)	0.33	61 (29)	0.09 (0.06)	0.3 (0.1)	0.14	14 (10)	0.1 (0.1)	1 (0.7)
<i>Fraxinus americana</i> (FACU)	0.17	31 (21)	1 (0.9)	5.8 (4.5)	0.20	47 (27)	1.9 (1.3)	9 (5)	0.14	51 (43)	0.7 (0.7)	4.7 (4.4)
<i>Fraxinus pennsylvanica</i> (FACW)	0.08	43 (43)	0.1 (0.1)	1.5	0.07	18 (18)	0.02 (0.02)	0.2	0	0	0	0
<i>Juglans nigra</i> (FACU)	0.25	36 (20)	2 (1)	7 (2)	0.07	13 (9)	0.1 (0.1)	2	0.29	25 (15)	4 (2.5)	15 (7)
<i>Maclura pomifera</i> (UPL)	0.08	6 (6)	0.04 (0.04)	0.5	0	0	0	0	0.14	43 (31)	1.5 (1.3)	7.9 (1.4)
<i>Morus alba</i> (UPL)	0.17	205 (155)	3 (2)	16 (4)	0.07	20 (20)	0.54 (0.54)	8	0	0	0	0

Table 2-4 (continued)

Species	Urban			Suburban			Rural					
	Freq.	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)	Freq.	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)	Freq.	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)
<i>Morus rubra</i> (FACU)	0.08	13 (9)	0.3 (0.3)	4	0	0	0	0	0	0	0	0
<i>Picea pungens</i> (NI)	0.08	8 (8)	0.03 (0.03)	0.3	0	0	0	0	0	0	0	0
<i>Platanus occidentalis</i> (FACW)	0.25	38 (23)	11 (6)	43 (14)	0.07	7 (7)	0.03 (0.03)	0.4	0.07	7 (7)	1 (1)	13
<i>Prunus cerasifera</i> (NI)	0.08	21 (21)	0.3 (0.3)	3	0	0	0	0	0	0	0	0
<i>Quercus rubra</i> (FACU)	0.08	4 (4)	0.007 (0.007)	0.1	0	0	0	0	0	0	0	0
<i>Thuja occidentalis</i> (FACW)	0.08	3 (3)	0.002 (0.002)	0.02	0	0	0	0	0	0	0	0
<i>Ulmus rubra</i> (FAC)	0.25	41 (24)	1 (1)	5.4 (5.2)	0.40	118 (67)	1.5 (1.3)	3.7 (3.1)	0.14	25 (19)	0.07 (0.05)	0.5 (0.2)
<i>Acer saccharinum</i> (FACW)	0	0	0	0	0.07	27 (27)	0.8 (0.8)	11	0	0	0	0
<i>Asimina triloba</i> (FACU)	0	0	0	0	0.13	20 (14)	0.06 (0.05)	0.4 (0.3)	0.21	101 (86)	0.1 (0.1)	0.6 (0.5)
<i>Cornus alternifolia</i> (NI)	0	0	0	0	0.07	35 (35)	0.2 (0.2)	2.8	0	0	0	0

Table 2-4 (continued)

Species	Urban			Suburban			Rural					
	Freq.	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)	Freq.	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)	Freq.	Mean Density (stems/ha)	Mean Basal Area ₁ (m ² /ha)	Mean Basal Area ₂ (m ² /ha)
<i>Staphylea trifolia</i> (FAC)	0	0	0	0	0.07	48 (48)	0.03 (0.03)	0.5	0	0	0	0
<i>Tilia americana</i> (FACU)	0	0	0	0	0.13	40 (34)	0.7 (0.7)	5 (5)	0	0	0	0
<i>Carpinus caroliniana</i> (FAC)	0	0	0	0	0	0	0	0	0.07	12 (12)	0.03 (0.03)	0.4
<i>Cercis canadensis</i> (FACU)	0	0	0	0	0	0	0	0	0.07	14 (14)	0.1 (0.1)	1
<i>Cornus florida</i> (FACU)	0	0	0	0	0	0	0	0	0.14	22 (15)	0.02 (0.01)	0.1(0.003)
<i>Juniperus virginiana</i> (FACU)	0	0	0	0	0	0	0	0	0.07	71 (71)	3 (3)	39
<i>Prunus serotina</i> (FACU)	0	0	0	0	0	0	0	0	0.07	4 (4)	0.01 (0.01)	0.2
<i>Quercus alba</i> (FACU)	0	0	0	0	0	0	0	0	0.07	4 (4)	0.2 (0.2)	3
SUM		825 (87)	32 (3.4)	153 (3.4)		742 (74)	12 (1.2)	56.5 (1.2)		533 (57)	14 (1.5)	98 (1.5)

*Wetland indicator status - Frequency of occurrence in wetlands and non-wetlands: OBL >99% in wetlands, FACW 67-99% in wetlands, FAC equally likely to occur in wetlands and non-wetlands, FACU 67-99% occurrence in non-wetlands, UPL >99% occurrence in non-wetlands, NI = not indicated.

Land use categories based upon subcatchment % impervious surface. Urban > 30% (n = 12), Rural < 10% (n = 14), Suburban between 10 and 30% IS (n = 15). 1&2 Basal area is the average within sites in each land use category including sites that did not contain the species (subscript 1) and including only sites containing the species (subscript 2). If the species was located in only one site, no standard error was given.

Table 2-5: Sapling frequency and mean density (+ S.E.) in land use categories at the subcatchment scale.

Species	Wetland Ind. Status*	Urban		Suburban		Rural	
		Freq	Mean Density (no./ha)	Freq	Mean Density (no./ha)	Freq	Mean Density (no./ha)
<i>Acer negundo</i>	FAC	0.25	97 (71)	0.13	167 (155)	0.21	45 (27)
<i>Acer saccharum</i>	FACU	0.08	83 (83)	0.07	78 (78)	0	0
<i>Carya cordiformis</i>	FACU	0.08	14 (14)	0.13	49 (38)	0	0
<i>Celtis occidentalis</i>	FACU	0.33	519 (342)	0.20	45 (26)	0.14	61 (46)
<i>Cornus sericea</i>	FACW	0.08	19 (19)	0	0	0	0
<i>Fraxinus americana</i>	FACU	0.08	28 (28)	0.20	59 (39)	0.14	25 (18)
<i>Fraxinus pennsylvanica</i>	FACW	0.08	23 (23)	0.07	139 (139)	0	0
<i>Fraxinus quadrangulata</i>	NI	0.08	35 (35)	0.07	23 (23)	0	0
<i>Maclura pomifera</i>	UPL	0.08	5 (5)	0	0	0	0
<i>Morus rubra</i>	FACU	0.17	28 (19)	0	0	0	0
<i>Thuja occidentalis</i>	FACW	0.08	5 (5)	0	0	0	0
<i>Aesculus glabra</i>	FACU	0	0	0.07	11 (11)	0.14	42 (36)
<i>Asimina triloba</i>	FACU	0	0	0.20	606 (324)	0.43	287 (168)
<i>Cercis canadensis</i>	FACU	0	0	0.07	12 (12)	0	0
<i>Cornus spp.</i>		0	0	0.07	111 (111)	0	0
<i>Prunus serotina</i>	FACU	0	0	0.07	11 (11)	0	0
<i>Quercus rubra</i>	FACU	0	0	0.07	12 (12)	0	0
<i>Staphylea trifolia</i>	FAC	0	0	0.07	667 (667)	0	0
<i>Tilia americana</i>	FACU	0	0	0.07	12 (12)	0	0
<i>Ulmus rubra</i>	FAC	0	0	0.13	92 (72)	0.21	30 (17)
<i>Juglans nigra</i>	FACU	0	0	0	0	0.07	6 (6)
<i>Robinia pseudoacacia</i>	FAC	0	0	0	0	0.07	60 (60)

SUM 854 (174) 2093 (324) 554 (103)

*Wetland indicator status - Frequency of occurrence in wetlands and non-wetlands: OBL >99% in wetlands, FACW 67-99% in wetlands, FAC equally likely to occur in wetlands and non-wetlands, FACU 67-99% occurrence in non-wetlands, UPL >99% occurrence in non-wetlands, NI = not indicated.

Land use categories based upon % impervious surface at the subcatchment scale. Urban > 30% (n = 12), Rural < 10% (n = 14), Suburban between 10 and 30% IS (n = 15).

Table 2-6: Tree seedling frequency and mean density (+ S.E.) in land use categories at the subcatchment scale.

Species	Wetland Ind. Status*	Urban		Suburban		Rural	
		Freq	Mean Density (stems/ha)	Freq	Mean Density (stems/ha)	Freq	Mean Density (stems/ha)
<i>Acer negundo</i>	FAC	0.42	3935 (3060)	0.60	1167 (362)	0.43	1210 (573)
<i>Aesculus glabra</i>	FACU	0.08	139 (139)	0.20	222 (128)	0.14	60 (40)
<i>Betula alleghaniensis</i>	FACU	0.08	23 (23)	0	0	0	0
<i>Celtis occidentalis</i>	FACU	0.33	1389 (651)	0.60	2556 (1520)	0.36	843 (486)
<i>Fraxinus americana</i>	FACU	0.25	255 (151)	0.20	283 (157)	0.21	1647 (1098)
<i>Fraxinus spp.</i>		0.08	69 (69)	0	0	0	0
<i>Gleditsia tricanthos</i>	FAC	0.08	139 (139)	0	0	0	0
<i>Morus rubra</i>	FACU	0.17	93 (71)	0	0	0	0
<i>Platanus occidentalis</i>	FACW	0.08	69 (69)	0.07	111 (111)	0.07	79 (79)
<i>Acer saccharum</i>	FACU	0	0	0.27	833 (467)	0.21	992 (654)
<i>Asimina triloba</i>	FACU	0	0	0.20	1444 (977)	0.29	327 (152)
<i>Carya cordiformis</i>	FACU	0	0	0.07	222 (222)	0.07	60 (60)
<i>Cercis canadensis</i>	FACU	0	0	0.07	111 (111)	0	0
<i>Cornus drummondii</i>	FAC	0	0	0.07	222 (222)	0	0
<i>Fraxinus pennsylvanica</i>	FACW	0	0	0.07	1222 (1222)	0	0
<i>Fraxinus quadrangulata</i>	NI	0	0	0.07	242 (242)	0.07	159 (159)
<i>Liriodendron tulipifera</i>	FACU	0	0	0.13	167 (121)	0	0
<i>Prunus serotina</i>	FACU	0	0	0.07	424 (424)	0	0
<i>Quercus shumardii</i>	FAC	0	0	0.07	61 (61)	0	0
<i>Tilia americana</i>	FACU	0	0	0.07	56 (56)	0	0
<i>Ulmus rubra</i>	FAC	0	0	0.07	424 (424)	0.14	149 (120)
<i>Acer spp.</i>		0	0	0	0	0.14	89 (64)
<i>Juglans cinerea</i>	FACU	0	0	0	0	0.07	89 (89)
<i>Quercus rubra</i>	FACU	0	0	0	0	0.07	159 (159)
<i>Robinia pseudoacacia</i>	FAC	0	0	0	0	0.07	30 (30)
SUM			6111 (1114)		9768 (880)		5893 (668)

*Wetland indicator status - Frequency of occurrence in wetlands and non-wetlands: OBL >99% in wetlands, FACW 67-99% in wetlands, FAC equally likely to occur in wetlands and non-wetlands, FACU 67-99% occurrence in non-wetlands, UPL >99% occurrence in non-wetlands, NI = not indicated.

Land use categories based upon % impervious surface at the subcatchment scale. Urban > 30% (n = 12), Rural < 10% (n = 14), Suburban between 10 and 30% IS (n = 15).

Table 2-7: Shrub frequency and mean density (+ S.E.) in land-use categories at the subcatchment scale.

Species	Wetland Ind. Status*	Urban		Suburban		Rural	
		Freq	Mean Density (stems/ha)	Freq	Mean Density (stems/ha)	Freq	Mean Density (stems/ha)
<i>Cornus spp.</i>		0.08	799 (799)	0	0	0	0
<i>Euonymus alatus</i>	NI	0.17	69 (48)	0.33	389 (226)	0.07	32 (32)
<i>Hibiscus syriacus</i>	NI	0.08	660 (660)	0	0	0	0
<i>Hypericum prolificum</i>	FACU	0.08	14 (14)	0	0	0	0
<i>Lagerstroemia indica</i>	NI	0.08	417 (417)	0	0	0	0
<i>Ligustrum sinense</i>	FACU	0.08	389 (389)	0.40	2456 (1484)	0.29	1345 (1160)
<i>Lindera benzoin</i>	FACW	0.25	458 (309)	0.27	728 (418)	0.57	1712 (665)
<i>Lonicera maackii</i>	NI	1.00	8005 (2023)	0.60	2767 (1401)	0.86	2618 (783)
<i>Rosa palustris</i>	OBL	0.08	97 (97)	0	0	0	0
<i>Amelanchier arborea</i>	FAC	0	0	0.07	56 (56)	0	0
<i>Cornus racemosa</i>	NI	0	0	0.07	89 (89)	0.07	42 (42)
<i>Hibiscus syriacus</i>	NI	0	0	0.07	111 (111)	0	0
<i>Rubus spp.</i>		0	0	0.07	222 (222)	0.07	595 (595)
<i>Hydrangea arborescens</i>	FACU	0	0	0	0	0.14	179 (129)
<i>Symphoricarpos orbiculatus</i>	UPL	0	0	0	0	0.21	4536 (4273)
<i>Viburnum acerifolium</i>	UPL	0	0	0	0	0.07	60 (60)
SUM			10907 (1643)		6817 (1015)		11118 (1322)

*Wetland indicator status - Frequency of occurrence in wetlands and non-wetlands: OBL >99% in wetlands, FACW 67-99% in wetlands, FAC equally likely to occur in wetlands and non-wetlands, FACU 67-99% occurrence in non-wetlands, UPL >99% occurrence in non-wetlands, NI = not indicated.

Land use categories based upon % impervious surface at the subcatchment scale. Urban > 30% (n = 12), Rural < 10% (n = 14), Suburban between 10 and 30% IS (n = 15).

Table 2-8: Tree diameter distribution in riparian zones by land-use category at the subcatchment scale.

Land Use	Species	Diameter Class (cm)						
		2.5 - 5	5 - 10	10 - 25	25 - 30	30 - 35	35 - 40	>40#
Rural	<i>Acer negundo</i>	12	25	37		6		
	<i>Acer saccharum</i>	6		19		12	6	
	<i>Aesculus glabra</i>	25	19	6				
	<i>Asimina triloba</i>	56	6					
	<i>Carpinus caroliniana</i>		6					
	<i>Celtis occidentalis</i>	12	6	6				
	<i>Cercis canadensis</i>		6	6				
	<i>Cornus florida</i>	12						
	<i>Fraxinus americana</i>	12	19	12	6			
	<i>Juglans nigra</i>						12	19 (53)
	<i>Juniperus virginiana</i>		6	12	6	6		
	<i>Maclura pomifera</i>	12	31	6		6		6 (47)
	<i>Platanus occidentalis</i>					6		12 (68)
	<i>Prunus serotina</i>		6					
	<i>Quercus alba</i>				6			
	<i>Ulmus rubra</i>	12	25		6			
	Total		161	155	106	25	37	19
Suburban	<i>Acer negundo</i>	49	49	81	24	8		
	<i>Acer saccharinum</i>		8	16	8			
	<i>Acer saccharum</i>	16	16	49				
	<i>Aesculus glabra</i>		8					
	<i>Asimina triloba</i>	16	8					
	<i>Celtis occidentalis</i>	33		8				
	<i>Cornus alternifolia</i>	8		8				
	<i>Fraxinus americana</i>	8		24	8			8 (45.5)
	<i>Fraxinus pennsylvanica</i>	8						
	<i>Juglans nigra</i>	8		8				
	<i>Morus alba*</i>		8	8	8			
	<i>Platanus occidentalis</i>		8					
	<i>Staphylea trifolia</i>	16						
	<i>Tilia americana</i>	8		41				
	<i>Ulmus rubra</i>	16	49	57		8		
Total		187	154	301	49	16	0	8

Table 2-8 (cont.)

	<i>Acer negundo</i>	15	146	66	7			7 (93)
	<i>Acer saccharinum</i>					7		
	<i>Acer saccharum</i>	22	7	15				
	<i>Aesculus glabra</i>				7			
	<i>Betula alleghaniensis</i>	7						
	<i>Carya cordiformis</i>		7		7		7	
	<i>Celtis occidentalis</i>	7	22	29	7	7		14 (53)
	<i>Fraxinus americana</i>		22	22				
Urban	<i>Fraxinus pennsylvanica</i>	7	7					
	<i>Juglans nigra</i>			29	15			
	<i>Maclura pomifera</i>		7	7				
	<i>Morus alba*</i>		22	58				
	<i>Morus rubra</i>		7	7				
	<i>Picea pungens</i>		7					
	<i>Platanus occidentalis</i>						7	14 (93)
	<i>Prunus cerasifera*</i>			7				
	<i>Quercus rubra</i>	7						
	<i>Thuja occidentalis</i>	7						
	<i>Ulmus rubra</i>	22	15		7			
	Total		95	270	241	51	22	7

†Land use categories based upon % impervious surface at the subcatchment scale. Urban > 30% (n = 12), Rural < 10% (n = 14), Suburban between 10 and 30% IS (n = 15).

*Non-native species

Number in parentheses is the largest diameter tree within the size class.

Table 2-9: ANOVA of woody plant richness by plant type among land-use categories at the subcatchment scale. The seedling category includes tree seedlings only.

Scale	Plant type	p-value
Near-stream plots	Tree	0.46
	Saplings	0.32
	Seedlings	0.026*
	Shrubs	0.87
Sites	Trees	0.82
	Saplings	0.8
	Seedlings	0.034*
	Shrubs	0.89

*p = 0.05

Richness = # spp./ log(sampled area)

Site richness determined by treating all plots at multi-plot sites as one sampling unit.

Table 2-10: Tukeys HSD pairwise comparisons of tree seedling richness by land-use category at the subcatchment scale.

Scale	Pairing	Difference	Adj. p-value
Near-stream plots	Suburban-Rural	1.21	0.17
	Urban-Rural	-0.66	0.61
	Urban-Suburban	-1.87	0.024
Sites	Suburban-Rural	1.05	0.26
	Urban-Rural	-0.80	0.49
	Urban-Suburban	-1.85	0.028

Table 2-11: Woody plant species composition for the most common species and significant indicator values for a 3-cluster configuration using optimal partitioning. Frequency values were calculated based on the proportion of sites within the cluster in which the species occurred. Indicator values were calculated using indicator species analysis, where a value of 0 means the species is never present in the cluster and a value of 1 means the species is always present and exclusive to the cluster. Probabilities for the indicator values were determined using a Monte Carlo randomization technique. Compare with the 5-cluster configuration in Table 2-12. *Celtis occidentalis* was included despite having no significant indicator value since it occurred most frequently (although not exclusively) in that cluster among the three growth stages.

Cluster	Species	Common name	Frequency	Indicator value	probability
1	<i>Symphoricarpos orbiculata</i>	Coralberry	0.66	0.66	0.001
2	<i>Lindera benzoin</i>	Spicebush	0.8	0.76	0.004
	<i>Asimina triloba</i>	Pawpaw			
		-Trees	0.41	0.42	0.023
		-Saplings	0.66	0.67	0.01
		-Seedlings	0.5	0.48	0.032
3	<i>Lonicera maackii</i>	Amur honeysuckle	0.88	0.84	0.001
	<i>Celtis occidentalis</i>	Hackberry			
		-Trees	0.26	-	-
		-Saplings	0.34	-	-
		-Seedlings	0.57	-	-

Table 2-12: Woody plant species composition and significant indicator values (IV) for 5-cluster configuration using optimal partitioning. Frequency values were calculated based on the proportion of sites within the cluster in which the species occurred. Indicator values were calculated using indicator species analysis, where a value of 0 means the species is never present in the cluster and a value of 1 means the species is always present and exclusive to the cluster. Probabilities for the indicator values were determined using a Monte Carlo randomization technique. Compare with the 3-cluster configuration in Table 2-11. Other species with frequencies ≥ 0.4 were included to illustrate species differences and similarities among groups.

Cluster	Species	Common name	Freq.	IV	Prob.	Other spp. (freq. ≥ 0.4)
1	<i>Lindera benzoin</i>	Spicebush	1	0.7	0.001	<i>Celtis occidentalis</i> , <i>Ulmus rubra</i>
	<i>Asimina triloba</i>	Pawpaw				<i>Aesculus glabra</i> seedlings, <i>Ligustrum sinense</i>
		Trees	0.55	0.56	0.01	
		Saplings	1	1	0.001	
		Seedlings	0.66	0.59	0.02	
2	<i>Acer negundo</i>	Boxelder	1	0.68	0.001	<i>Celtis occidentalis</i> seedlings, <i>Lindera benzoin</i>
3	<i>Lonicera maackii</i>	Amur honeysuckle	1	0.57	0.001	<i>Acer negundo</i> seedlings, <i>Celtis occidentalis</i> saplings/seedlings
4		Outlier cluster				
5	<i>Acer negundo</i>	Boxelder (seedlings)	0.8	0.5	0.03	<i>Symphoricarpos orbiculata</i> , <i>Ligustrum sinense</i> , <i>Ulmus rubra</i> saplings

Table 2-13: Environmental and National Land Cover Database (NLCD) metrics used for non-metric multidimensional scaling.

Environmental variables	NLCD values (500-m)
-Bank height adjacent to the site	-Open-space developed
-% impervious surface: subcatchment, 1-km buffer, 500-m buffer	-High-, medium, low-intensity developed
-Area sampled	-Deciduous forest
-Stream order	-Mixed forest
-Catchment Area	-Herbaceous forest
-Property value	-Crops
	-Pasture
	-Wetland

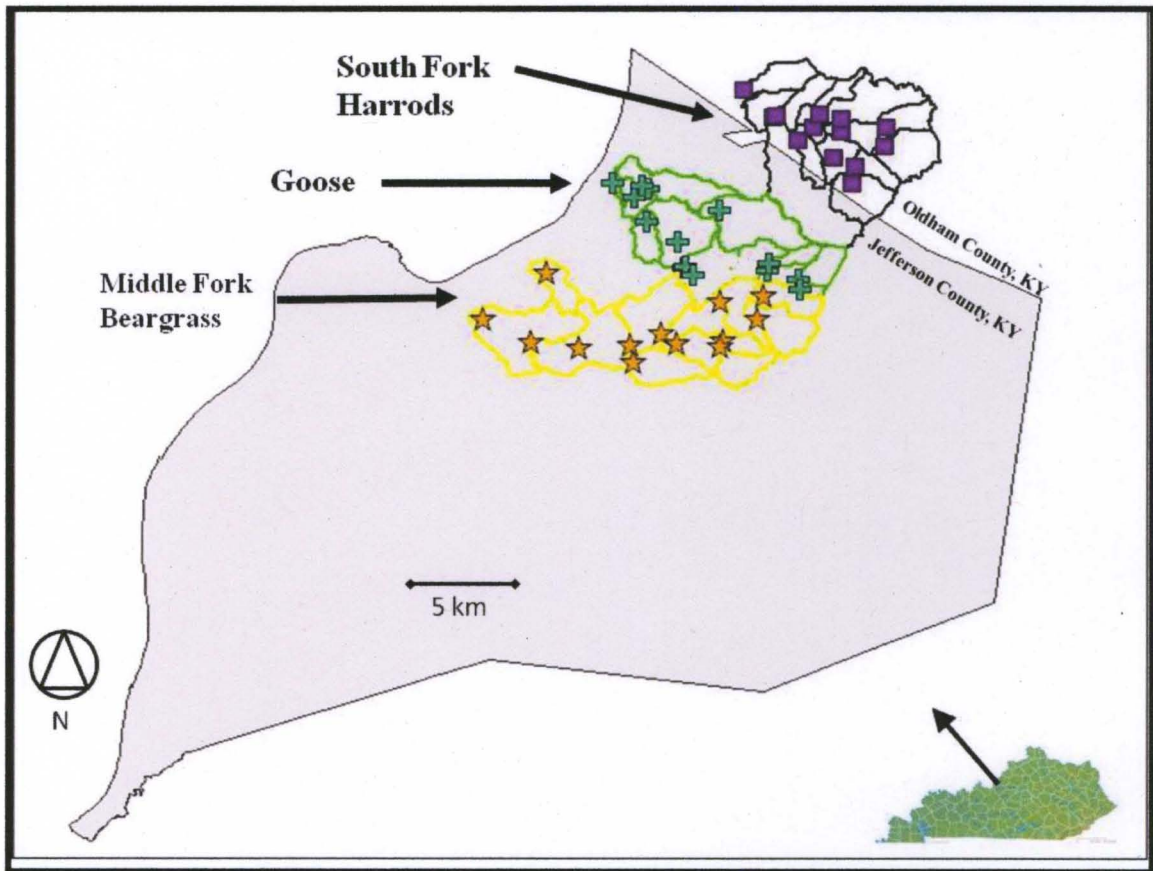
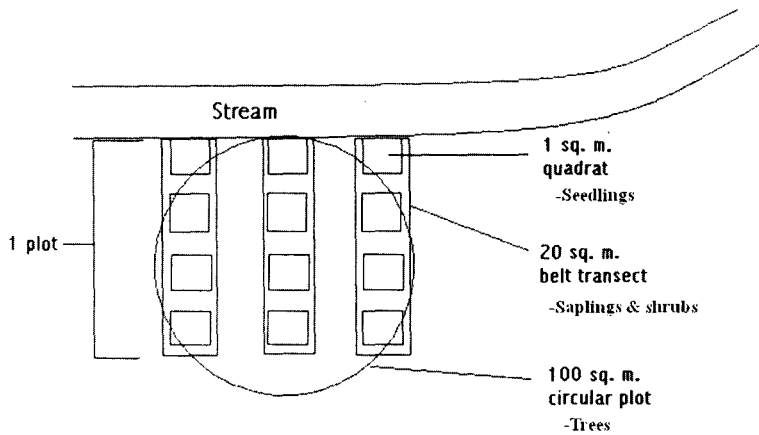
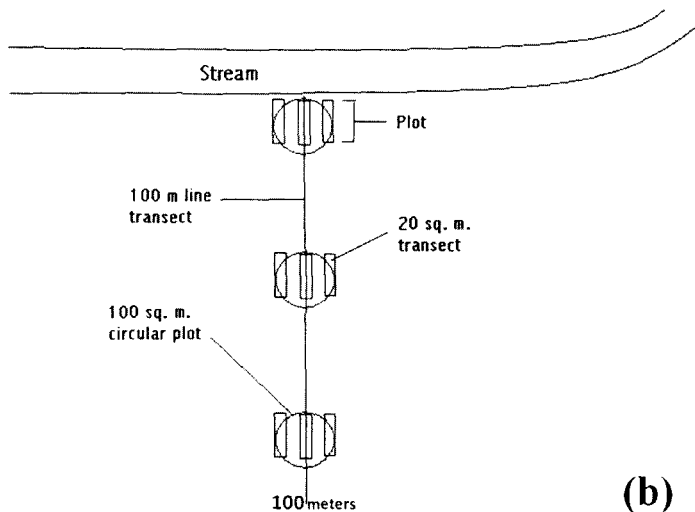


Figure 2-1: Schematic map of the three watersheds in Jefferson and Oldham counties in Kentucky. Research sites are indicated with markers.



(a)

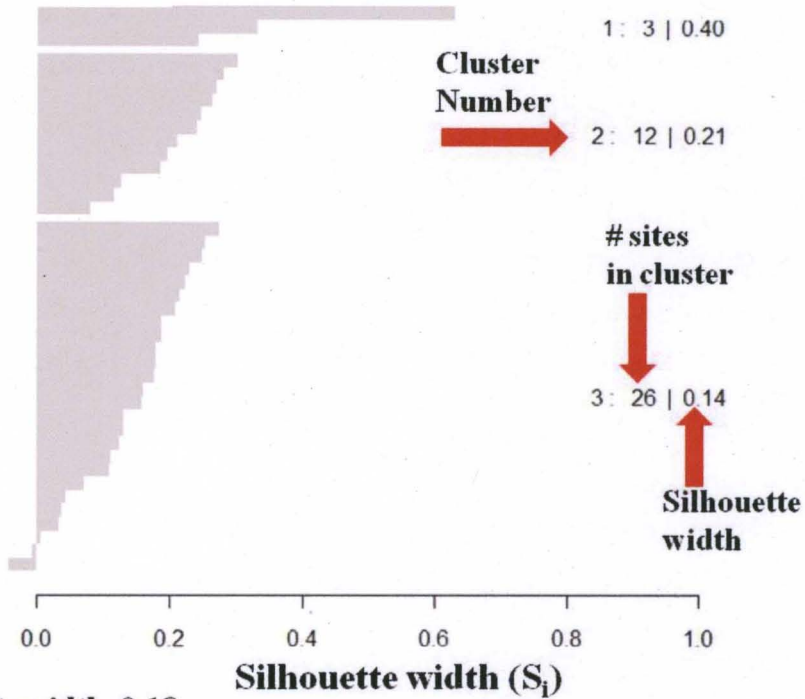


(b)

Figure 2-2: Plot and site design for sampling adult trees (≥ 2.54 -cm DBH), saplings, tree seedlings, and shrubs. **(a)** Each site consisted of at least one 100-m² circular plot for sampling tree counts and DBH, except in the cases when the riparian width could not accommodate a complete circular plot. Nested within the circular plot were three belt transects measuring 10 x 2 m with midlines spaced four meters apart for counting saplings and shrub stems. Quadrats measuring 1 x 1 m were nested within the belt transects for determining percent cover and counts of tree seedlings. Adjacent quadrat spacing within belt transects was three meters at the midpoint. **(b)** Where riparian width was sufficient, sites consisted of a midline transect stretched the length of the riparian zone up to 100 meters with up to 3 plots per transect with midpoints spaced 40-m apart.

Silhouette plot – 3 clusters

n = 41 sites



Average silhouette width: 0.18

Figure 2-3: Silhouette plot of a cluster analysis (optimal partitioning using Bray-Curtis distance) resulting in three groupings of research sites. Sites within clusters are represented by gray bars, the length of each demonstrating the silhouette width (value on the x-axis) of that particular site within the cluster. Cluster numbers, number of sites within the cluster, and cluster widths are to the right of each cluster. Overall silhouette widths are located below the numerical axis. Explanation of values given in Methods.

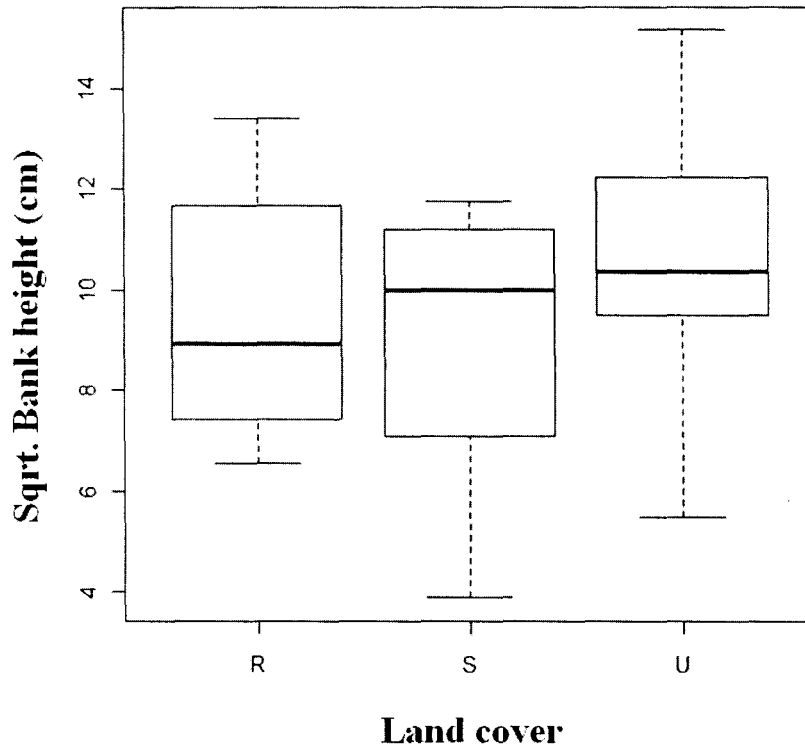


Figure 2-4: Bank height measurements for land-use categories determined at the subcatchment scale (R = Rural, n = 22; S = Suburban, n = 23; U = Urban, n = 22). The bold line indicates the median, boxes delineate 25th and 75th percentiles, and whiskers indicate minimum and maximum values. Mean urban bank height (122-cm) was higher than suburban bank height (86-cm; $p = 0.068$) and rural bank height (96-cm).

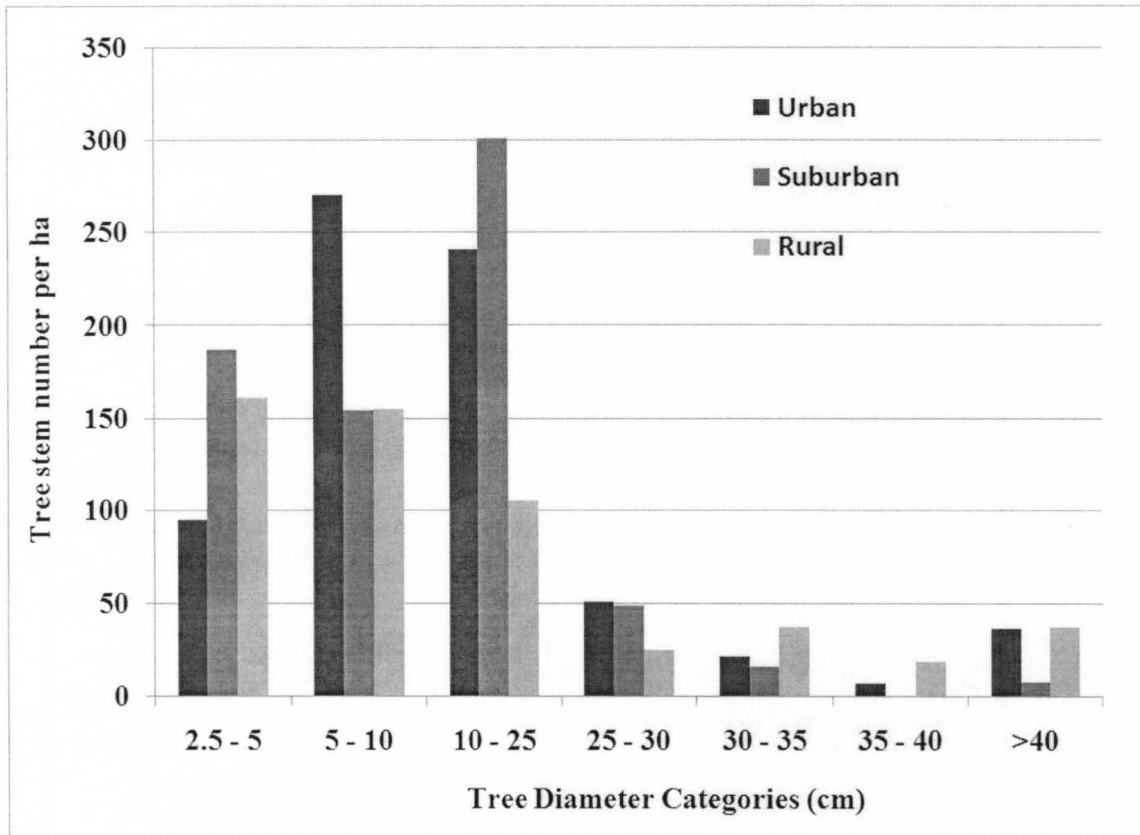


Figure 2-5: Tree diameter distributions by land-use category in riparian zones. Land-use categories were determined at the subcatchment scale.

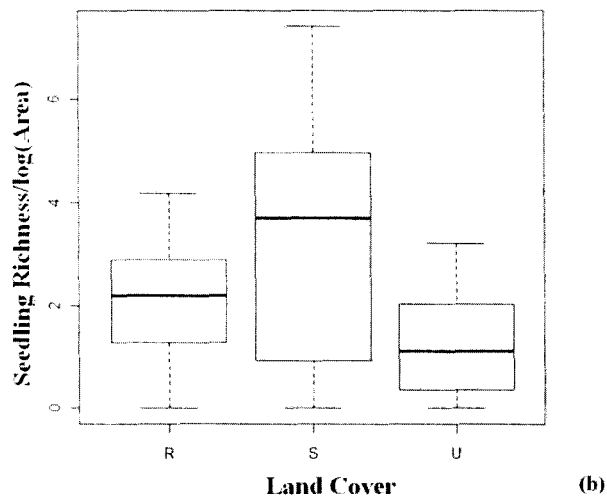
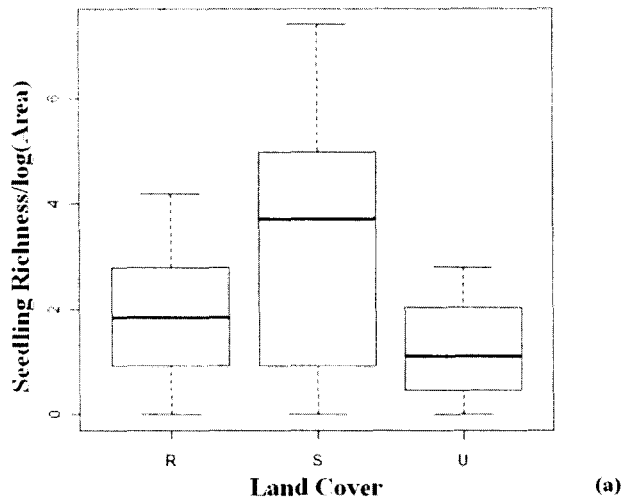


Figure 2-6: Tree seedling richness/ $\log(\text{Area sampled})$ for (a) near-stream plots only ($n = 41$ nearstream plots), and (b) sites ($n = 41$ sites). In (b), plots at the multi-plot sites were combined and treated as one sampling unit. Land use assignment for both plot and multi-plot sites was determined at the subcatchment scale (R = Rural, S = Suburban, U = Urban). The bold line indicates the median value, boxes delineate 25th and 75th percentiles, and whiskers indicate minimum and maximum values. Mean suburban tree seedling richness (Site and nearstream means = 2.8 species per site and nearstream plot) was significantly higher than mean urban tree seedling richness (site mean = 1.5 species; nearstream mean = 1.3 species) at $p = 0.05$ (nearstream plots only $p = 0.024$; all plots, $p = 0.028$).

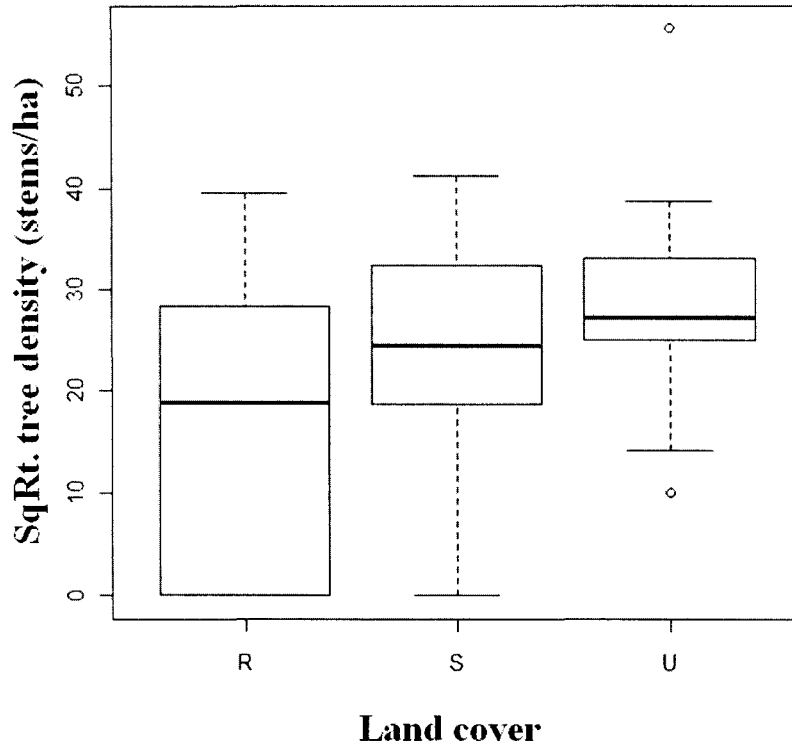


Figure 2-7: Square root of the tree density averaged across all nearstream plots in each land-use category. Land use categories were determined at the subcatchment scale (R = Rural, S = Suburban, U = Urban). The bold line indicates the median, boxes delineate 25th and 75th percentiles, and whiskers indicate minimum and maximum values. Circular markers represent potential outliers, but were not excluded from analysis. Mean rural tree density (477 stems/ha) was nearly half that of urban tree density (954 stems/ha; $p = 0.073$). Mean suburban tree density was intermediate (705 stems/ha).

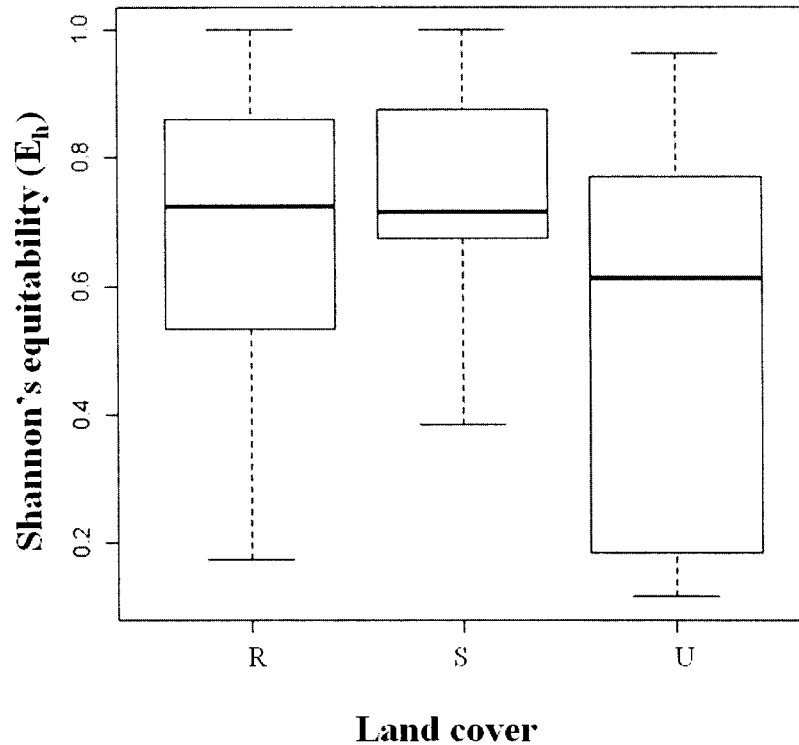
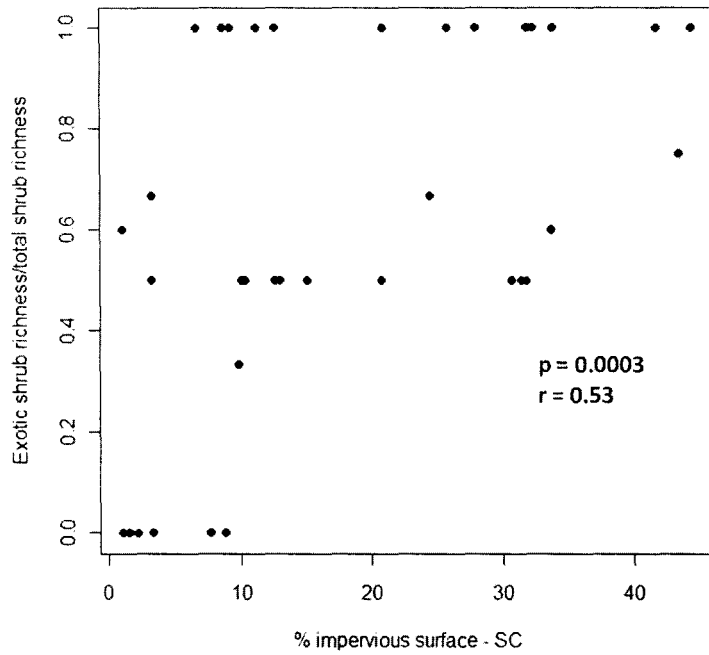
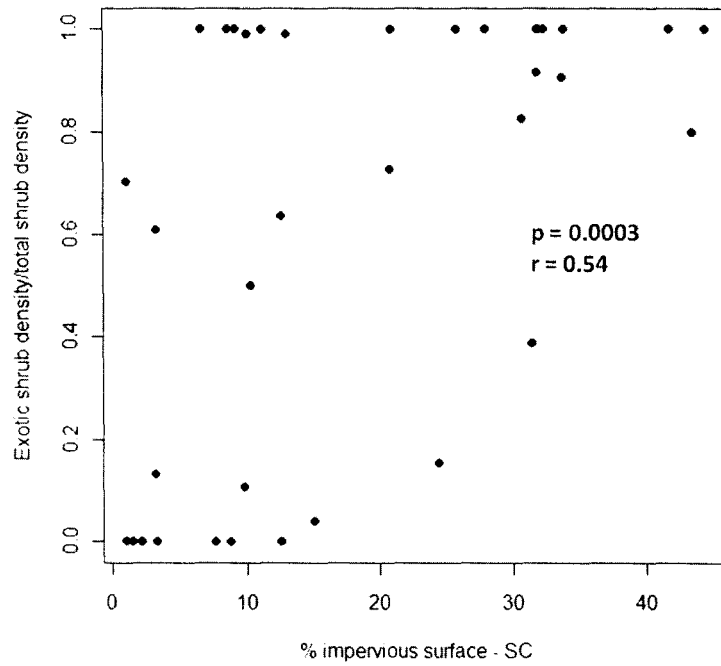


Figure 2-8: Shannon's equitability index (E_h) for all woody species using nearstream plots only for comparisons. Land-use categories were determined at the subcatchment scale (R = Rural, S = Suburban, U = Urban). The bold line indicates the median, boxes delineate 25th and 75th percentiles, and whiskers indicate minimum and maximum values. Mean urban woody plant evenness (0.54) was significantly lower ($p = 0.037$) than suburban community evenness (0.77), while mean rural woody plant evenness (0.68) was similar to that of suburban, but more variable.



(a)



(b)

Figure 2-9a-b: Exotic shrub species richness (a) and density (b) proportions with increasing impervious surface (subcatchment scale).

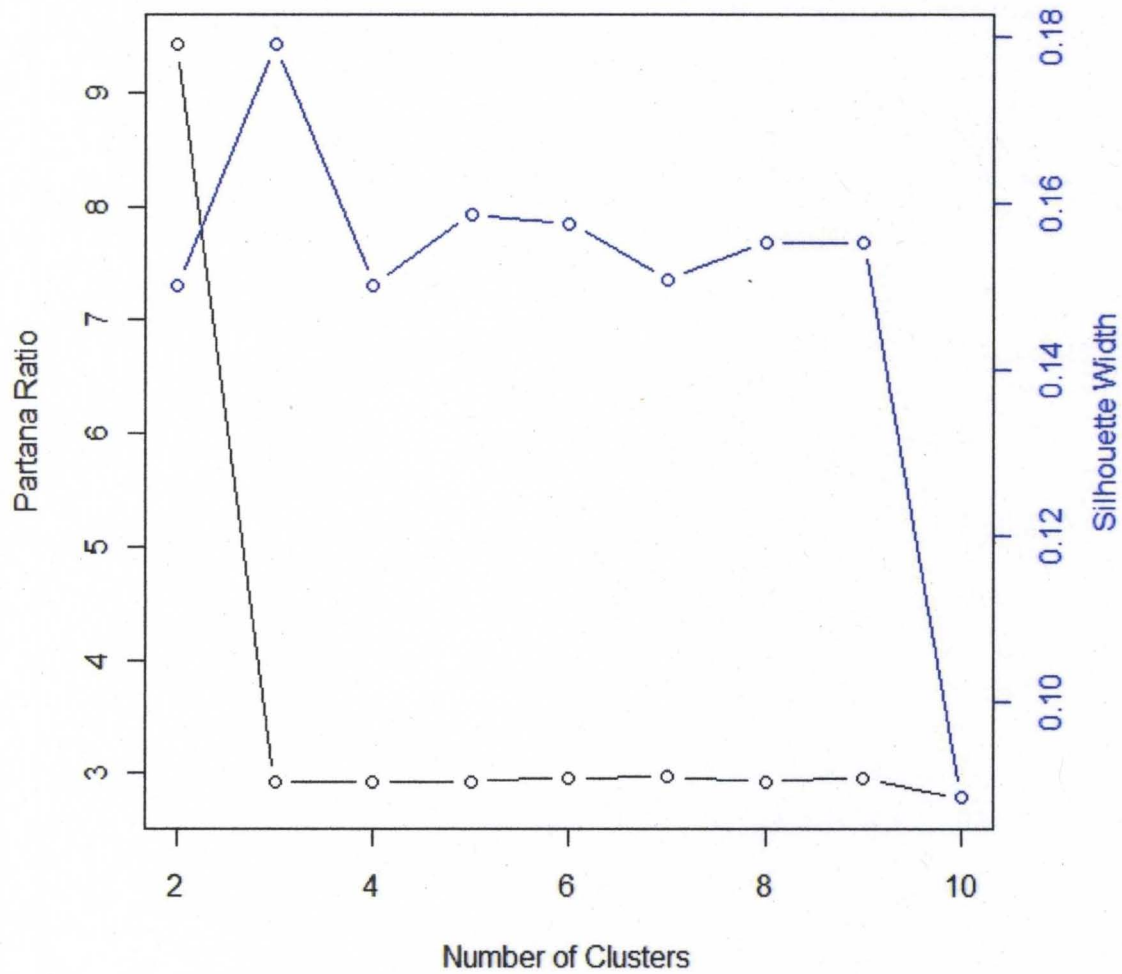


Figure 2-10: Partana ratios and overall silhouette widths for 2 to 10 clusters using optimal partitioning. The 2-cluster configuration, while giving the highest silhouette width and partana ratio, results in two clusters, with one cluster containing all but one of the sites, and the other containing one site. I chose to group the sites into three and five clusters since those two options gave the highest values for overall silhouette width (0.18 and 0.16, respectively). Explanations of silhouette width given in Methods.

Silhouette plot – 5 clusters

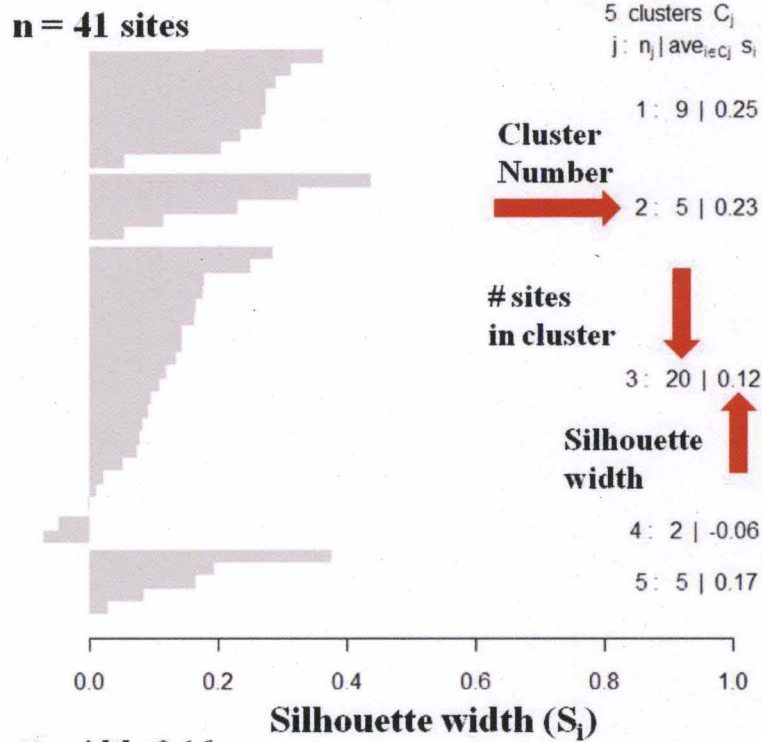


Figure 2-11: Silhouette plot of cluster analysis using optimal partitioning and Bray-Curtis distance resulting in five groupings of research sites. Sites within clusters are represented by gray bars, the length of each demonstrating the silhouette width (x-axis) of that particular site within the cluster. Cluster numbers, number of sites within the cluster, and mean cluster widths are to the right of each cluster. Average silhouette width, located below the numerical axis, is the mean of all silhouette widths for each site. Explanation of values given in Methods under the heading ‘Data Analysis’.

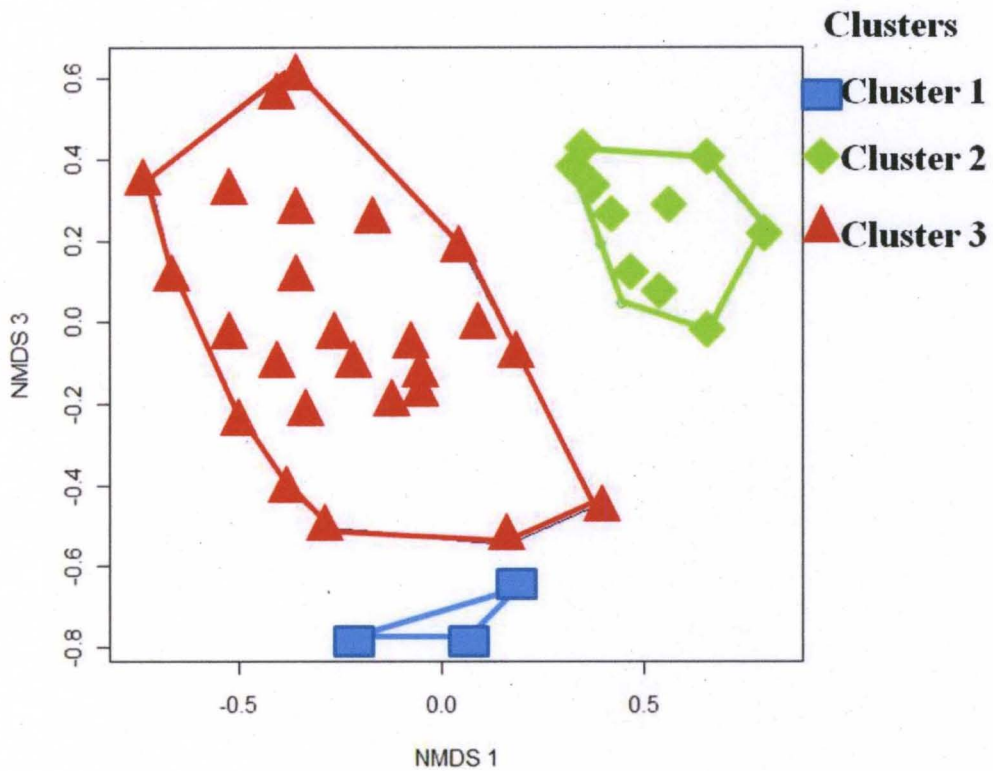


Figure 2-12: 3-dimensional configuration (only 2 dimensions shown) of woody plant data using non-metric multidimensional scaling (NMS) with sites (points) assigned to their respective groupings from the 3-cluster configuration from cluster analysis. Grouped clusters are shown along axes 1 & 3. No site overlap occurred in these three groupings in 3-dimensions indicating satisfactory agreement between the cluster analysis and the NMS solution. Plots in Cluster-1 (blue) are associated with the native shrub, *Symphoricarpos orbiculata* as its indicator species (Table 2-11), Cluster-2 in green is associated with the native shrub, *Lindera benzoin*, and the largest Cluster-3 (red) is associated with the exotic shrub, *Lonicera maackii*.

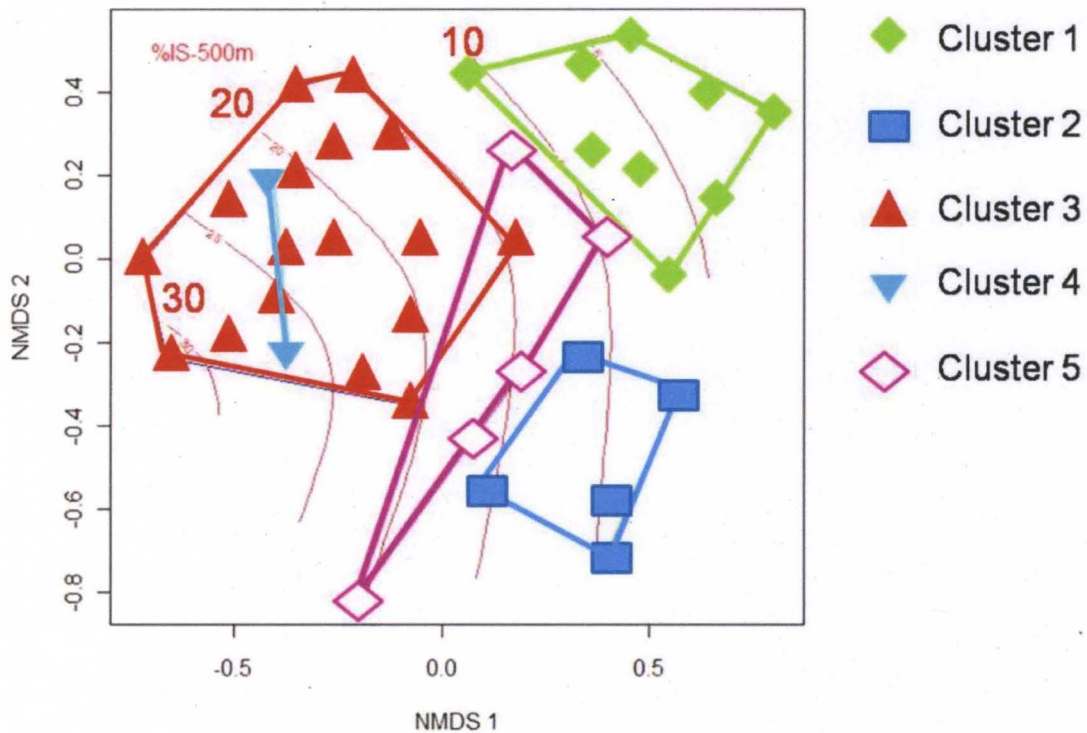
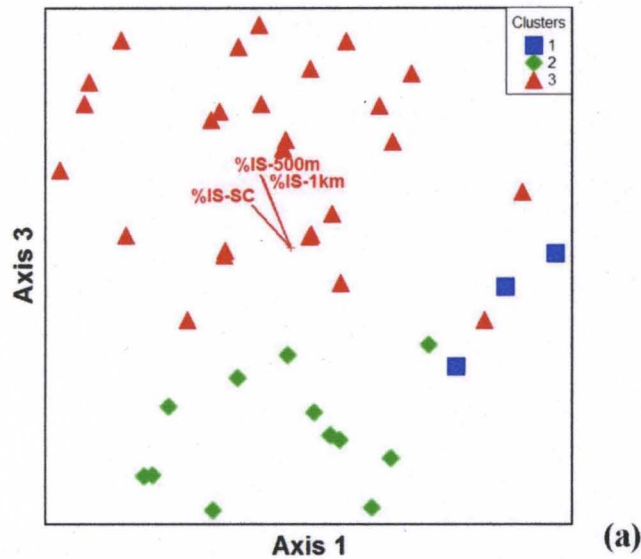
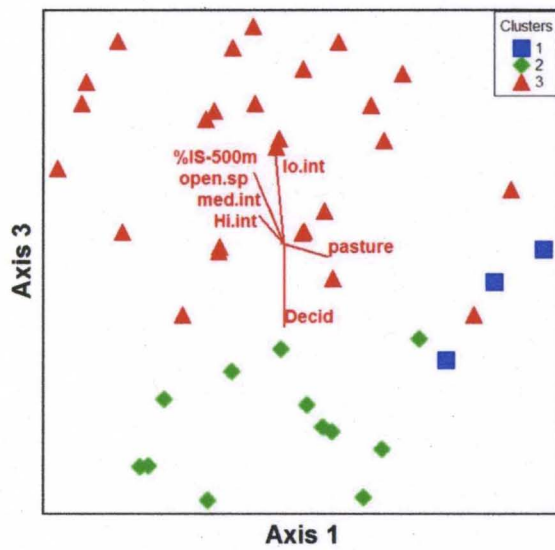


Figure 2-13: 3-dimensional configuration (only 2 dimensions shown) of woody plant data using non-metric multidimensional scaling coupled with cluster assignments from the 5-cluster configuration from cluster analysis. Isobar lines representing proportion of impervious surface within a 500-m radius of the research sites are presented to show the relationship to the clusters. Cluster-1 (green) is associated with the native shrub, *Lindera benzoin*, as a primary indicator species (Table 2-12), Cluster 2 (dark blue) with the native tree, *Acer negundo*, as its primary indicator species, Cluster 3 (red) with the exotic shrub, *Lonicera maackii*, as its primary indicator species, Cluster 4 (light blue) with anomalous outlier plots, and Cluster-5 (purple) with *Acer negundo* seedlings as its primary indicator species.



(a)



(b)

Figure 2-14a-b: Non-metric multidimensional scaling of woody plant data coupled with cluster assignments from the 3-cluster configuration using cluster analysis, shown as colored shapes. Clusters-1 and -2 contain native shrubs as indicator species, while Cluster-3 contains an exotic shrub as an indicator species. Vectors representing site environmental metrics are pointing in the direction of increasing proportions, with the length of each vector representing the strength of correlation. Vectors for environmental metrics are shown only if they have an $r^2 \geq 0.15$ along at least one axis. Environmental metrics shown along (a) axes 1 & 3 are the proportion of impervious surface (IS) surrounding each site at 3 scales: subcatchment (SC), 1-km and 500-m radius. Relationships of clusters with other environmental metrics are shown along the same axes in (b) and include low- medium- (med.int), and high-intensity (Hi.int) development, open-space developed (open.sp), % IS within a 500-m site radius (%IS-500m), pasture, and deciduous forest (Decid). Definitions are included in Appendix 3. Compare with Table 2-11 to get dominant and indicator species of the clusters. Explained variance (r^2) along each axis is as follows: Axis 1 - 0.14, Axis 2 (not shown) - 0.20, Axis 3 - 0.34.

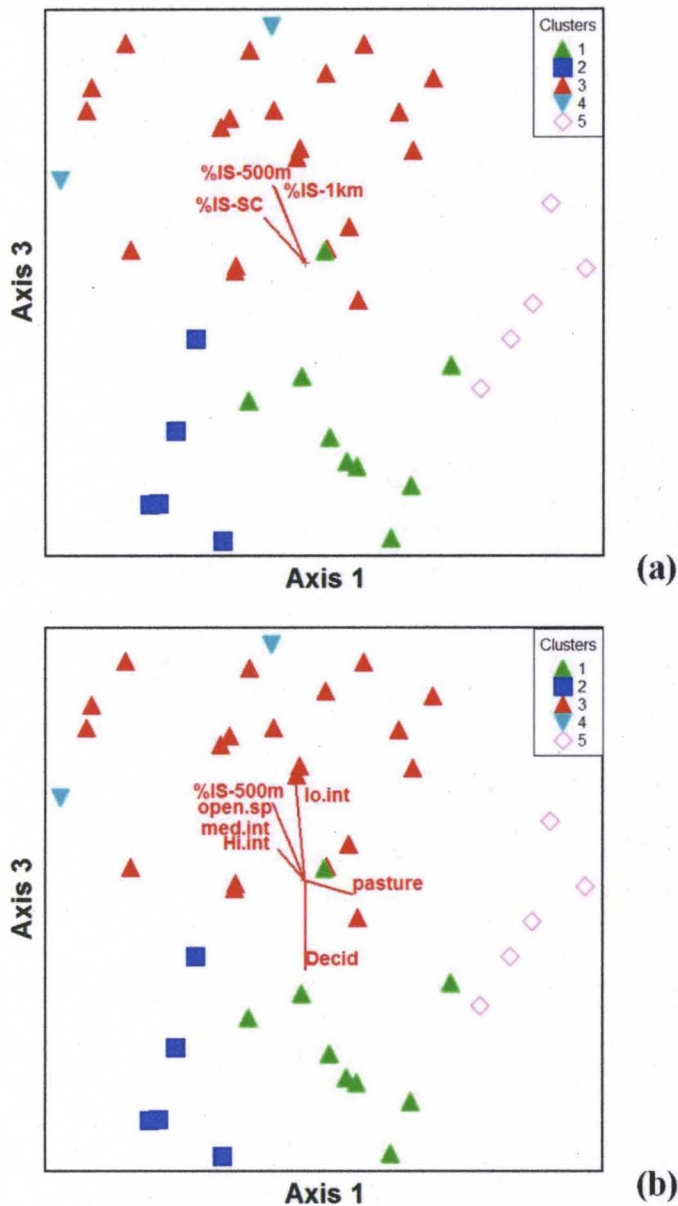


Figure 2-15a-b: Non-metric multidimensional scaling of woody plant data coupled with cluster assignments from the 5-cluster configuration using cluster analysis. Vectors representing site environmental metrics are pointing in the direction of increasing proportions, with the length of each vector representing the strength of correlation. Vectors for environmental metrics are shown only if they have an $r^2 \geq 0.15$ along at least one axis. Environmental metrics shown along (a) axes 1 & 3 are proportion impervious surface (IS) surrounding each site at 3 scales: subcatchment (SC), 1-km and 500-m radius. Environmental metrics shown along the same axes in (b) include low- (lo.int), medium- (med.int), and high-intensity (Hi.int) developed, open-space developed (open.sp), % IS within a 500-m site radius (%IS-500m), pasture, and deciduous forest (Decid). Definitions are included in Table 13. Compare with Table 2-12 to get dominant and indicator species of the clusters. Explained variance (r^2) along each axis is as follows: Axis 1 - 0.14, Axis 2 (not shown) - 0.20, Axis 3 - 0.34.

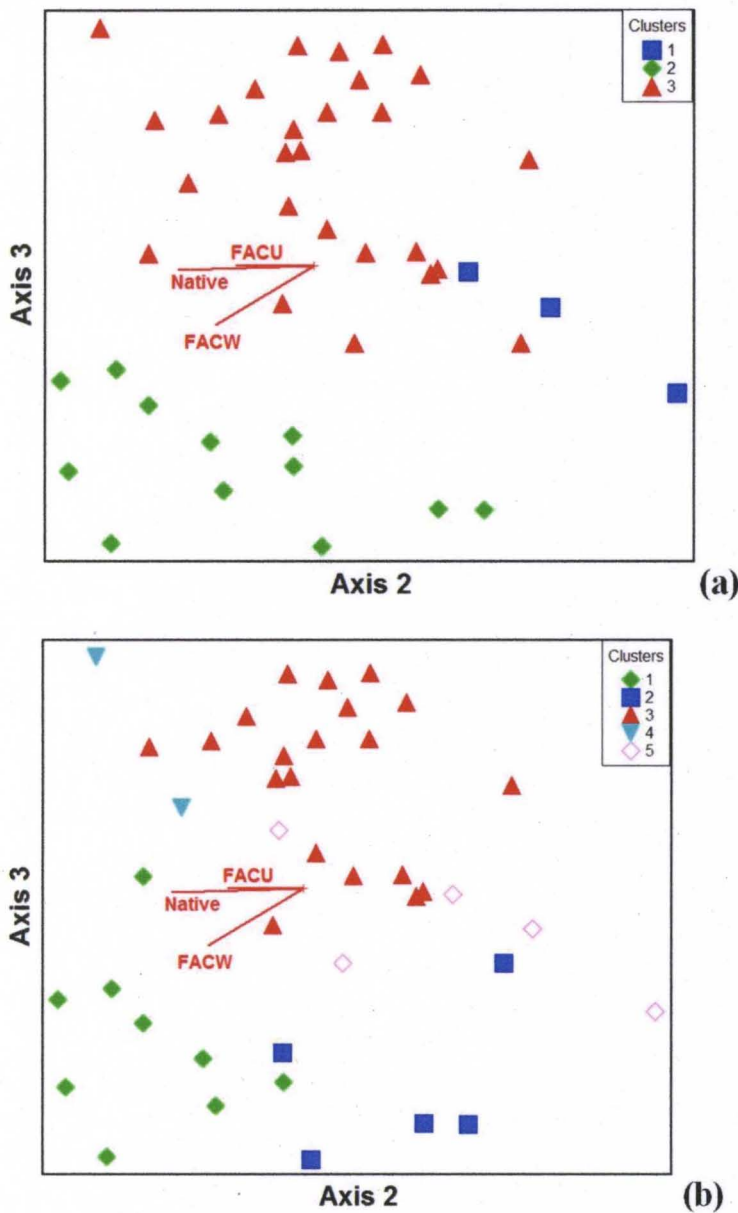


Figure 2-16a-b: Non-metric multidimensional scaling of research sites arranged according to woody species composition. Sites are color coded according to their cluster assignments from 3- and 5-cluster configurations using cluster analysis. Vectors representing species attributes are pointing in the direction of increasing importance values, with the length of each vector representing the strength of correlation. Vectors for species attributes are shown only if they have an $r^2 \geq 0.15$ along at least one axis. The 3-cluster assignments are pictured in (a) and (b). The 5-cluster assignments are pictured in (c) and (d), with identical site configurations shown in (a) and (c), and (b) and (d). The species attributes shown are facultative-upland (FACU), facultative wetland (FACW), and native species (N). Definitions and the full list of attributes included in the analysis are located in Table 14. Compare with Tables 8 and 9 to get dominant and indicator species of the clusters. Explained variance (r^2) along each axis is as follows: Axis 1 (not shown) - 0.14, Axis 2 - 0.20, Axis 3 - 0.34.

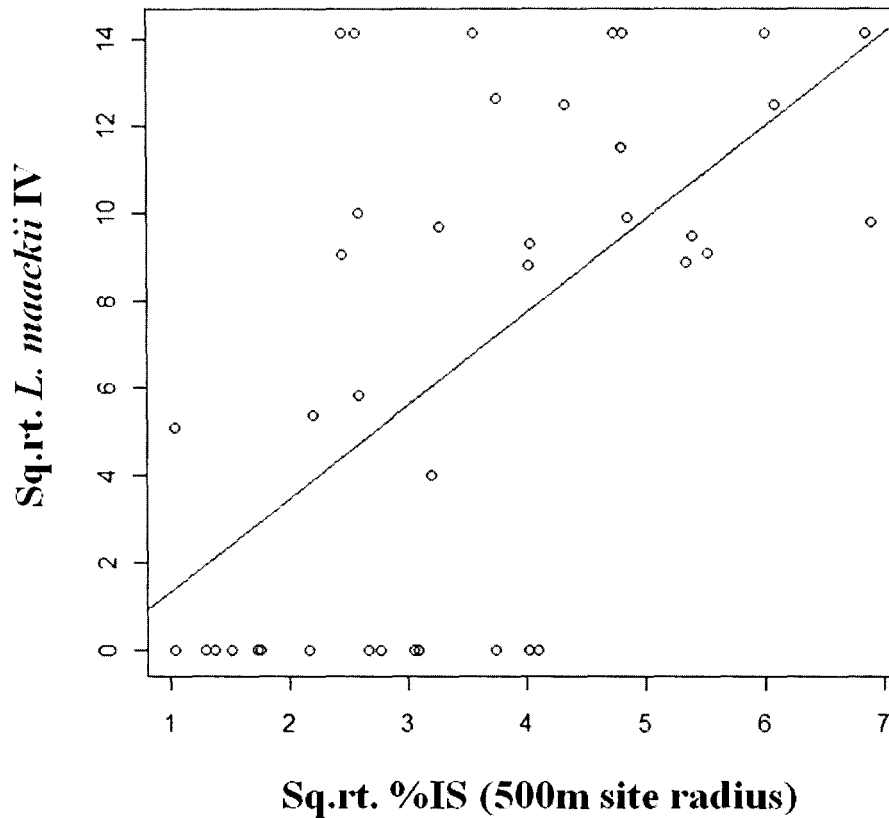


Figure 2-17: Linear regression of *L. maackii* relative importance value (IV) against proportion of impervious surface within a radius of 500-m of site centers. *L. maackii* IV significantly increased in relation to increasing proportions of impervious surface ($p = 0.000057$; adj. $r^2 = 0.34$). Data were square root transformed prior to regression. Regression of *L. maackii* densities against %IS within a 500-m site radius resulted in a lower coefficient of determination and p-value ($p = 0.0002$, adj. $r^2 = 0.28$) so the data are not shown.

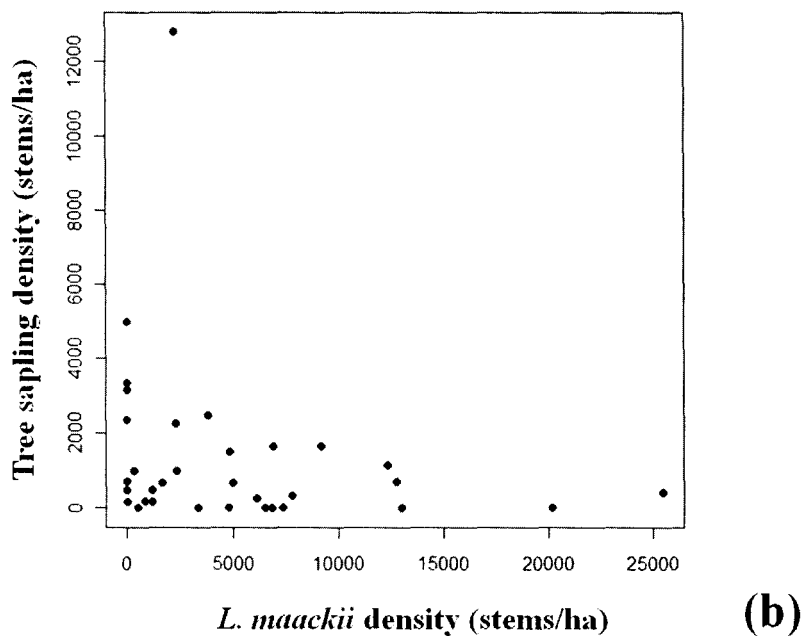
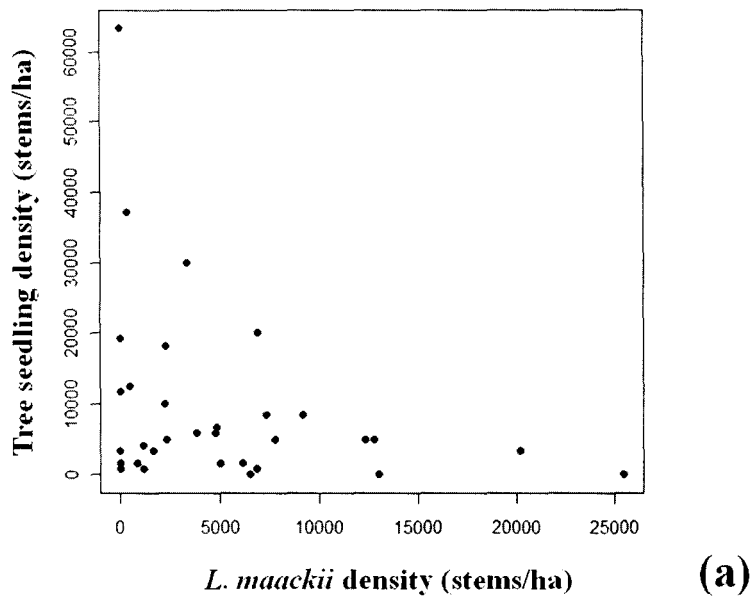


Figure 2-18a-b: Regressions of tree seedling and sapling densities compared to *Lonicera maackii* density at each site. Tree seedling density (a) and, to a lesser extent, tree sapling density (b) decreased in relation to increasing densities of *L. maackii*. The highest seedling and sapling densities occur at the lowest densities (<5000 to 10000 stems/ha) of *L. maackii*, and the lowest seedling and sapling densities at *L. maackii* concentrations >10000 stems/ha.

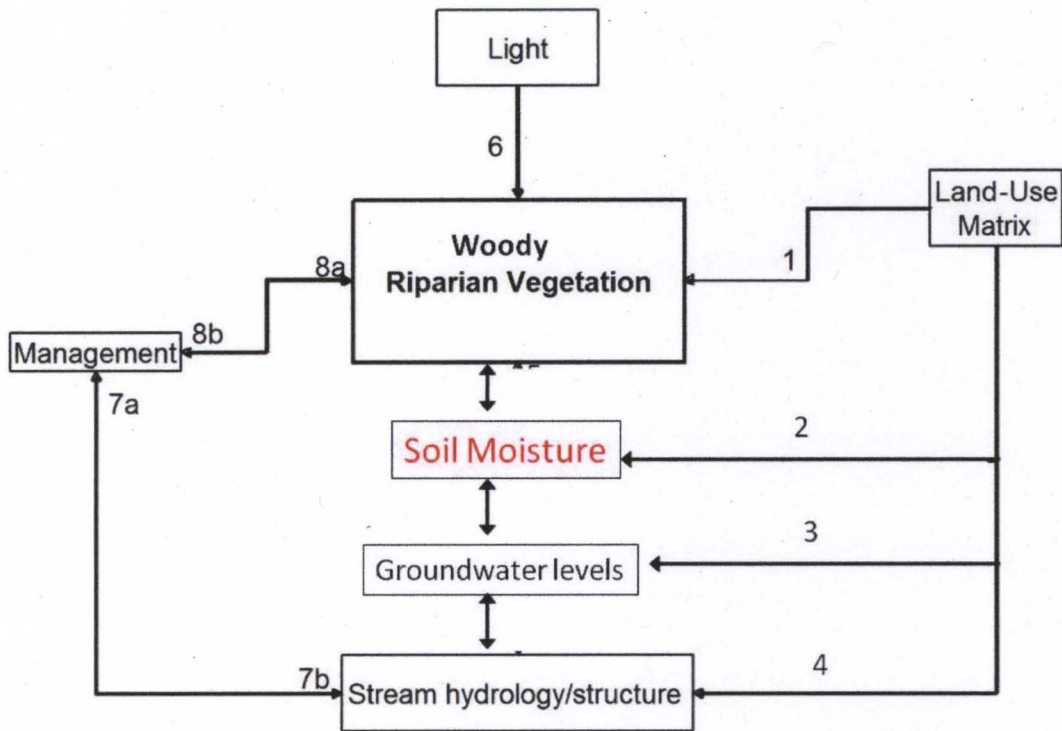


Figure 2-19: New conceptual model of variables that affect and are affected by vegetation within a riparian area modified from Figure 1-1. A direct link between groundwater levels and riparian vegetation was not found. However, since wetland species were found to a greater extent in rural riparian zones (Table 2-7, Fig. 2-16) soil moisture may be influencing plant species composition, although this would need to be verified in the field.

CHAPTER 3: VARIATION IN RIPARIAN GROUNDCOVER PLANT COMMUNITIES SURROUNDED BY URBAN, SUBURBAN, AND RURAL LAND- USE IN LOUISVILLE, KENTUCKY

Introduction

Urbanization can affect the composition of riparian herbaceous and groundcover species at various scales. The existence of a greater number of exotics in the urban species pool creates the potential for their dominating riparian areas at landscape and regional scales. However, the plant communities realized in any location are highly influenced by site-level factors such as micro-scale soil nutrient, light and moisture differences (Guillaume 2002). As such, urban influences, such as nutrient deposition, on these species are likely to occur within a smaller site radius than for woody vegetation, which is more greatly influenced by macro-scale influences, such as forest management (Guillaume 2002). Localized urban effects, such as edge creation through reduction in size of woody riparian buffers, will likely influence groundcover (herb and vine) species composition and vulnerability to exotic species invasion. For example, Canadian riparian zones had greater numbers of exotic annuals at the urban end of an urban-to-rural gradient, while more perennial natives occurred primarily at the rural end (Moffatt and McLachlan 2004).

The interaction of urbanization and variable physical properties of the riparian zone can also result in different attributes of the plant community. Plant diversity has been shown to vary with location within riparian zones. Changing conditions of moisture

and light availability with increasing lateral movement away from and longitudinal movement downstream have both been shown to relate to predictable changes in riparian plant diversity (Nilsson et al. 1994, Lite et al. 2005). For example, species richness of riparian herbaceous communities in arid environments in Arizona was found to decrease laterally away from the stream channel during a dry year (Lite et al. 2005). This finding was attributed to lower water availability farther from the stream, as the water table was found to drop with lateral distance from the channel. Furthermore, the opposite was true following monsoon rains and flooding, as herbaceous species richness increased with distance from the stream channel following increases in the water table. How plant species diversity changes with lateral distance from streams flowing through varying land-uses is less well understood.

In addition, riparian plant diversity has been shown to vary with catchment area and stream order. Nilsson et al. (1994) showed that plant species richness along a river in Sweden was higher along the main channel than its tributaries. One possible explanation offered was variation in mean annual stream discharge and peat and silt cover along streambanks. Lite et al. (2005) found a pattern of increasing mean species richness in an upstream direction following summer monsoon and flooding events in an arid environment, as herbaceous annual species rapidly responded to removal of resource limitation upstream. Whether such patterns exist for riparian herbaceous communities in response to changing resource availabilities across an urban-to-rural land-use gradient has yet to be determined.

The overarching goal of this project was to determine how groundcover plant communities (herbs and woody vines growing over the soil) along streams vary with

degree of urban and suburban development in three watersheds in Jefferson and Oldham counties in Kentucky. The first objective was to determine whether potential relationships existed between riparian groundcover vegetation assemblages and impervious surface cover and NLCD land-cover types at different spatial scales. Specific predictions associated with this objective were that distinct groundcover vegetation communities exist that can be partially correlated with impervious surface cover, used as an index of urbanization. The second prediction was that the sphere of influence of different land covers would correlate more strongly with differences in groundcover vegetation with decreasing site radius, given the sensitivity of herbaceous vegetation to microscale as opposed to macroscale environmental variation (Guillaume 2002).

The second objective was to determine whether differences in plant species diversity existed along this urban-to-rural land-use gradient as well as with increasing catchment area and lateral distance from the streambank. There are three predictions associated with this objective, the first being that groundcover plant diversity along this gradient would be higher in suburban riparian areas than in either urban or rural riparian areas for two reasons: 1) because of the likely greater availability of diverse seed from residential areas than in rural locations (McKinney 2008), and 2) because of the prevalence in urban riparian areas of *Lonicera maackii*, an invasive shrub known to reduce herb layer diversity (Collier et al. 2002, Loewenstein and Loewenstein 2005) (see Chapter 2). The second prediction was that species richness would increase with total upstream catchment area, since other studies have shown that plant species richness increases with downstream distance (Bendix 1997, Bendix and Hupp 2000, Lite et al. 2005). The third prediction was that groundcover plant species richness would increase

with lateral distance from the stream due to the decreasing influence of floodwaters farther from the stream (Lite et al. 2005). To my knowledge, this is the first study to have compared herb and woody vine species richness in the ground layer of riparian environments across different land-use types and with lateral distance from the stream.

The third objective was to determine the extent to which native and non-native groundcover vegetation was associated with varying land-cover types. Since one consequence of urbanization is creation of edges, I predicted that native vegetation would be more dominant in riparian assemblages associated with greater proportions of forested land-cover types, and exotic vegetation would take on greater importance in riparian assemblages associated with urban land-cover types. The fourth objective was to determine the extent to which wetland and upland species were associated with various land-cover types. Because of impervious runoff, stream incision due to greater impervious cover, and the urban heat island effect, I proposed that higher streambanks would be found in urban areas and that this trend would be correlated with groundcover species composition. I specifically predicted that obligate and facultative wetland species would assume greater dominance in riparian assemblages associated with forested rural land-cover types and that facultative and upland species would exhibit greater dominance in riparian assemblages associated with urban land-cover types. The fifth objective was to determine groundcover species associations with and diversity within plots dominated by the shrub, *Lonicera maackii*. Since *L. maackii* is an exotic species with allelopathic properties and strong shading capabilities, I predicted that herb layer species composition would differ greatly and that species richness and other measures of diversity would be lower under this shrub than where this shrub was less dominant.

Methods

Study Area

The study area consisted of three individual watersheds located within and adjacent to Louisville, Kentucky, USA (Latitude 38°15'N, Longitude 85°46'W), which has a population of approximately 700,000 with a mean density of 695 people km⁻² (U.S. Census Bureau 2008). These watersheds, Beargrass Creek (Middle Fork), Goose Creek, and Harrods Creek (South Fork), lie adjacent to each other with all streams flowing in a primarily westerly direction and eventually emptying into the Ohio River (Fig. 3-1).

The three watersheds are of approximately equivalent sizes, containing varying proportions of impervious surface cover (ISC; Table 3-1). Beargrass Creek watershed, located in the north-northeast portion of Jefferson County, KY, is approximately 65 km² (Metropolitan Sewer District 1999b) and contains the greatest proportion of impervious surface cover (33% ISC). This Middle Fork of Beargrass Creek joins the South Fork before flowing into the Ohio River. Several stretches of this stream are featured on Kentucky's Division of Water 303d list, which list streams with impairment issues as well as the pollutant(s) causing the impairment (Kentucky Environmental and Public Protection Cabinet 2008). Since combined sewer overflows (CSOs) commonly occur in this stream, the presence of fecal coliform and sewage are stated as the primary problem pollutants within Beargrass Creek.

Goose Creek is a 50-km² watershed also in Jefferson County, in-between and adjacent to Beargrass and Harrods Creek watersheds (Metropolitan Sewer District 1999a). Of the three watersheds in this study, impervious surface cover is intermediate (20% ISC). The creek is composed of a pair of forks that meet before emptying into the

Ohio River. Goose Creek is also listed on the 303d list, with cadmium and fecal coliform levels stated as impairment issues.

South Fork Harrods Creek watershed (60-km²), which has the lowest impervious surface cover of the three (10% ISC), is located primarily in neighboring Oldham County, but extends into northeast Jefferson County. South Fork Harrods Creek eventually joins North Fork Harrods Creek to become Harrods Creek before flowing into the Ohio River. Sections of the larger Harrods Creek network made the 303d list in years past, specifically the portion passing through Jefferson County which is a more developed area. As in the other two watersheds, fecal coliform has traditionally been a problem (Metropolitan Sewer District 1999c). The current 303d list contains Harrods Creek with fecal coliform as an impairment issue, although this information is for the stream section downstream of where the North Fork and the South Fork Harrods Creek join.

Watershed classification

Watersheds were chosen from digitized catchment layers downloaded from the Louisville/Jefferson Information Consortium (LOJIC) database (LOJIC 2010). LOJIC is a multi-agency consortium that maintains a geographic information systems (GIS) database to serve primarily Jefferson County, KY, although limited information is available for a portion of Oldham County, KY. After checking for topographical accuracy, I divided catchments into subcatchments based on stream order and topography from the original LOJIC layers. I further divided subcatchments of higher stream orders with longer reaches into two or three subcatchments. Data layers from the LOJIC

database were clipped to each of these subcatchments. Beargrass Creek watershed contained thirteen subcatchments, Goose Creek watershed contained twelve subcatchments, and Harrods Creek watershed contained fourteen subcatchments.

Impervious land-cover classification was determined using three data layers from the LOJIC database digitized in 2001. These layers included buildings, miscellaneous transportation, and roads. The buildings layer included all built structures, whether commercial or residential. The roads layer included all paved road surfaces, excluding parking lots. The miscellaneous transportation layer included secondary transportation features such as driveways, parking lots, and sidewalks. The area of land covered by impervious surfaces for each subcatchment was determined using ArcGIS 9 (ESRI 2010) and was divided by the total land area for each subcatchment to determine the proportion of impervious surfaces covering the landscape.

Categorization of subcatchments into urban, suburban, and rural land-use categories was modified from Schuler (1994). For this study, urban subcatchments were defined as containing $\geq 30\%$ impervious surface cover. Rural subcatchments contained $\leq 10\%$ impervious surface cover and suburban subcatchments contained between 10% and 30% impervious surface cover. To explore the degree of association between riparian plant communities and proximity of different types of land use and cover, research sites were classified by land use based on impervious surface cover at three scales. The first and largest areal scale was the subcatchment level, where sites were classified as urban, suburban, or rural based on the proportion of impervious cover in the subcatchment in which they were located. Impervious surface cover was calculated at two additional and smaller scales surrounding the sites. These site buffers at a radius of 1-km and 500-m

from the site center were created using ArcGIS 9. The proportion of impervious surface cover was determined for these buffers in the same way as for the subcatchments. Sites were then classified into one of the three land-use categories in the same way as the subcatchment. Urban, suburban, rural categorization of sites was changed for each relevant analysis depending on the scale at which impervious cover was calculated.

Site selection and plot establishment

Forty-one riparian sites were chosen from within the aforementioned 39 catchments. Riparian zones were defined by topography and structural confinements. Areas of level ground between the streambank and either an upland slope, structural barrier (roads, parking lots, buildings), or other obstruction (grazed pasture, ephemeral stream channel) were considered as being within the riparian zone. Sites were chosen randomly using ArcGIS 9 as follows. A vertical measurement line with units in meters was drawn the length of the stream section within the subcatchment. A random number multiplied by 100 meters was used to find the location of the site along the vertical measurement line. Sites were visited and chosen if they were at least partially covered with unmanaged vegetation and at least 100 meters from the nearest fork to avoid sampling dual riparian zones from separate channels. Riparian zones located completely within regularly used pastures and those found to be mown to the edge of the streambank were not used for this study. Only one side of the stream was sampled at each site. If both sides contained vegetation, the larger side was chosen if it fit the criteria above, if it was reachable, and if the proper permissions could be attained. If only one side of the stream contained vegetation then that side was chosen. Locations where at least one full plot

could not be placed (riparian width < 12-m) were sampled with a partial plot due to the time and expense of acquiring permission to access sites, and to avoid potential biases of purposefully choosing a site of a particular width. The distribution of riparian widths by stream order and land-use category for the sites chosen is presented in Table 3-2.

After visiting the locations identified by ArcGIS selection, the location of the actual study site was chosen by taking a randomly determined number of steps to establish the center of a transect. I used a compass to extend a transect tape perpendicular to and away from the stream channel up to 91 meters, if possible. Circular plot centers (plot radius = 5.64-m) were established beginning at 6, 46, and 86 meters away from and perpendicular to the stream channel. Three belt transects measuring 10 x 2 meters were nested within the circular plots, with the long edge perpendicular to the stream. The distance between midlines of adjacent belt transects was four meters. Twelve 1 x 1 meter quadrats were nested within the belt transects (four quadrats per belt transect). Quadrats were placed every two meters down the midline of each belt transect, so that the centers of adjacent quadrats within a belt transect were three meters apart (Fig. 3-2).

The circular plots were used in a previous study to estimate the abundance and species identity of trees. Belt transects were used to measure shrub live stem densities. Percent cover and identity of groundcover species (herbaceous and woody vine species), as well as number and percent cover of *L. maackii* seedlings, were determined in each 1 x 1 meter quadrat. For all vegetation data, only living plants were considered. *L. maackii* sampling was conducted in 2005 (June to August) and in 2006 (May to October). Groundcover plant data were collected from May to October, 2006.

Plant identification and classification

Plant identifications were made in the field whenever possible. The primary source used for identification of shrubs and vines was Wharton and Barbour (1973). Plants that could not be identified in the field were collected for later keying using Jones (2005). Collections included as many parts of the plant as possible, including stem, leaf, and flowering parts. However, the authority for all species names used here was the USDA PLANTS Database (2010). Species provenance at the continental level was determined using Jones (2005) and the USDA PLANTS Database (2010). Two categories of species provenance at the continental scale were used: native and exotic. Term usage is as defined in the USDA Natural Resources Conservation Service (2009). Native plants are those that have developed over hundreds of years in an area (in this instance, the continental U.S.) and were present prior to European settlement. Exotic plants are defined herein as those introduced with human assistance to a continent on which they were not previously found prior to European settlement.

Importance value and diversity

For each site (one to three plots), an importance value (IV) was calculated for each groundcover species as the sum of average percent cover and frequency. Average importance values were calculated across quadrats at each site based on the arithmetic mean of the percent cover of each species. Percent frequency was determined for each species based on the number of quadrats in which a particular species was present divided by the total number of quadrats at each plot and multiplied by 100.

Eq. 1

$$\text{Groundcover species IV (\%)} = \text{Average \% cover} + \% \text{ Frequency}$$

Species richness (S), Shannon's diversity (H') and Simpson's reciprocal (1/D) indices, as well as Shannon's equitability index (E_h) were computed for each site using data collected only from the plot nearest the stream (see Chapter 2 Methods for full equations and references). Since site and plot sizes varied, richness was calculated as $S/\text{Log}(\text{Area sampled})$ at the site scale (for up to three plots) and at the nearstream plot scale. Log transforms were used on richness data to normalize variance. Two sites with the lowest richness were eliminated from site and nearstream analyses to normalize residuals and stabilize variance. Data from only nearstream plots were compared using Shannon's, Simpson's, and equitability diversity analyses when examining potential relationships between plant community diversity and land cover and use. Average % cover for each species was computed for each nearstream plot and multiplied by 100 to obtain all integer values. Shannon's equitability index (E_h) was computed as a measure of species evenness as $E_h = H'/\ln(S)$, where H' is Shannon's diversity index and S is species richness.

Community diversity was expressed at each site using species richness, Shannon's Index and Simpson's Reciprocal Index. In addition to nearstream plot richness, site richness was calculated by totaling species across all plots for a site. Because of different riparian widths and plot sizes, richness was calculated for all sites as well as nearstream plots as the number of species divided by the log of the area sampled (Conner and McCoy 1979, Nilsson et al. 1997). Average percent cover of groundcover

species (herbaceous species and vines) was used to compute Simpson's reciprocal (1/D) (Simpson 1949, Magurran 2004) and Shannon's (H') (Shannon 1948, Magurran 2004) indexes for plots nearest the stream. EstimateS (Colwell 2009) was used for computing diversity indices. In EstimateS, diversity index estimators were not used, so indices were computed using the observed sample order (no randomization).

Metrics for wetland indicator status (WIS) and species provenance (NE; N=native, E=exotic) were computed for each site to assess correlations with various land-cover variables. Species were labeled with their WIS from the USDA PLANTS database as well as NE as describe previously. WIS and NE codes and meanings are shown in Appendix 2. A numerical value was assigned to each site for each category of WIS and NE using the groundcover IV's calculated for each species in each site. Values were calculated by averaging the IV's of the species in each WIS and NE category and adding a weighting factor to account for the number of species at each site within that particular category. The weighting factor was the number of species within a particular category divided by the total number of species within the site. Being based upon absolute IV's, the maximum value for any species indicator category was 300 (maximum 200 for average absolute IV + maximum 100 for the weighting factor). Values were converted to percent form by multiplying by 100.

Eq. 2

$$\text{Native metric} = (\text{Native Sp}_1 \text{ IV} + \text{Native Sp}_2 \text{ IV} + \text{Native Sp}_3 \text{ IV} + \dots + \text{Native Sp}_N \text{ IV})/N + \text{Weighting factor } [\# \text{ native spp.}/\text{Total \# spp.} * 100]$$

Eq. 3

$$\text{Exotic metric} = (\text{Exotic Sp}_1 \text{ IV} + \text{Exotic Sp}_2 \text{ IV} + \text{Exotic Sp}_3 \text{ IV} + \dots + \text{Exotic Sp}_N \text{ IV})/N + \text{Weighting factor } [\# \text{ exotic spp.}/\text{Total \# spp.} * 100]$$

The same form of equation was used to compute metrics for each category of WIS. These values were included in an environmental matrix and used to determine correlations with research sites arranged according to similar species composition in ordination space.

Distance From Stream and Catchment Area Analyses

Relationships were also explored between groundcover plant diversity and perpendicular distance from the stream by land-use type. Only nearstream plots were used, since all sites had at least this plot in common. At each nearstream plot, groundcover species richness was computed for each row ($n = 1$ to 4 rows per plot depending on plot size) of quadrats ($n = 3$ per row). Quadrats were arranged as previously described, with rows parallel to the stream spaced every two meters beginning one meter from the streambank edge. The four rows were positioned from 1 to 2, 4 to 5, 7 to 8, and 10 to 11 meters from the streambank. Richness values for the two rows nearest the stream were log transformed prior to analysis to meet assumptions of normality. Richness for the furthest two rows met assumptions of normality without data transformation. In some instances, mown lawns became the only vegetative cover beyond a certain lateral distance from the stream. In these cases the lawns were treated as monocultures and richness was assigned a value of 1.

Linear regression was used to compare groundcover richness for nearstream plots with cumulative upstream catchment area and bank height adjacent to the plot as a metric for depth to groundwater. Lawns were not included in determining richness values in these analyses, since lawns can be considered a type of land-use. Richness was then normalized to account for varying plot sizes as (richness/Log (area sampled)). Plots were also assigned to urban, suburban and rural land uses based on % impervious surface at the subcatchment scale. For each land-use category, groundcover richness was regressed as a response variable against cumulative catchment area (km²) upstream of and including the subcatchment in which the plot was located, as well as against bank height (cm) measured adjacent to the midline of each plot. Cumulative catchment area and groundcover richness were natural log transformed to better meet assumptions of the linear regression model.

Honeysuckle analysis

The relationship between *L. maackii* stem density and measures of groundcover diversity was assessed using linear regression. Again only nearstream plots were used in these analyses. In the case of exploring potential relationships between *L. maackii* density and percent cover of the vine *Euonymus fortunei*, two different approaches were used. In the first, plots were included in the analysis if they contained *E. fortunei*, but not necessarily *L. maackii* to demonstrate the abundance of *E. fortunei* in the presence and absence of *L. maackii*. Data were square root transformed prior to analysis, and one site was removed to normalize residuals and variance. In the second analysis, sites were included only if they contained both species occurring together in the same plot to

demonstrate the degree to which the abundances of the species were related to each other. Data were log transformed and two sites were removed to normalize residuals and variance.

To explore relationships between groundcover species diversity and *L. maackii* and *E. fortunei* abundances, I chose sites where those two species occurred and regressed values for site diversity indices (H' , $1/D$, E_h , S) against either *L. maackii* density or *E. fortunei* percent cover. In regressions involving *L. maackii*, stem densities of *L. maackii* and diversity index values (except H') were log transformed, and two sites were excluded prior to analysis to normalize residuals and variance. In regressions with *E. fortunei*, the two sites with the lowest and highest E_h were excluded from analysis to reduce heteroscedasticity. In addition, *E. fortunei* percent cover was square root transformed prior to analysis with $1/D$ and H' , and one site was removed prior to regressions against H' to normalize residuals and variance.

Environmental parameters

Several environmental variables within and external to the plots were used to explore their ability to explain the variation in plant species composition. More proximate site variables included bank height, canopy openness, and plot size. Factors that integrated influences from the greater landscape included % impervious surface cover (at three scales: the subcatchment, within 0.5-km radius and within a 1-km radius of site center), land-cover categories, stream order, cumulative catchment area, and property value.

Bank height was measured from the first terrace out of the channel and referenced to the depth at middle of the stream bed. Where the center of the channel was not reachable from the bank or by wading, bank height was measured at arm's reach. Bank height was measured once at the midline of each site ($n = 41$). These measurements were used as a proxy for depth to groundwater in order to determine correlations with species composition at the sites. Additional measurements were taken 50 meters to either side at thirteen of the sites where future studies of soil respiration were conducted. These thirteen additional measurements were averaged with the 41 site measurements to assess differences in overall bank height by land-use category.

Since canopy openness and hence light can also be a determinant of species presence and dominance, a spherical densiometer (Lemmon 1956, 1957), held over the middle of each of twelve 1-m^2 quadrats, was used to determine this measure. These values were averaged to obtain percent canopy openness for each site, as well as for each distance interval from the stream as described previously under the 'Distance Analysis' heading. Plot sizes were based on sampling areas of circular plots (Fig. 3-2) and ranged from a minimum of 28-m^2 to a maximum of 300-m^2 . Thirteen sites out of 41 had an area less than 100-m^2 , and seven sites had an area greater than 100-m^2 (one being 150-m^2 , five being 200-m^2 , and one site being 300-m^2). The remaining sites were 100-m^2 .

Impervious surface cover was determined as described previously and the three scales were used to explore which scale better predicted plant community composition. Finer-grained land-cover categorizations (developed -open space, -low, -medium, and -high intensity, deciduous forest, mixed forest, herbaceous, pasture, cultivated crops, and wetlands) were made using the U.S. Geological Survey's National Land Cover Database

from 2001 (NLCD 2001; Appendix 3) acquired through LOJIC. NLCD 2001 is a database developed from a consortium initiated by the U.S. Geological Survey to interpret land-cover attribute categories from Landsat 5 and 7 images (Homer et al. 2004). Pixels from these images were assigned digitized landcover attribute categories with a spatial resolution of 30-m. This digitized layer was used to determine the areal proportion of land represented by each category at the two buffers surrounding each site as mentioned previously (1-km and 500-m buffers). The proportions were entered as environmental variables into an ordination analysis for species composition at each site to determine which categories correlated best with species composition.

Stream order was determined using ArcGIS 9.0, with first order streams having no permanent tributaries, second order streams occurring at the junction of two first order streams, and third order streams at the junction of two second order streams. Cumulative catchment area was determined by adding subcatchment areas upstream of the site to the area of the subcatchment in which the site was located. This was done because catchment size is believed to affect stream conditions; therefore, it could be important for riparian plant composition. To explore relationships between species identity, percent exotic species, and the measures of community diversity with a measure of socio-economic status of land owners, property values were used, as listed by the Property Valuation Administrator offices in Jefferson and Oldham counties. The value of the parcel of land in which the site was located was taken as the property value. In instances where the site was located on the edge of a park abutting residential housing units or businesses, the values of surrounding parcels was used. Where sites were located in the interior of a park or natural area, the property was assigned a value of \$1.

Statistical Analyses

Potential relationships between plant species composition, degree of impervious surface, NLCD land-cover categories, and other environmental parameters listed below were explored using non-parametric ordinations, indicator species analysis (Dufrene and Legendre 1997), and cluster analyses. Software packages used were PC-Ord v. 4.41 (McCune and Mefford 1999) and R v.2.11 (R Development Core Team 2009). Non-metric multidimensional scaling (NMS) in PC-Ord was executed according to the recommended procedure outlined in McCune & Grace (2002). For each analysis, a separate starting configuration was supplied based on an initial analysis with the following configuration: Sorensen distances, 50 runs with real and random data, a random starting configuration, and 200 iterations to evaluate stability of the final solution (stability criterion ≤ 0.0005). Final solutions were completed using one run with real data. Ordinations run using R were given an optimal geometric starting configuration based on principal coordinates analysis of the same data. The statistical significance of the indicator value is assessed using a Monte Carlo test for significance, with a p-value indicating the probability of finding the computed stress value for a specific number of dimensions by chance (based on 50 permutations).

Indicator species analysis is a method of determining the value of a species to a particular grouping of sites (Dufrene and Legendre 1997). This method utilizes relative abundance and relative frequency of occurrence of a species within a particular group of sample units to assign a proportional indicator value. The statistical significance of the indicator value is assessed using a Monte Carlo test for significance, with a p-value

indicating the probability of finding the observed indicator value by chance (based on 1000 permutations).

The goal of cluster analysis was to obtain groupings of research sites based on commonalities in species composition. To do this, a site dissimilarity matrix was constructed based on groundcover species composition. Several dissimilarity measures and clustering techniques were applied to the data in an attempt to achieve the best cluster configuration for the sites. I decided upon complete clustering as it produced the highest within-to-among cluster similarity (Partana ratio) with the most even distribution of sites among clusters. The Bray-Curtis distance measure was used to create a dissimilarity matrix of the sites prior to cluster analysis. To decide on the number of groupings and assess their overall quality, I used the Partana ratio, as mentioned, as well as silhouette plots. Figure 3-3 shows an example of a silhouette plot for three groupings. Silhouette plots assign numerical values (between -1 and 1) termed “silhouette widths” (S_i) to each research site (i) within a cluster. The silhouette width is obtained by comparing a research site’s mean similarity to other sites within the cluster, followed by comparison to its mean similarity to sites within the nearest cluster. A silhouette width of +1 means the within- cluster similarity is much higher than the between-cluster similarity, indicating a good fit of that site to the cluster. A value of -1 means the between-cluster similarity is much higher than within-cluster similarity, indicating a poor fit of that site to the cluster. An average silhouette width is computed for each cluster in order to compare quality of the groupings. An average silhouette width is determined based on site silhouette widths in order to compare different numbers of groupings, where

the goal is to obtain a value as high as possible without compromising the quality of the individual clusters.

To determine whether measures of riparian groundcover species abundance and diversity (% cover, diversity indices) varied with the proportion of subcatchment impervious surface, I used scatterplots and a single factor model (ANOVA, R v.2.11). Scatterplots were generated using all sites, and a subset of sites occurring along first order streams to control for increasing catchment area. ANOVA was used for partitioning variance among land-use categories (urban, suburban, rural), and a post-hoc test (Tukey's HSD pairwise comparisons) was used for determining statistical significance of the variance between pairs of land-use categories. Data were assessed for homoscedasticity prior to analyses and were transformed to better meet those assumptions, if necessary. In addition, any potential outliers were identified using quantile by quantile (Q-Q) plots and graphs of leverage vs. standardized residuals plotted with Cook's distance. Points appearing to assert a disproportional influence on the data were sequentially eliminated from the analysis to assess effects on the statistical assumptions of homoscedasticity and homogeneity of variance. Data points were permanently removed from the analyses if their elimination improved these statistical assumptions.

Results

Subcatchment delineation & classification

Forty subcatchments were delineated based on topography within and near the three main catchments. Sites were classified into land use categories based on % impervious surface cover (ISC) at three scales: subcatchment, 1-km and 500-m radius site

buffers (Table 3-1). There were twelve urban subcatchments, with one in Goose Creek watershed and eleven in Beargrass Creek watershed. Rural subcatchments included eleven in Harrods and three in Goose Creek. Suburban subcatchments spanned all three catchments, with eight in Goose and three each in Beargrass and Harrods Creek watersheds (Table 3-1). Impervious surfaces within 500-m and 1-km of each site generally decreased compared with values at the subcatchment scale. Therefore, as the scale of the buffer distance around each site decreased, the number of sites classified as urban fell, while sites classified as suburban and rural increased (Table 3-1). One site in Beargrass Creek watershed on a privately owned farm was reclassified as rural when analyzed using the 500-m radius site buffer. The site was kept primarily in mown grass, with a thin strip of woody vegetation allowed to persist adjacent to the creek. All sites with $\geq 30\%$ ISC within 1-km and 500-m radii of a site were located in Beargrass Creek watershed.

Riparian zone widths

Riparian width varied with catchment scale land-use. Suburban sites exhibited the smallest mean width, while rural sites had the highest mean/median width and so had the greatest number of multi-plot sites (Table 3-2). Six of fourteen rural sites, four of fifteen suburban and three of twelve urban sites were less than twelve meters wide, resulting in smaller nearstream plot sizes in these sites. Only two urban sites contained more than one plot. One of the sites, located in a city park (Cherokee Park), contained two plots. The other site accommodated three plots, since it extended approximately 100-m before

intersecting a highway. All suburban sites contained only one plot as all vegetated riparian sites were less than 30 meters wide.

The distribution of riparian widths at urban and rural multi-plot sites was similar (Table 3-2). In rural subcatchments, two of the multi-plot sites were along 1st order reaches, one occurred along a 2nd order reach, and two lay adjacent to 3rd order streams. One of the rural multi-plot sites along a 1st order reach appeared to have an altered stream flowpath, as it was located along a residential street and directly abutted an adjacent upland slope. The two multi-plot sites in the urban subcatchments occurred along 2nd and 3rd order reaches.

The distribution of narrow riparian zones with regard to stream order exhibited greater variation between land-use categories (Table 3-2). Seven of the sites with riparian widths less than 12 meters were found along first and second order streams. In suburban subcatchments, three of the smaller plots were located along 1st order streams, and one was located along a 3rd order stream. Two of the urban riparian sites with riparian widths less than 12 meters were located along 1st order streams, and one along a 3rd order stream. Both of the narrow sites along 3rd order streams (one along urban and one along a suburban stream) appeared to have been dramatically altered, possibly to contain floodwaters. In the urban area, the first terrace out of the stream channel contained a narrow riparian zone (< 12-m) which led to a vertical incline and second terrace approximately six feet higher (at the urban site) than the riparian zone. The opposite bank on the other side of the stream channel was practically identical. Such uniformity suggests human intervention. Alterations of this sort, while not surprising,

carries implications for some of the ecosystem services provided by riparian zones in urban areas.

Variation in Plant Community Structure by Land Use

Across all 41 plots in all three watersheds I found there were 96 groundcover species, with 88 being herbs and 8 woody vines, and with 78 being native and 18 exotic. Across all sites the top four most frequently occurring species were *Ageratina altissima*, *Sanicula odorata*, *Glechoma hederacea* (exotic herb) and *Euonymus fortunei* (exotic woody vine). *A. altissima* was the most frequently occurring species in rural and suburban sites, and *E. fortunei* was the most frequently occurring species in urban sites (site land-use categories determined at the subcatchment scale). *E. fortunei* also had the highest mean % cover in urban riparian areas (16.8%), which was approximately five times greater than the second most abundant species, *A. altissima* (3.1%, Table 3-3). *G. hederacea* had the highest mean % cover of all species in suburban areas (8.1%), which was nearly twice that of the next highest species, *E. fortunei* (4.3%). *A. altissima* was the most abundant species in rural riparian areas (3.9%), covering approximately the same average area in urban and suburban locations.

In addition to differences in individual species abundances, differences in groundcover species diversity were discovered as well. I found that groundcover species richness was significantly reduced with increasing % IS at the subcatchment scale ($p = 0.003$, adj. $r^2 = 0.17$; Fig. 3-4). Total species richness across all plots increased from urban to suburban and rural sites from 38 to 58 to 77 species, respectively, with woody vine species remaining fairly constant (7 to 8 species) for all land uses (Table 3-3). Other

diversity indices (Shannon's, Simpson's reciprocal, and Shannon's equitability) did not exhibit a linear pattern of change with % IS, so I explored differences among these measures using categorical analyses. Groundcover plant diversity in urban nearstream plots was lower than that in rural riparian areas. Both Simpson and Shannon's indices showed similar trends at this scale, but only Shannon's index was statistically significant ($p = 0.023$; Fig. 3-5a). Shannon's index for rural plots was higher (mean = 1.61) than for urban plots (0.99; $p = 0.02$). While Simpson's reciprocal index did not differ statistically among land-use categories ($p = 0.088$; Fig. 3-5b), it did show a higher mean value in rural plots (mean = 3.88) than in urban plots (mean = 2.35; $p=0.077$ for this two-way comparison). Equitability of the riparian plots did not differ among land-use categories (urban mean $E_h = 0.53$, suburban and rural means $E_h = 0.61$; Fig. 3-5c).

Abundance measures of total groundcover and herbs did not show any significant linear or uniform non-linear trends with increasing subcatchment %IS (Appendix 6). Vine cover increased with increasing impervious surface, but the data were highly heteroscedastic. Percent herbaceous cover declined in a portion of the sites located in subcatchments with greater than 20% impervious surface cover. Percent vine cover attained a maximum of 70% where subcatchment impervious surface cover reached 30%, and herb cover steadily declined to below 30% in those sites with 40% or more impervious surface cover. Proportions of exotic groundcover (herbs and woody vines) revealed no significant patterns regardless of this uneven distribution of vine and herb cover (Appendix 6).

A subset of sites along first order streams were assessed separately to control for variation in plant community variables related to differences in cumulative catchment

area. A significant reduction in diversity measures (richness, Shannon's and Simpson's indices) occurred with increasing %IS cover (determined at 1-km site radius scale; Fig 3-6). Pearson's r values did not exceed 0.5 for any of the diversity measures. Correlations with % IS were as follows: Shannon's diversity, $r = 0.48$ ($p = 0.01$); Simpson's diversity, $r = 0.41$ ($p = 0.02$); and species richness, $r = 0.4$ ($p = 0.03$). Evenness (Shannon's equitability index) did not vary with % IS, but did show a positive correlation with bank height adjacent to plots along first order streams ($p = 0.004$, Pearson's $r = 0.57$; Appendix 7). The pattern, however, seemed driven by a small number of plots at the very low end of bank height (< 50-cm) and at the high end (200-cm). The plot with the highest bank height only contained two groundcover species, each having less than 1% cover, but with approximately equivalent cover values, resulting in a relatively high evenness value. Abundance measures (% cover) did not show any significant relationships with %IS; however, woody vine % cover was greatest within two plots at 25 and 30 %IS cover (determined at the subcatchment scale; Appendix 7). The vine that covered most of the area within those plots was *E. fortunei*.

While species diversity differed among land-use categories, diversity also varied with lateral distance from the stream along the riparian zone. Species richness showed differences between urban, suburban, and rural riparian zones with lateral distance from the stream, although no differences were evident within those land-use categories. In urban subcatchments, species richness was half that of rural subcatchments within 8 meters of the stream (Fig. 3-7, Table 3-4). Urban richness (mean = 4.9 species) was also significantly lower ($p = 0.037$) than suburban richness (mean = 8.4) between 1 and 2 meters from their respective streams. Mean species richness in suburban areas decreased

by two and three species from the 1-2 meter to 10-11 meter distance from the stream, but remained approximately the same with distance from streams in urban and rural riparian zones. These differences could not be attributed to changes in canopy openness among the intervals, as no significant differences were evident within or among land-use categories in canopy openness with increasing distance from the stream. Two urban and two suburban nearstream plots were partially covered by lawn. One plot each in both urban and suburban categories contained lawn between 4 and 11 meters from the stream. A second site in both land-use categories contained lawn between 7 and 11 meters from the stream. A third urban plot was reduced to a single species (*Eunonymus fortunei*) at the 7-8 meter interval. Groundcover richness with distance from the stream appears more stable in urban and rural riparian zones than in suburban riparian zones, but with a lower mean richness in urban and a higher mean richness in rural riparian zones.

Riparian plant species richness also differed with downstream distance within land-use categories. I found a significant positive relationship between species richness in nearstream plots and cumulative catchment area within suburban riparian zones (Fig. 3-8). In contrast, no significant linear relationship was detected between these two variables within urban or rural land-use categories. Bank height adjacent to the plot was also used as an integrative index of flood disturbance, but no significant linear relationships were apparent with either groundcover species richness or other community diversity indices.

NMS & Cluster analysis – Groundcover species

I conducted a cluster analysis to obtain groupings of research sites based on commonalities in species composition. Cluster analysis of the groundcover layer resulted

in three clusters of 39 sites (out of 41) using complete clustering and Bray-Curtis distance. Two sites were not used since each one formed a separate cluster, indicating they were not a good fit with any of the other sites. The Partana ratio was 1.74 with an average silhouette width of 0.11. Cluster-1 contained the greatest number of sites (nineteen) with an average silhouette width of 0.13. Cluster-2 contained the fewest sites (eight) with an average width of 0.12. Cluster-3, with twelve sites, was the weakest cluster with an average silhouette width of 0.06.

Clusters were analyzed for differences in species composition using frequency of occurrence within sites making up the clusters and indicator species analysis. Those species with significant indicator values in each of the clusters ($p \leq 0.05$) are shown in Table 3-5. Assemblage identity of cluster-1 was defined by the indicator species *Glechoma hederacea* (an exotic herb) and non-lawn (unmanaged) grasses. However, the grasses, *Microstegium vimineum* and *Elymus riparius*, were not included in this category as I was able to identify them to species. Other noteworthy species occurring with greater frequency though not exclusively in this cluster were the two native species, *Verbesina alternifolia* and *Impatiens capensis*, a wetland species. All sites in cluster-2 contained the indicator species *Euonymus fortunei*, an exotic vine that also occurred in cluster-1 with lesser frequency. The native woody vine, *Toxicodendron radicans*, was ubiquitous, but occurred with the greatest frequency in cluster-2. A mix of species defined cluster-3, as all indicator species occurred with a frequency of less than 0.5. *Lysimachia nummularia*, *Symphiotrichum cordifolium*, *Juncus* spp., and *Packera obovata* were present in at least 25% of all sites in this cluster. *L. nummularia* and *Juncus* spp. are classified as wetland (facultative or obligate) species, and all these species are native.

Non-metric multidimensional scaling (NMS) was used to determine cluster associations with environmental variables and species attributes. NMS was run using the 39 sites organized into a distance matrix (using Sorensen's index) based on groundcover species importance values (IVs). A 3-dimensional configuration was chosen, achieving a final stress of 16.6 and instability of 0.00048 for the final configuration. When sites were assigned to their respective clusters, no site overlap was visible among the three clusters in 3-dimensional space, indicating satisfactory agreement between the cluster analysis and the NMS solution (Fig. 3-9). Clusters were analyzed in relation to environmental variables that might correlate with and potentially help explain the observed variation in species composition among clusters (Table 3-6). Environmental variables calculated for each site were entered into a site-by-variable matrix and included as a secondary matrix in NMS using PC-Ord. The biplot feature in PC-Ord was used to generate a line for each variable with an $r^2 \geq 0.15$ for one or more axes, with the direction of the line corresponding to the direction of positive correlation of the variable and the length of the line indicating the strength of the correlation along one or more axes. Implied in the ordination plot but not visible, a vector of equal length but opposing sign points in the opposite direction to indicate a negative correlation.

NLCD categories and % impervious surface at various scales were included in the NMS as environmental variables to evaluate cluster identities with fine and coarse scale land-cover variables (Table 3-6; Fig. 3-10). The importance value of *L. maackii* for each site was included as an environmental variable to determine if the species might be a possible influence on groundcover species composition. Cluster-2 in the 3-cluster configuration was the most "urban" of the clusters, being most strongly and positively

correlated with % IS (ranging between 12% and 48%) within a 1-km radius of each site center, as indicated by the length of its vector (Fig. 3-10a,b). Since impervious surface cover at the 1-km scale showed the strongest correlation with the NMS axes, NLCD database land-cover categories at that scale were used for further analyses. Low-intensity developed land most strongly correlated with this cluster (Fig. 3-10c,d). This type of land-cover is typically composed of single-family residential housing, which was the most frequently observed land-cover type found in this study. To a lesser extent, medium-intensity and open-space developed land also positively correlated with the species composition of cluster-2, followed by high-intensity developed land, which occurred around a few of the most urban sites. *Lonicera maackii* was slightly more important in explaining cluster-2 than the other clusters. While all clusters contained sites with honeysuckle, most sites within cluster-2 had an *L. maackii* IV greater than 75. Only one site contained no honeysuckle (IV = 0), and two sites had a *Lonicera* IV of 200 (maximum IV). Clusters-1 and -3 were most positively correlated with the proportion of deciduous forest cover and pasture (Fig. 3-10c-d). Cluster-1 contained between 8% and 82% deciduous forest cover and cluster-3 between 18% and 68% deciduous forest cover within a 1-km radius of the sites. Pasture comprised between 0% and 39% cover in both clusters. Wetland cover was located primarily within a radius of cluster-1 sites, with eight of the sites having between 0.1 and 0.5 % cover for this habitat type. Most sites in clusters-1 and -3 contained less than 24% IS cover within a 500-m radius, but three sites contained up to 45%. Low-intensity developed land cover occurred with a proportion up to 27% around the sites in these two clusters. While primarily located within a mixture of

deciduous forest and pasture, the influence of urban development was therefore detectable.

Species attributes (Appendix 2) were included in the analyses as environmental variables to assess which attributes associated with the same site clusters as the land-cover variables. In this way an indirect correlation could be drawn between species attributes and land-cover variables. Figure 3-11 shows the direction of correlation of these attributes in relation to the various site clusters. Obligate wetland (OBL) and native species attributes most strongly correlated with clusters-1 and -3. Facultative species (FAC) did not positively associate with any particular grouping of sites. The vector for exotic species (E) extended in opposition to the native vector (N), correlating most strongly with Clusters-1 and -2.

L. maackii and *E. fortunei* analysis

Since *L. maackii* and *E. fortunei* were important species associated with urban clusters, I decided to determine if their distributions and densities were related. Only nearstream plots were used for these analyses. First, all plots containing *E. fortunei* were included in the analysis, regardless of whether they contained *L. maackii* (Fig. 3-12a). A strong positive correlation was shown between *L. maackii* stem density and *E. fortunei* percent cover (Pearson's $r = 0.73$, $p = 0.0001$). When only plots containing both species were included in the regression (Fig 3-12b), an even stronger positive correlation emerged (Pearson's $r = 0.81$, $p = 0.001$).

I then determined whether measures of groundcover species diversity in nearstream plots decreased with increasing density and cover of *L. maackii* and *E.*

fortunei. Where *E. fortunei* was used as a correlate with diversity measures, that species was not used in calculations of species diversity. In plots containing *L. maackii*, a weak negative correlation was found between *L. maackii* density and groundcover species richness (adj. $r^2 = 0.19$, $p = 0.019$; Fig. 3-13a) and Shannon diversity (adj. $r^2 = 0.20$, $p = 0.012$; Fig. 3-13b). No trends were detected between *L. maackii* density and Shannon's equitability index. However, % cover of *E. fortunei* was negatively, but weakly, correlated with Shannon's equitability index (adj. $r^2 = 0.12$, $p = 0.06$; Fig. 3-14). Relationships with Shannon's index, Simpson's reciprocal index, and species richness can be seen in Figure 3-15. Values of *E. fortunei* % cover above 30% showed consistently low levels of diversity.

Discussion

Exotic woody and herbaceous vine species were the most abundant groundcover occurring in urban and suburban riparian zones. *E. fortunei* occurred with an average site cover of 17% in urban riparian sites, and *G. hederacea* covered 8% of suburban riparian sites. *E. fortunei* was the second most abundant species in suburban riparian zones, with an average cover of 4.3%. The dominant cover in rural riparian zones, the native herb *A. altissima*, also occurred in urban and suburban riparian sites with approximately the same average cover (rural mean = 3.9%, suburban mean = 3.3%, urban mean = 3.1%). Urban and suburban areas are notorious for their associations with exotic species. These findings indicate that while these areas are more susceptible to invasion by exotic species, native species that are abundant in rural environments are not necessarily being excluded from urban and suburban riparian zones.

Species richness of all groundcover species significantly declined with increasing subcatchment impervious surface (Fig. 3-4 and 3-6a). This differed from my original prediction of greater richness in suburban areas in that richness was just as high or higher at the rural end of the gradient. This may indicate more resource heterogeneity or less stressful conditions across rural riparian areas enabling support of a greater number of species. Reduction in groundcover species richness along the gradient could also be partially attributed to the densities of *L. maackii*, which I (Fig. 3-13a) and others (Luken and Thieret 1996, Hutchinson and Vankat 1997, Borgmann and Rodewald 2005, Castellano and Boyce 2007) have shown to be associated with reduced richness of groundcover plants. I had shown in a study of woody vegetation at these same sites that *L. maackii* importance value was significantly positively correlated with proportion of impervious surface within a 500-m site radius (See Fig. 2-17 in Dissertation Chapter 2), demonstrating it to be a dominant riparian species in urban Louisville.

Other measures of species diversity were used as response variables to gauge how degree of subcatchment urbanization affects groundcover plant communities. For nearstream plots both Shannon's and the Simpson's reciprocal indices of diversity were lowest for urban and highest for rural subcatchments (Fig. 3-5). Scatterplots from riparian sites along first order streams also supported this pattern (Fig. 3-6b-c). Since Shannon's equitability index did not differ between land-use categories, differences in the indices were primarily due to differences in species richness. While fewer groundcover species exist in urban riparian areas than in suburban or rural riparian areas, the species that did exist are distributed in similar proportions, regardless of surrounding land-cover/land use. Comparison of the groundcover layer among land-use categories supports the conclusion

that vegetation in rural riparian zones is generally more diverse than in urban riparian zones.

As resources and environmental conditions can change within a riparian zone laterally as well as longitudinally up and downstream, I predicted that groundcover species diversity would vary with distance from the streambank. No significant differences were found between distance intervals within land-use categories. However, when comparing distance intervals between land-use categories, urban richness was shown to be significantly lower than rural richness within eight meters of the stream, but not beyond that distance. Urban richness was also lower than in suburban plots within two meters of the stream bank edge (Table 3-4, Fig. 3-7). These findings suggest that, in addition to the availability of a larger species pool, nearstream conditions within rural riparian zones could be a factor favoring higher species diversity than those in urban riparian zones. Suburban riparian zones possess a greater species pool only at the distance interval nearest the stream. As distance from the streambank increases, suburban riparian plant diversity becomes more similar to urban diversity. However, this low suburban diversity pattern was in part due to the presence of lawns, which I considered as monocultures within urban and suburban riparian sites. Lite et al. (2005) also found decreasing herbaceous species richness with increasing distance from the stream channel in an arid environment in Arizona. Following rains and flooding at the same sites, the pattern was reversed, with herbaceous richness increasing farther from the stream. These patterns were attributed to changes in soil water availability and drainage. Therefore, variation in soil water availability is one possible explanation for richness differences between urban, suburban, and rural areas in this study. I measured streambank height as a

proxy for groundwater levels in all plots, and actually measured depth to groundwater in a subset of nearstream plots to examine whether relationships existed between these variables and variation in riparian plant communities. The highest measured streambank heights occurred adjacent to urban plots (mean = 122-cm, vs. 96- and 86-cm in rural and suburban plots, respectively). Streambank height and mean depth to groundwater (see Dissertation Chapter 4) could then partially explain the lower richness at urban sites at least within the 1-2 meter interval. Mean depth to groundwater was found to be lower along urban riparian zones than suburban or rural riparian zones over a period of one year (see Dissertation Chapter 4). The contrast was most stark in the spring, when rainfall allowed recharge of rural and suburban groundwater tables but not urban water tables (see Fig. 4-4 in Dissertation Chapter 4). Further research would need to be conducted in these areas to determine whether groundwater decreased or increased with lateral distance from the stream to more fully understand how this variable might affect herb and woody vine cover in riparian areas along urban-to-rural gradients.

Upstream-to-downstream (longitudinal) gradients in plant species diversity have also been shown to exist in riparian environments. Since flooding intensities and frequencies vary with upstream catchment area, it is a likely contributor to the differences in richness seen with longitudinal distance downstream. As such, I used cumulative upstream catchment area in lieu of longitudinal downstream distance, even though this measure has been referenced in previous studies (Bendix 1997, Bendix and Hupp 2000, Lite et al. 2005). My prediction of higher species richness with greater upstream catchment area was only partially supported. While cumulative upstream catchment area was significantly and positively correlated with groundcover species richness within the

suburban land-use category (Fig. 3-8), it was not a predictor of species richness for riparian areas within either urban or rural land uses. Several streamflow parameters, which may have contributed to this pattern, have been shown to increase or decrease with increasing catchment impervious surface cover (Paul and Meyer 2001). These parameters include increases in bankfull and peak discharge, and decreases in lag time. As catchment area increases moving downstream, disturbance to the riparian zone can increase as flood velocity and volume increases (Bendix 1997). Although bank height increases with increasing upstream catchment area, no trends were detected between bank height and groundcover richness. Stream channel structure and width-to-depth ratios are additional measures that should be compared and which might explain this species richness phenomenon, as both could affect severity of flooding.

Plant community analysis

Site groupings via cluster analyses and NMS generally supported my prediction of distinct vegetation communities being correlated with impervious surface cover. Three communities were defined and analyzed for correlations with environmental factors and species attributes. Cluster-1 contained the greatest proportion of sites with surrounding wetland cover and was defined by unmanaged grasses and the exotic species *G. hederacea* (Table 3-5). The most urban cluster (cluster-2) was defined by the exotic species *E. fortunei* (Fig. 3-10). Although I expected vegetation communities to be most strongly correlated with impervious surface at the smallest scale measured (500-m site radius), this was not the case. This cluster correlated most strongly with measures of impervious surface and urban land cover within a 1-km radius of the sites.

Environmental correlates with the three clusters suggest they are composed of groundcover species with differential affinities for site conditions associated with proportions of impervious surface and deciduous forest cover, with wetland species possessing low affinities for surrounding impervious surfaces and certain exotic species exhibiting higher affinities. These findings provide partial support for my predictions regarding exotic vs. native and wetland vs. non wetland species. While some species associations correlated to a greater degree with impervious surface and urban land cover, no clusters showed complete fidelity to urban vs. suburban vs. rural categories, as each cluster contained several sites with at least moderate proportions of impervious surface and urban cover within a 1-km radius. Furthermore, the absence of strong patterns regarding exotic species richness and abundance proportions along an impervious surface gradient (Appendix 6) suggests the “urbanness” of an area does not necessarily predict the presence or abundance of exotic species. However, cluster correlations with the exotic shrub, *L. maackii*, suggest that this species influences groundcover species at the plot scale, no matter what land-cover type the plot was surrounded by within a 1-km radius.

Groundcover plant associations with shrub honeysuckle

In another study (see Dissertation Chapter 2), I found a strong positive correlation between importance value of the exotic shrub, *L. maackii*, in plots and impervious surface cover within a 500-m radius of research sites. In this study I also found that the exotic woody groundcover vine, *E. fortunei*, often co-occurred with this shrub. The percent cover of *E. fortunei* dramatically increased in the presence of high densities of *L. maackii* (Fig. 3-12), approaching 70% cover at the highest densities of the shrub. Where *L. maackii* was

not present, *E. fortunei* percent cover did not exceed 5% cover (Fig. 3-12a). Castellano and Boyce (2007) discovered a very different pattern regarding *L. maackii* and *Juniperus virginiana* along a road cut in northern KY, USA. Both species were found to coexist at the same site, where *L. maackii* demonstrated a clumped distribution, and *J. virginiana* exhibited a more random distribution. These two species were able to co-exist since they were able to partition resources between themselves at the same sites, with *J. virginiana* inhabiting areas with higher photosynthetically active radiation, indicating lower shade tolerance. In the case of riparian sites in this study, *E. fortunei* appears to thrive beneath dense *L. maackii* clusters, suggesting either that *L. maackii* may be facilitating growth of this species where they occur together, or conversely *E. fortunei* may promote successful germination and growth of *L. maackii*.

I also examined whether measures of groundcover species diversity decreased with increasing density and cover of *L. maackii* and *E. fortunei*. Both groundcover species richness and Shannon's index decreased with increasing densities of *L. maackii* (Fig. 3-13). This decreasing pattern of plant species richness with increasing exotic cover was also shown by Loewenstein and Loewenstein (2005), where understory plant species richness (saplings, shrubs, herbs, vines) decreased with increasing cover of the exotic shrub *Ligustrum sinense* in riparian forests near Columbus, Georgia, USA. In the Louisville sites, Shannon's equitability index was negatively but weakly correlated with *E. fortunei* percent cover (Fig. 3-14), but no correlation was found with species richness. However, in sites where this vine exceeded 30% cover groundcover diversity measures were all relatively low. While *L. maackii* and *E. fortunei* may co-occur, they appeared to interact differently with groundcover plants, with elimination of species under *L. maackii*,

and changes in species proportions where *E. fortunei* occurs. These results help explain variation in composition of species assemblages within the clusters. Cluster-2 was defined by *E. fortunei*, and nearstream equitable distributions of species declined as *E. fortunei* cover increased. Since *E. fortunei* cover increases in relation to *L. maackii* density, the shrub layer also helps to define groundcover communities in the more urban riparian sites where *L. maackii* dominated the shrub community (see Dissertation Chapter 2), lending support to my predictions that *L. maackii* is involved in structuring the plant community.

Conclusions

I found that urbanization in the Louisville Metro area is associated with decreasing diversity of riparian groundcover plant communities, reduced importance of obligate wetland species, and increased importance of the exotic vine, *E. fortunei*, a species sold by nurseries in the area for residential and commercial plantings. In addition, this study demonstrates the negative effects that the exotic shrub, *L. maackii*, is likely having on riparian herb and woody vine diversity. The co-occurrence of both these exotic species may be synergistically decreasing native species diversity in urban riparian sites. Furthermore, declining forested buffers reduce both riparian and instream diversity. I found presence of managed vegetation (lawns) contributed to the declining diversity of suburban riparian zones moving laterally from the stream. Moore and Palmer (2005) found declining instream invertebrate diversity with a declining proportion of forested riparian areas. These findings carry important implications since they show high amounts of impervious surface are not necessarily needed to affect riparian and instream diversity.

Simply reducing the complexity of riparian vegetation is sufficient to negatively affect this aspect of riparian environments. Also found in suburban riparian sites, increasing cumulative catchment area results in increasing species richness in but did not appear to explain species richness variation in urban and rural sites. Further research should concentrate on assessing the intensity and frequency of floodwaters in suburban environments, and assessing how they differ from urban and rural locations as upstream catchment area increases.

These findings suggest a change is needed on the conceptual model in Figure 1-1, with addition of an intermediary between the water table and riparian groundcover (Fig. 3-16). Although I found no direct link between bank height (and thus the water table) and species composition or richness, decreasing plant richness has been shown in arid environments with increasing depth to water table (Lite et al. 2005). However, I did find indirect evidence of a soil moisture difference with changes in proportion of impervious surface. Obligate wetland species were rare in most urban communities (Fig. 3-11), assuming greater importance in a more forested environment. Facultative and obligate wetland species are indicators of wetter soils at critical times of the year needed for germination and establishment. Therefore, a soil moisture gradient among sites is implied via the presence of these species in less urban environments. Therefore, I propose the changes in Figure 3-16 to better reflect potential conditions contributing to species composition in this system. Results from the woody species layer also support this model since facultative wetland species were more highly associated with more rural site clusters (see Fig. 2-14, 2-15, and 2-16 in Dissertation Chapter 2).

Table 3-1: Area of research catchments and the number of study sites classified as urban, suburban and rural at the subcatchment scale, and within a 1-km and 500-m radius from each study site. Classification as urban, suburban and rural land cover was based on proportion of impervious surface with $\geq 30\%$ being urban (U), $\leq 10\%$ being rural (R), and between 10 and 30% being suburban.

Catchment	Area (km ²)	SC			1 km			500 m		
		U	S	R	U	S	R	U	S	R
Beargrass Creek	65	11	3	0	8	6	0	6	7	1
Goose Creek	50	1	8	3	0	8	4	0	6	6
Harrods Creek	60	0	3	11	0	1	13	0	2	12

Table 3-2: Distribution of riparian width intervals in this study by stream order and land-use category at the subcatchment scale. Riparian zone width was measured along areas of level ground beginning at the streambank edge and extended until reaching a structural barrier (such as a road, parking lot, or building), upland slope, or other obstruction (grazed pasture, ephemeral stream channel). Classification as urban, suburban and rural land cover was based on proportion of impervious surface with $\geq 30\%$ being urban, $\leq 10\%$ being rural, and between 10 and 30% being suburban.

Land use	Stream Order	Riparian width (m)			
		<12	12-45	46-85	>85
Urban	1	2	4	0	0
	2	0	1	0	1
	3	1	2	1	0
Total		3	7	1	1
Suburban	1	3	7	0	0
	2	0	4	0	0
	3	1	0	0	0
Total		4	11	0	0
Rural	1	4	2	2	0
	2	2	0	1	0
	3	0	1	2	0
Total		6	3	5	0

Table 3-3: Mean % cover (+S.E.) for the top ten most abundant herb and woody vine species in urban, suburban and rural land-use categories.

Species	Urban plots ₁ Mean % cover (S.E.)	Suburban plots ₂ Mean % cover (S.E.)	Rural plots ₃ Mean % cover (S.E.)
<i>Ageratina altissima</i> (L.) King & H.E. Robins.	3.1 (1.5)	3.3 (1.4)	3.9 (2.4)
<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande	1.8 (1.2)	0.76 (0.71)	0
<i>Asarum canadense</i> L.	0.26 (0.18)	0	0
<i>Carex blanda</i> Dewey	1.0 (1.0)	0	0
<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz.*	16.8 (6.9)	4.3 (2.2)	0.94 (0.91)
<i>Glechoma hederacea</i> L.	1.6 (0.9)	8.1 (3.4)	3.3 (1.7)
<i>Impatiens capensis</i> Meerb.	0.28 (0.24)	0	0
<i>Solidago</i> spp. L.	0.92 (0.41)	1.4 (0.71)	0
<i>Urtica dioica</i> L.	0.59 (0.32)	0	2.7 (2.7)
<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	2.1 (1.4)	1.4 (0.72)	0.71 (0.33)
<i>Arundinaria gigantea</i> (Walt.) Muhl.	0	1.8 (1.3)	0
<i>Microstegium vimineum</i> (Trin.) A. Camus	0	3.4 (2.6)	3.2 (1.4)
<i>Parthenocissus quinquefolia</i> (L.) Planch.	0	1.1 (0.90)	0
<i>Viola</i> spp. L.	0	1.2 (0.77)	0.58 (0.21)
<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0	0	1.3 (0.57)
<i>Securigera varia</i> (L.) Lassen*	0	0	0.68 (0.46)
<i>Vinca minor</i> L.*	0	0	0.93 (0.93)
Mean % cover (herb and woody vine species)!	30.1 (6.8)	32.5 (6.1)	24.6 (5.1)
Mean % cover (herbs only)!	13 (3.7)	25.8 (5.9)	21.7(5.2)
Herb species richness	31	50	70
Woody vine species richness	7	8	7

₁ n=12, ₂ n=15, ₃ n=14 plots

*Woody vines

Land-use categories based upon % impervious surface at the subcatchment scale. Urban \geq 30%, Rural \leq 10%, Suburban between 10 and 30% IS.

! No significant differences were detected (ANOVA) among land-use categories regarding groundcover ($p = 0.63$) or herb-only ($p = 0.24$) mean % cover (arcsin transformed).

Table 3-4: Riparian groundcover mean species richness by distance from the stream bank and land-use categories determined at the subcatchment scale.

Land cover category	Distance interval from the stream			
	1-2 meters*	4-5 meters*	7-8 meters	10-11 meters
Rural	9.9 (0.009)	9.1 (0.025)	8.6 (0.029)	9.3
n =	14	14	10	8
Suburban	8.4 (0.037)	7.2	6.9	5.9
n =	15	13	12	12
Urban	4.9	5.0	4.2	4.8
n =	12	12	9	8

*Species richness was natural log transformed for 1-2-m and 4-5-m intervals for analysis, but values in the table are untransformed p-values (in parentheses) represent significant differences (at $\alpha = 0.01$ or 0.05) from urban riparian richness

Table 3-5: Frequently occurring and significant indicator species (using indicator species analysis (Dufrene and Legendre 1997)) for 3-cluster configuration using complete clustering. Frequency values were calculated based on the proportion of sites within the cluster in which the species occurred. Indicator values (IV) were calculated using indicator species analysis, where a value of 0 means the species is never present in the cluster and a value of 1 means the species is always present and exclusive to the cluster. Probabilities for the indicator values were determined using a Monte Carlo randomization technique.

Cluster	Species	Common name	Freq.	IV	p-value
1	<i>Glechoma hederacea</i>	Creeping Charlie	1	0.96	0.001
	Unmanaged grass	Grass	0.63	0.51	0.01
	<i>Verbesina alternifolia</i>	Wingstem	0.73		
	<i>Impatiens capensis</i>	Jewelweed	0.51		
2	<i>Euonymus fortunei</i>	Winter creeper	1	0.73	0.001
	<i>Toxicodendron radicans</i>	Poison ivy	0.62		
3	<i>Lysimachia nummularia</i>	Creeping Jenny	0.5	0.42	0.026
	<i>Symphiotrichum cordifolium</i>	Common blue wood aster	0.33	0.29	0.022
	<i>Juncus</i> spp.	Rush	0.25	0.22	0.041
	<i>Packera obovata</i>	Roundleaf ragwort	0.25	0.22	0.043

Table 3-6: Environmental and National Land Cover Database (NLCD) metrics used for non-metric multidimensional scaling.

Environmental variables	NLCD values (500-m)
-Bank height adjacent to the site	-Open-space developed
-% impervious surface: subcatchment, 1-km buffer, 500-m buffer	-High-, medium, low-intensity developed
-Plot size and distance from channel	-Deciduous forest
-Stream order	-Mixed forest
-Catchment Area	-Herbaceous forest
-Property value	-Crops
-Earthworm species number & weight	-Pasture
- <i>Lonicera maackii</i> IV	-Wetland

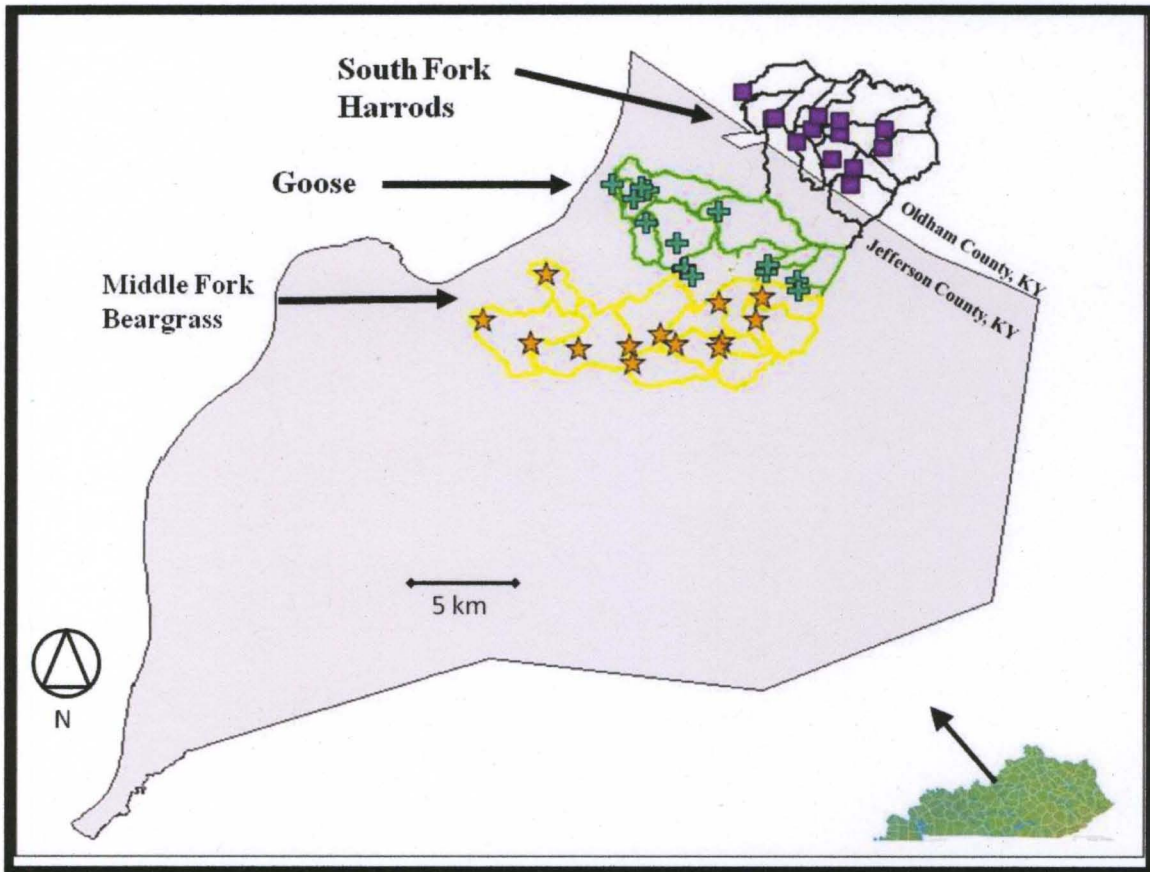
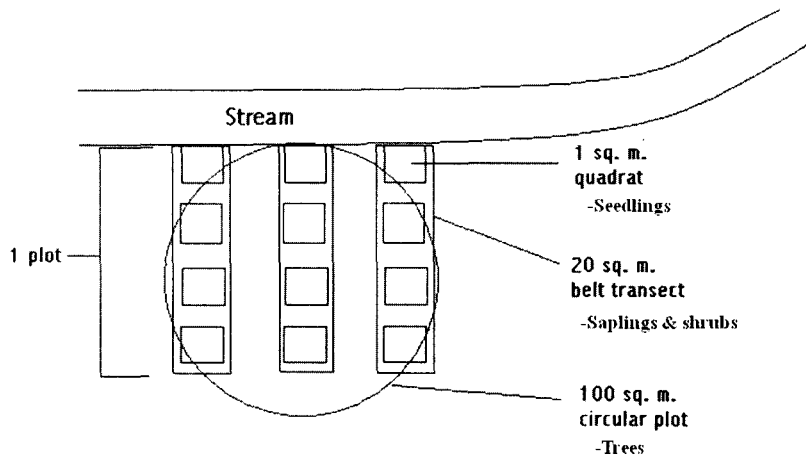
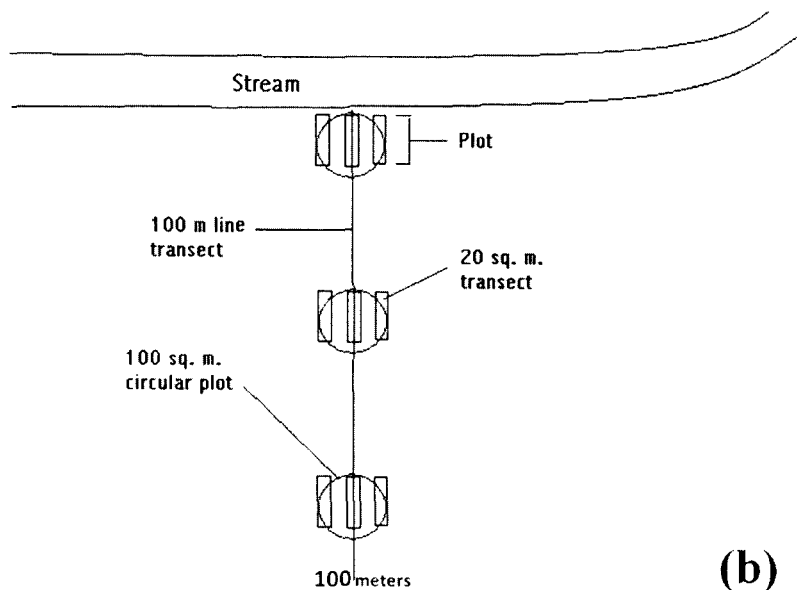


Figure 3-1: Schematic map of the three watersheds in Jefferson and Oldham counties in Kentucky. Research sites are indicated with markers.



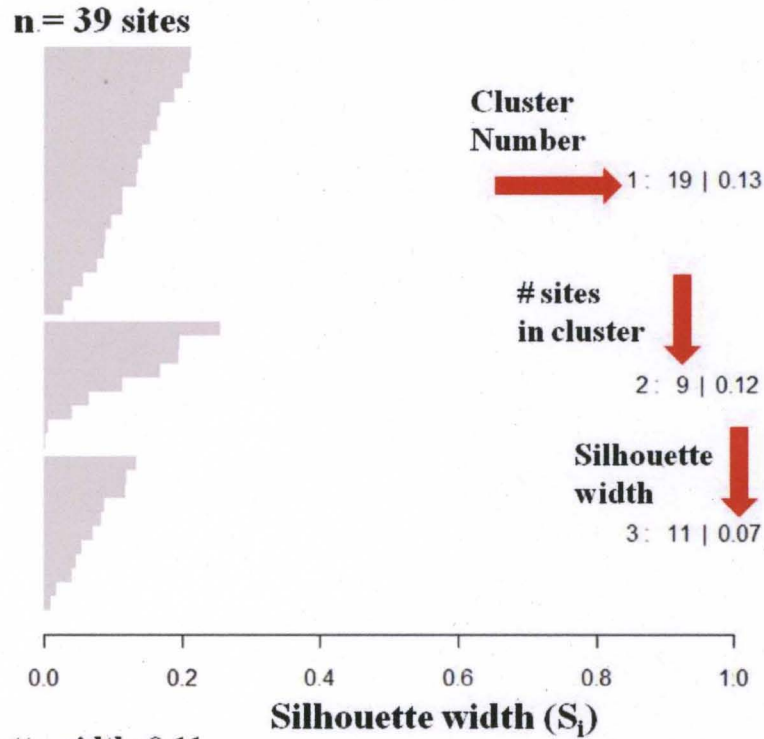
(a)



(b)

Figure 3-2a-b: Plot and site structure for sampling adult trees (≥ 2.54 cm DBH), saplings, tree seedlings, and shrubs. **(a)** Plots consisted of a 100-m² circular plot for sampling tree counts and DBH. Nested within the circular plot were three belt transects measuring 10 x 2 m with midlines spaced four meters apart for sampling sapling and shrub counts. Quadrats measuring 1 x 1 m were nested within the belt transects for determining percent cover and counts of tree seedlings. Adjacent quadrat spacing within belt transects was three meters at the midpoint. **(b)** Sites consisted of a midline transect stretched the length of the riparian zone up to 100 meters with up to 3 plots per transect with midpoints spaced 40-m apart.

Silhouette plot – 3 clusters



Overall silhouette width: 0.11

Figure 3-3: Silhouette plot of a cluster analysis (complete clustering using Bray-Curtis distance) resulting in three groupings of research sites. Sites within clusters are represented by gray bars, the length of each demonstrating the silhouette width (value on the x-axis) of that particular site within the cluster. Cluster numbers, number of sites within the cluster, and cluster widths are to the right of each cluster. Overall silhouette widths are located below the numerical axis. Explanation of values given in the text.

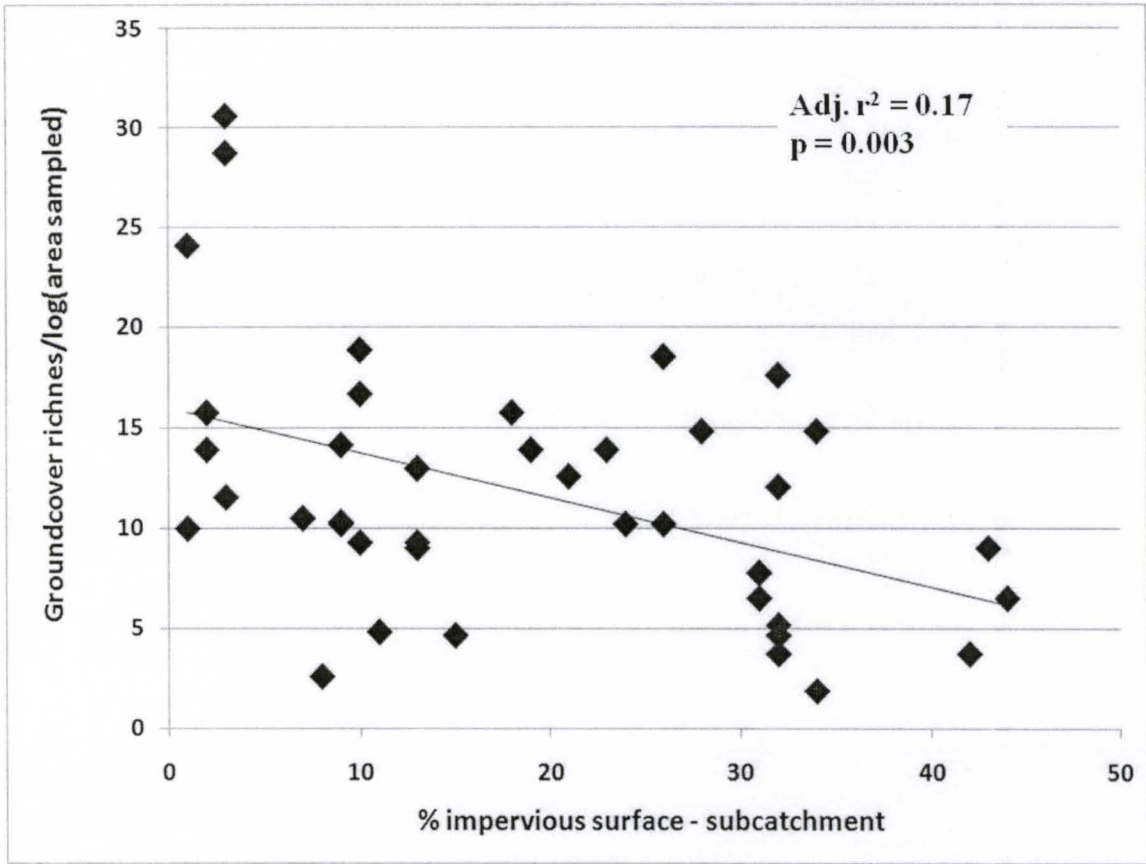


Figure 3-4: Groundcover species richness variation in nearstream plots with % impervious surface at the subcatchment scale. Species richness (S) was first normalized prior to analysis as $S/\log(\text{area sampled})$. The adjusted $r^2 = 0.17$ and the p-value = 0.003.

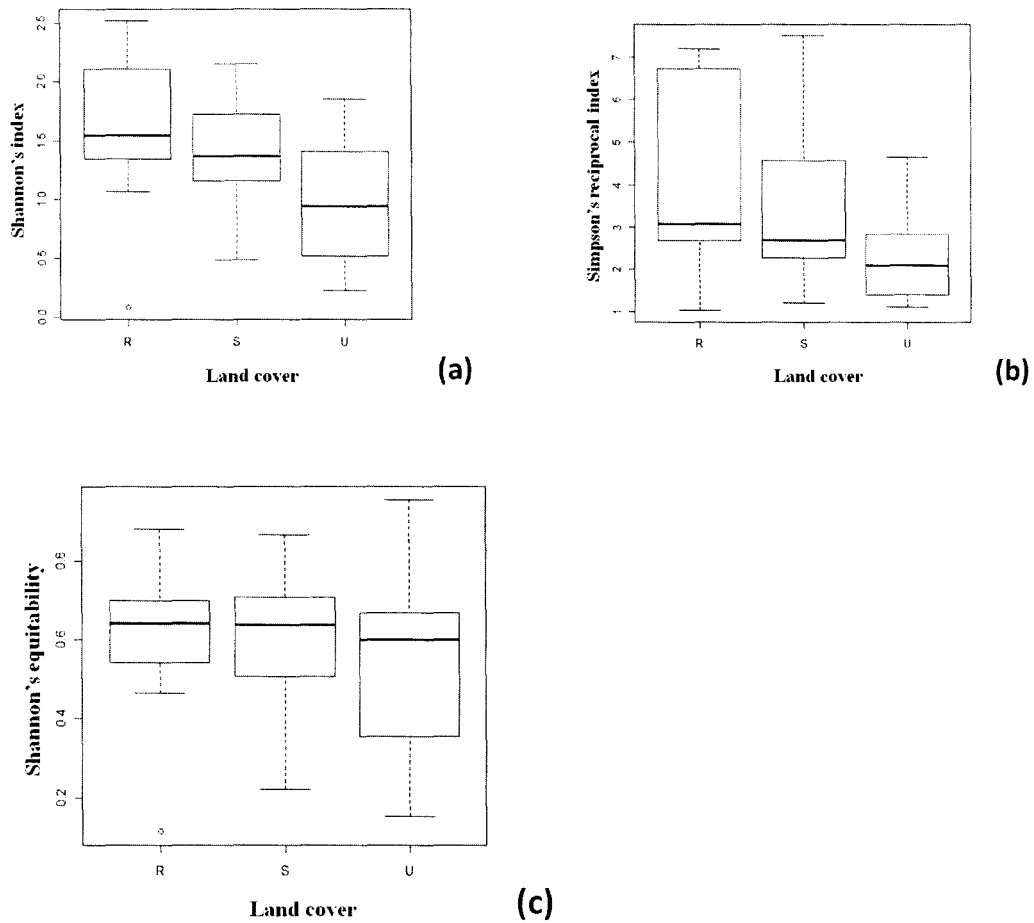
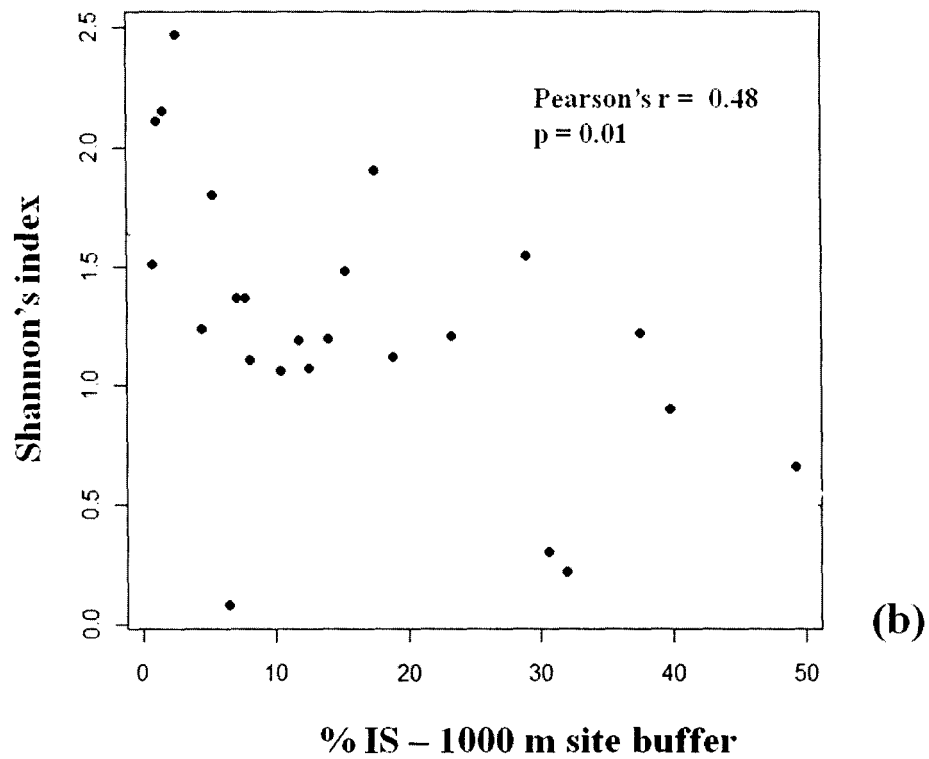
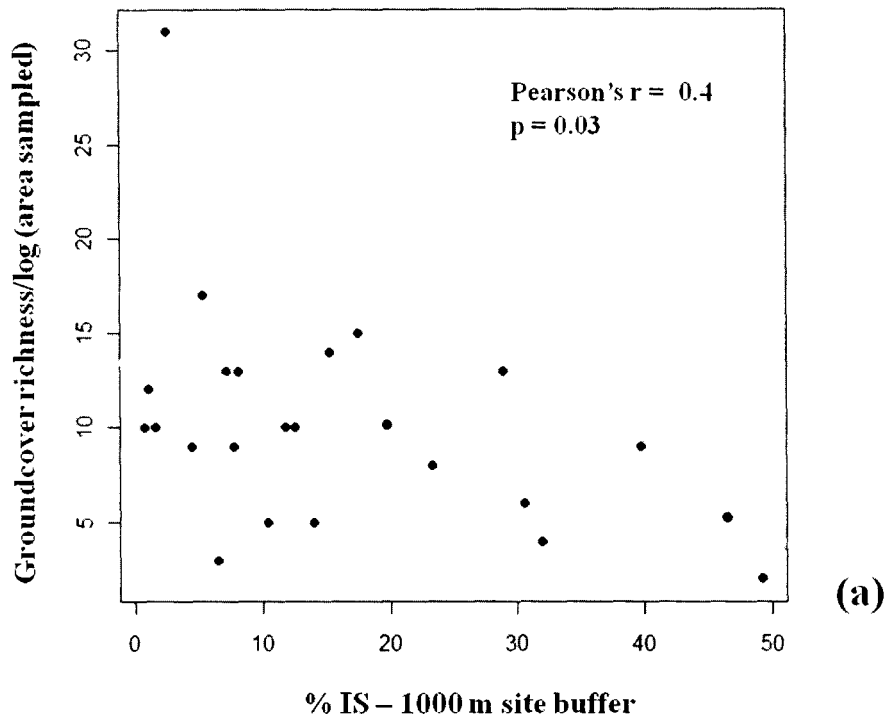


Figure 3-5a-c: Box-and-whisker diagrams of diversity indices computed for groundcover species in urban, suburban, and rural riparian zones. The bold line indicates the median, boxes delineate 25th and 75th percentiles, and whiskers indicate minimum and maximum values. (a) Shannon's, (b) Simpson's reciprocal, and (c) Shannon's equitability indexes were determined for all nearstream plots. Land use assignment for plots was determined at the subcatchment scale (U=Urban; S=Suburban; R=Rural). ANOVA's revealed a significant difference in Shannon's index (a) between land use categories ($p = 0.023$). Follow up analysis using Tukey's HSD pairwise comparisons revealed a higher Shannon's index in rural plots (mean = 1.61) than in urban plots (mean = 0.99; $p = 0.020$). Simpson's reciprocal index, while not exhibiting any significant differences ($p = 0.088$), did show the same trend where rural plots had a higher mean index value (mean = 3.88) than urban plots (mean = 2.35; $p = 0.077$). Shannon's equitability index did not significantly differ among land use categories. Open circles represent potential outliers.



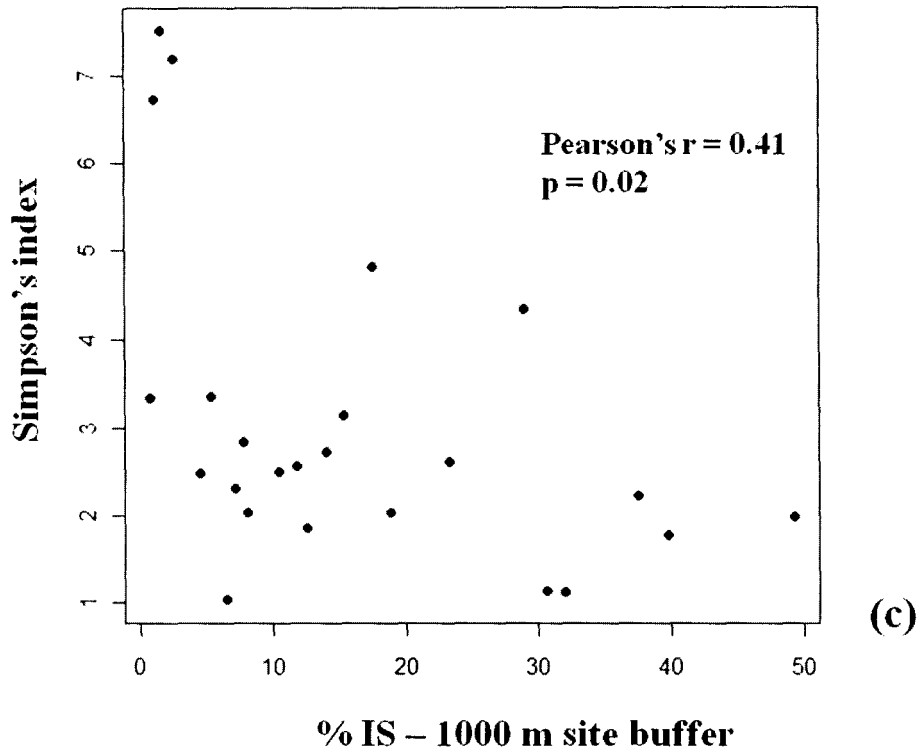


Figure 3-6a-c: Relationship between diversity measures of groundcover species at riparian plots along first order streams and % impervious surface cover within 1000-m of each plot. Diversity measures shown are species richness (a), Shannons index (b), and Simpson's reciprocal index (c). Species richness (S) was normalized based on plot size as $S/\log(\text{area sampled})$ since nearstream plot sizes varied.

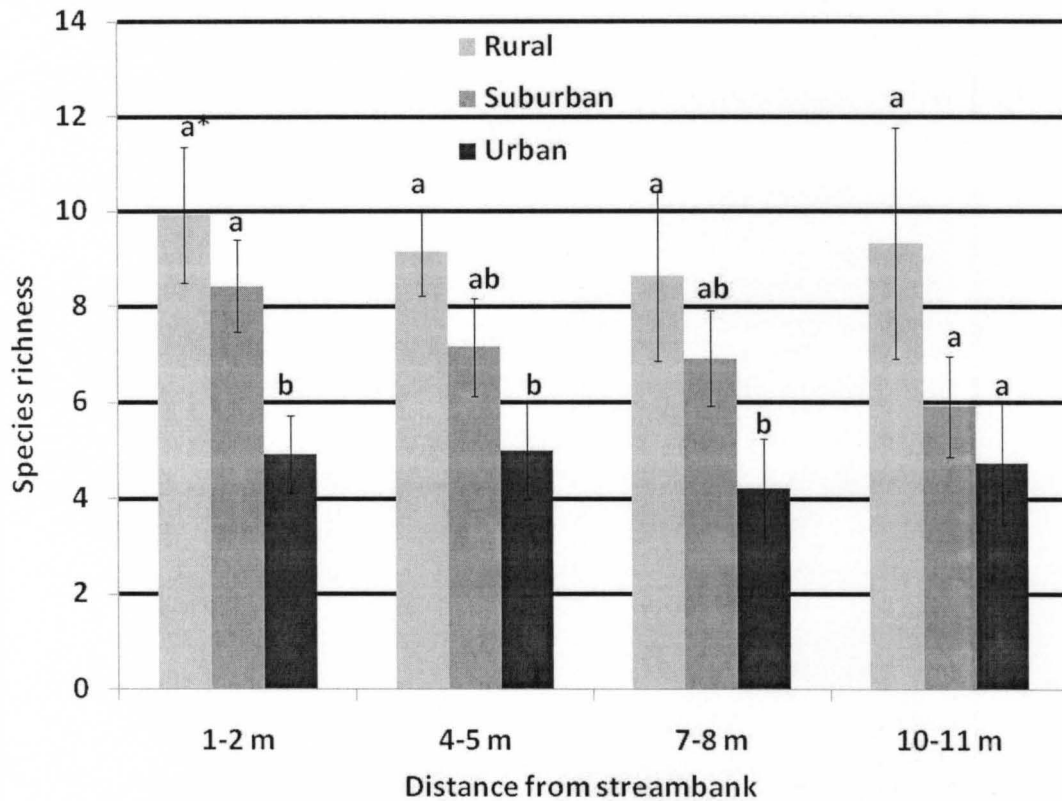


Figure 3-7: Mean groundcover riparian species richness at indicated distance intervals from the streambank in urban, suburban, and rural riparian zones. Land use assignment for plots was determined at the subcatchment scale. Data were obtained from plots nearest the stream. Each distance interval at each plot was composed of three 1 x 1 m² quadrats. Species richness was averaged by distance interval for each land use category. Differing letters above the standard error bars indicate significant differences at $\alpha = 0.01^*$ or 0.05. Richness values were natural log transformed for the 1-2 and 4-5-m intervals prior to analysis to meet the assumptions of ANOVA, although raw mean species richness values were used to create graphics.

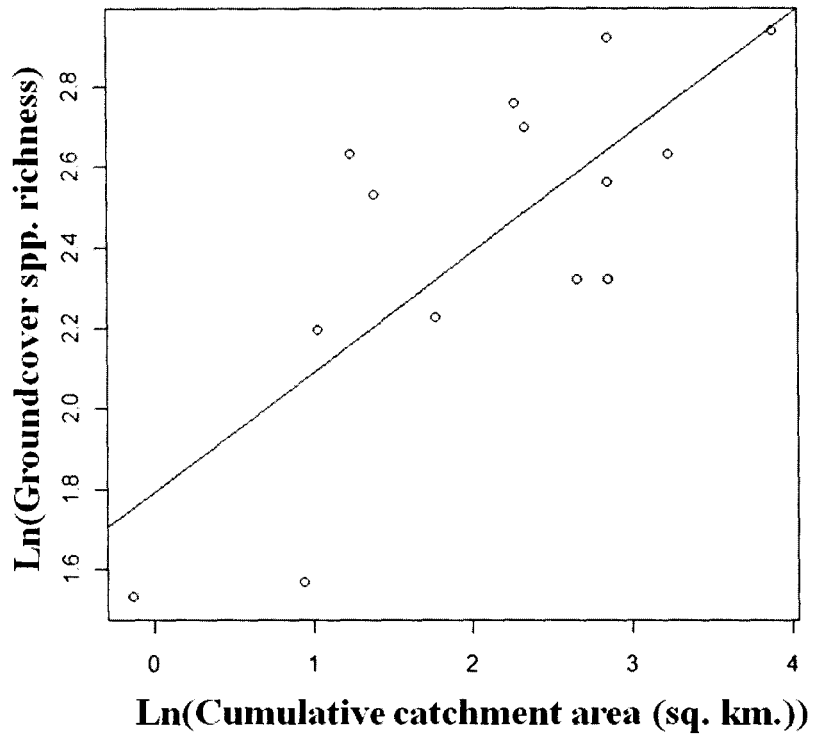


Figure 3-8: Linear regression model of cumulative catchment area upstream of the subcatchment for each suburban nearstream plot against normalized groundcover species richness ($S/\log(\text{area sampled})$). The adjusted $r^2 = 0.54$, and the p-value = 0.0011.

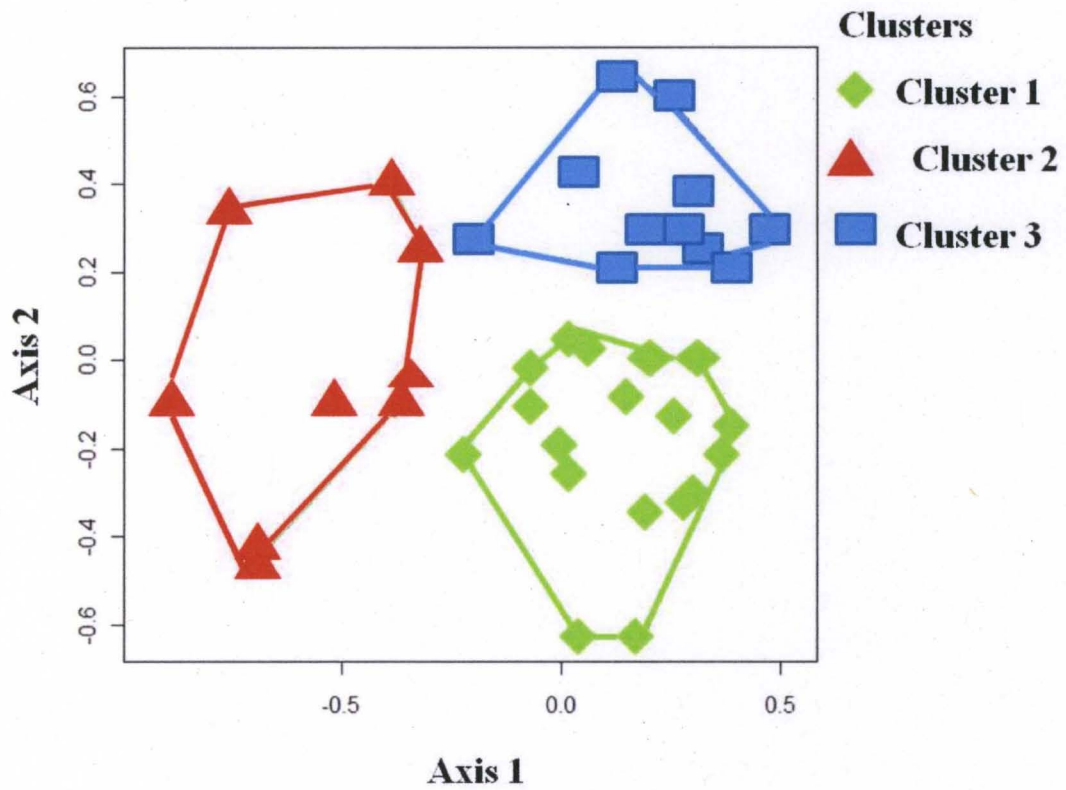
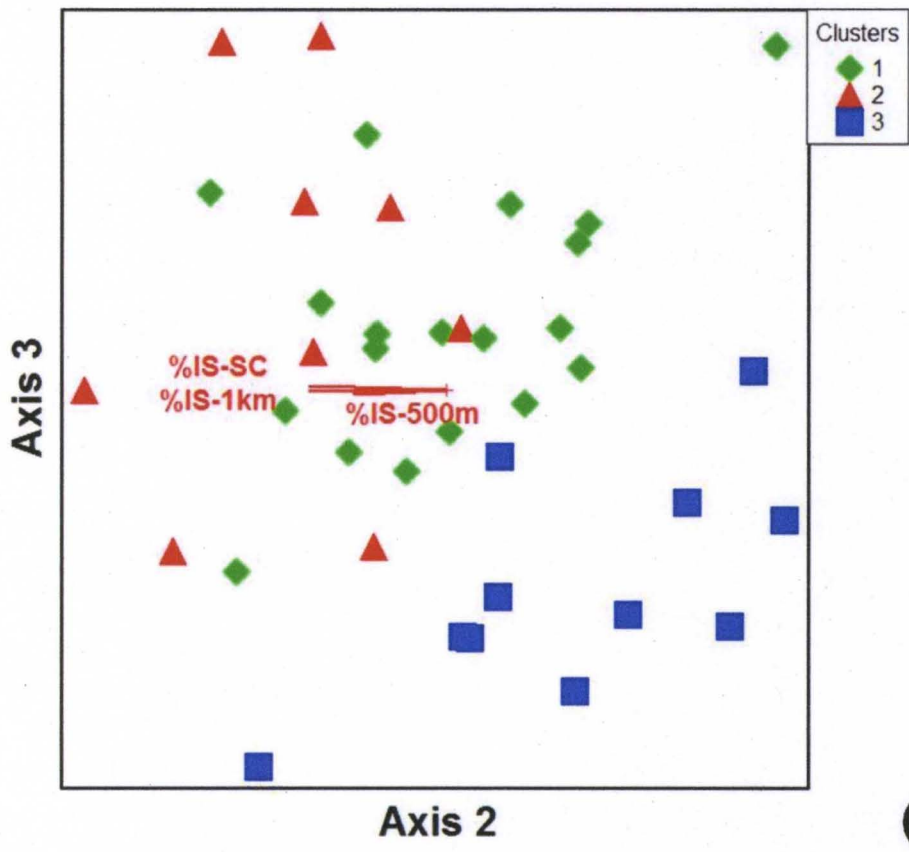
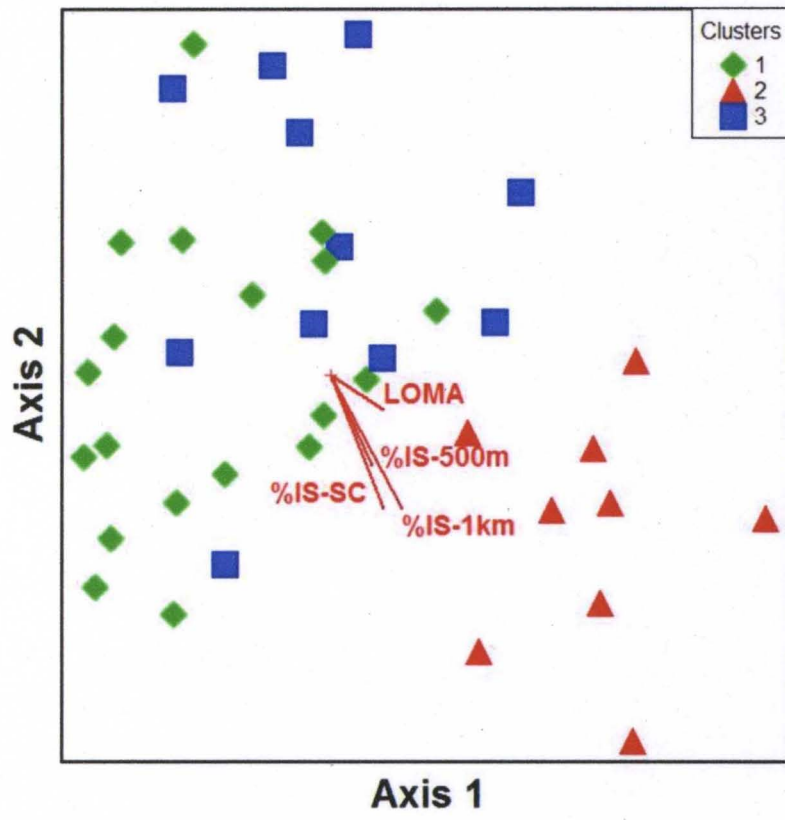
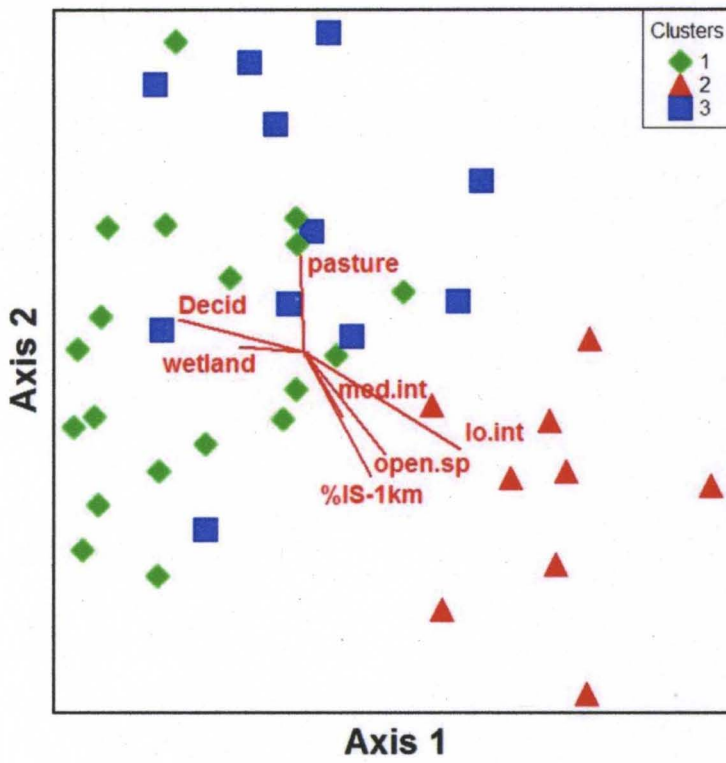
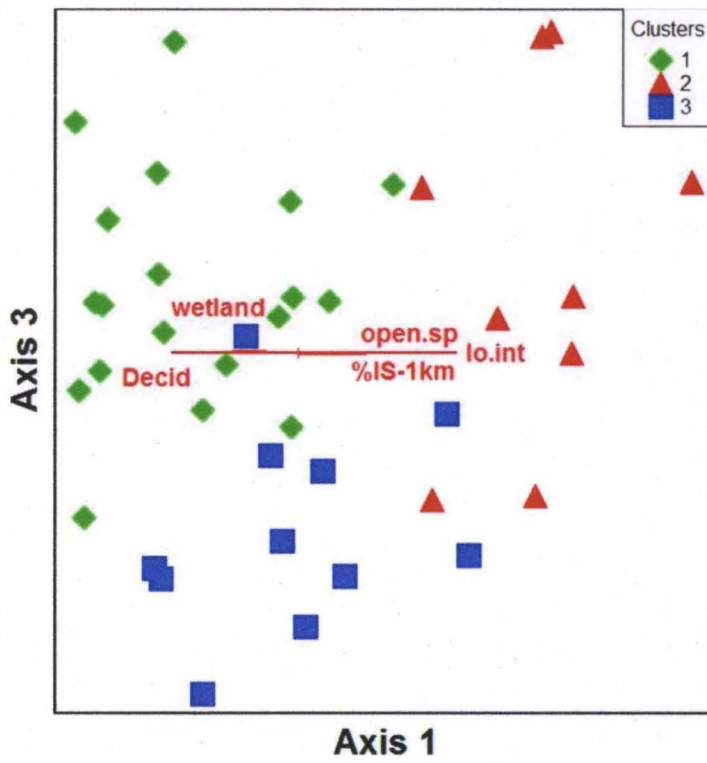


Figure 3-9: Non-metric multidimensional scaling (NMS) 3-dimensional configuration of research sites arranged by groundcover species composition. Research sites (points) assigned to their respective groupings from the 3-cluster configuration from cluster analysis. Grouped clusters are shown along axes 1 & 2.





(c)



(d)

Figure 3-10a-d: Non-metric multidimensional scaling (NMS) 3-dimensional configuration of research sites arranged by groundcover species composition and color-coded by cluster assignments from the 3-cluster configuration using cluster analysis. Cluster identities are identified in the legend. Vectors representing site environmental metrics (created using the biplot feature in PC-Ord (McCune and Mefford 1999)) are pointing in the direction of increasing proportions, with the length of each vector representing the strength of correlation. Vectors for environmental metrics are shown only if they have an $r^2 \geq 0.15$ along at least one axis. Environmental metrics shown along (a) axes 1 & 2 and (b) axes 2 & 3 are the proportion of impervious surface (%IS) surrounding each site at 3 scales: subcatchment (SC), 1-km and 500-m radius from the site center. Since %IS at the 1-km scale exhibited the strongest correlation with the site arrangement (a & b), I chose to use environmental metrics from the National Land Cover Database. Environmental metrics are shown along the same axes in (c) and (d) include low- (lo.int) and medium- intensity (med.int) developed, open-space developed (open.sp), % IS within a 1-km site radius (%IS-1km), pasture, and deciduous forest (Decid). Definitions are included in Appendix Table 3. Explained axis variance (r^2) is as follows: Axis 1 – 0.25, Axis 2 – 0.22, Axis 3 – 0.25.

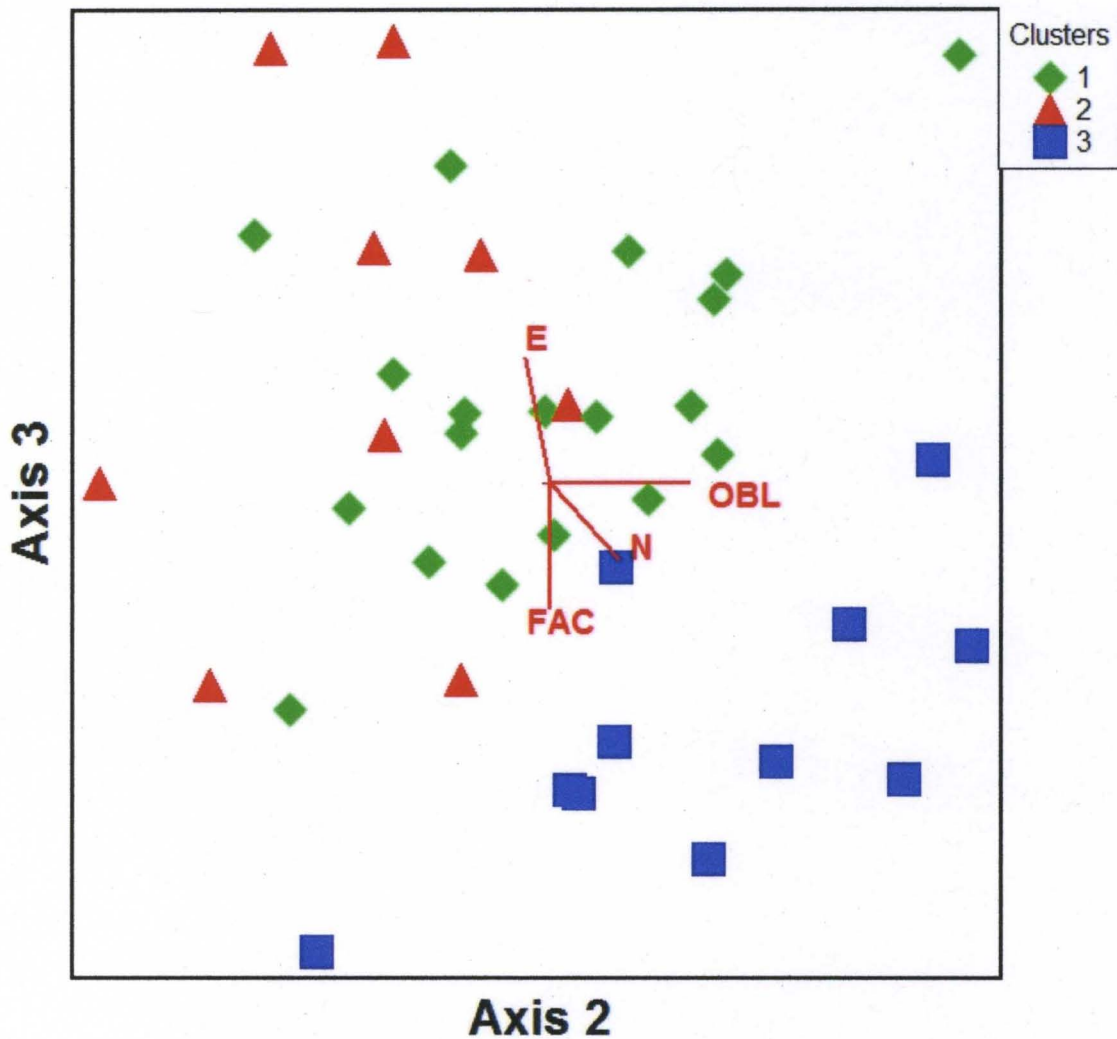
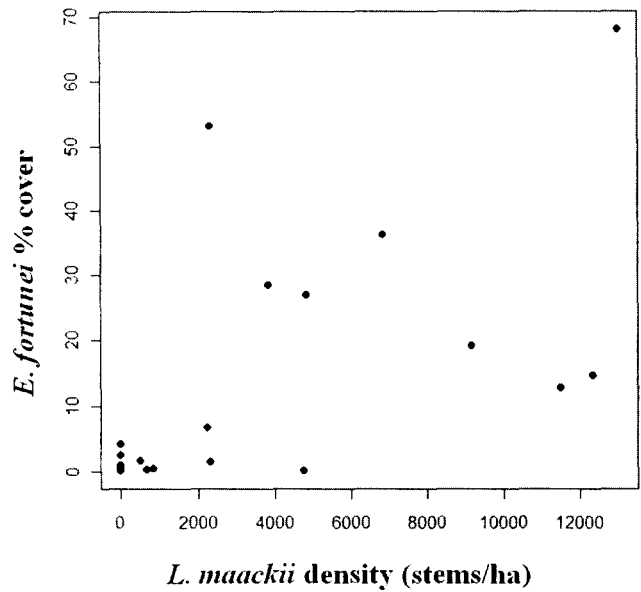
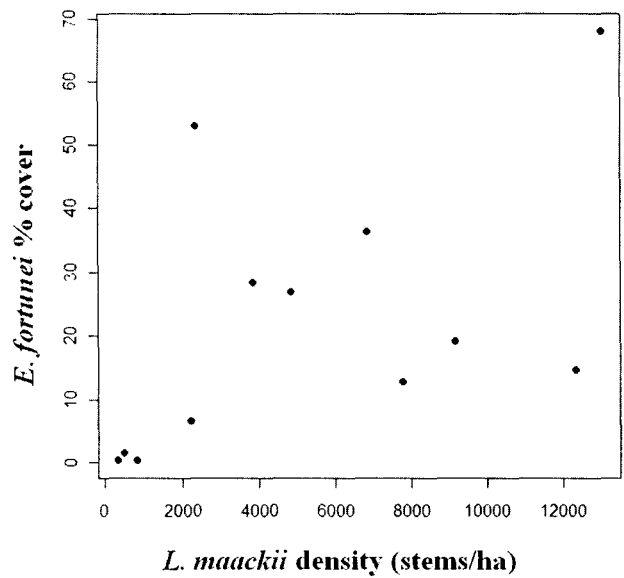


Figure 3-11: Non-metric multidimensional scaling of research sites arranged according to groundcover species composition. Sites are color coded according to their cluster assignments from the 3-cluster configuration using cluster analysis. Cluster identities are identified in the legend. Vectors representing species attributes (created using the biplot feature in PC-Ord (McCune and Mefford 1999)) are pointing in the direction of increasing importance values, with the length of each vector representing the strength of correlation. Vectors for species attributes are shown only if they have an $r^2 \geq 0.15$ along at least one axis. The species attributes shown are facultative (FAC), obligate wetland (OBL), native (N) and exotic (E) species. Definitions and the full list of attributes included in the analysis are in Appendix Table 3. As a 3-dimensional view is not an option using the biplot feature, separation of sites are shown only along axes 2 and 3 to enable better visualization of the relationship between environmental variables and cluster categories in two dimensions. Explained axis variance (r^2) is as follows: Axis 2 – 0.22, Axis 3 – 0.25.



(a)



(b)

Figure 3-12a-b: Scatterplots of *Euonymus fortunei* percent cover in response to *Lonicera maackii* density. All nearstream plots were included in the regression that contained *E. fortunei* (a), or that contained both *E. fortunei* and *L. maackii* (b). Data were square root (a) or natural log (b) transformed prior to regression to better meet statistical assumptions. One outlier was removed from graph (a), and two were removed from graph (b) following outlier analysis to better meet statistical assumptions. Pearson's r and p -values were (a) Pearson's $r = 0.73$ and $p = 0.0001$, and (b) Pearson's $r = 0.81$ and $p = 0.001$.

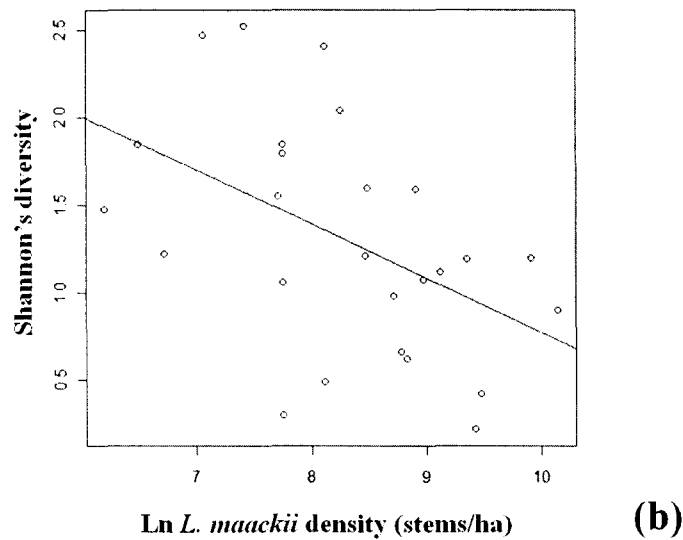
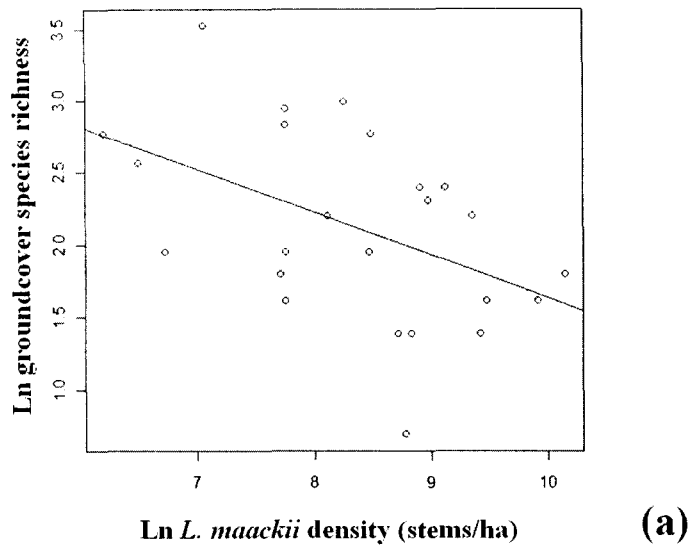


Figure 3-13a-b: Species richness (a) and Shannon's diversity (b) of groundcover species in nearstream plots in response to *Lonicera maackii* density. Species richness and *L. maackii* density was natural log transformed prior to regression in (a). Only plots that contained *L. maackii* were included. Two outliers were removed from graphic (a) following outlier analysis to better meet statistical assumptions. The adjusted coefficient's of determination and p-values were (a) $r^2 = 0.19$ and $p = 0.019$, and (b) $r^2 = 0.20$ and $p = 0.012$.

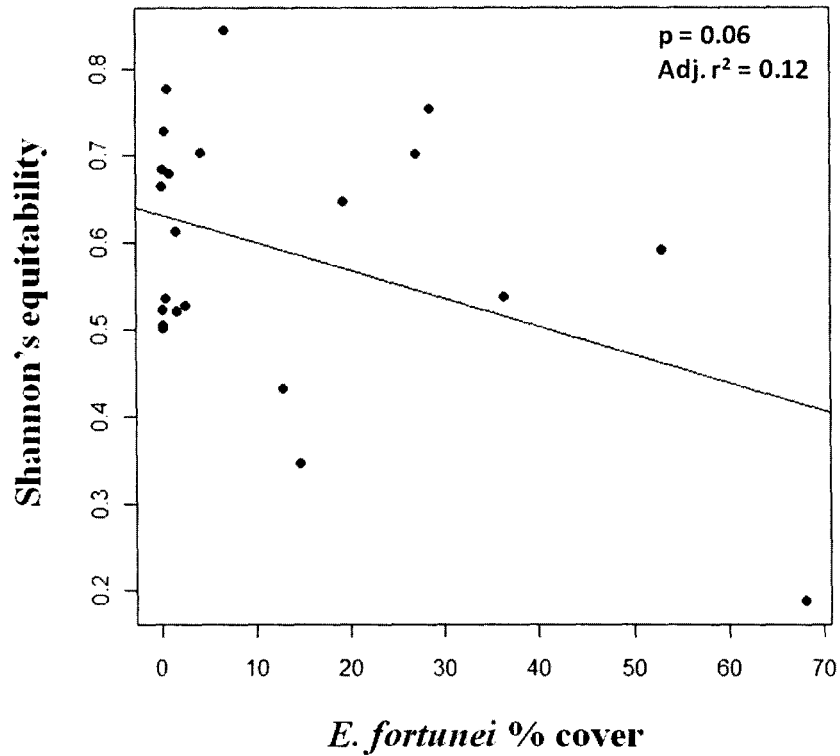
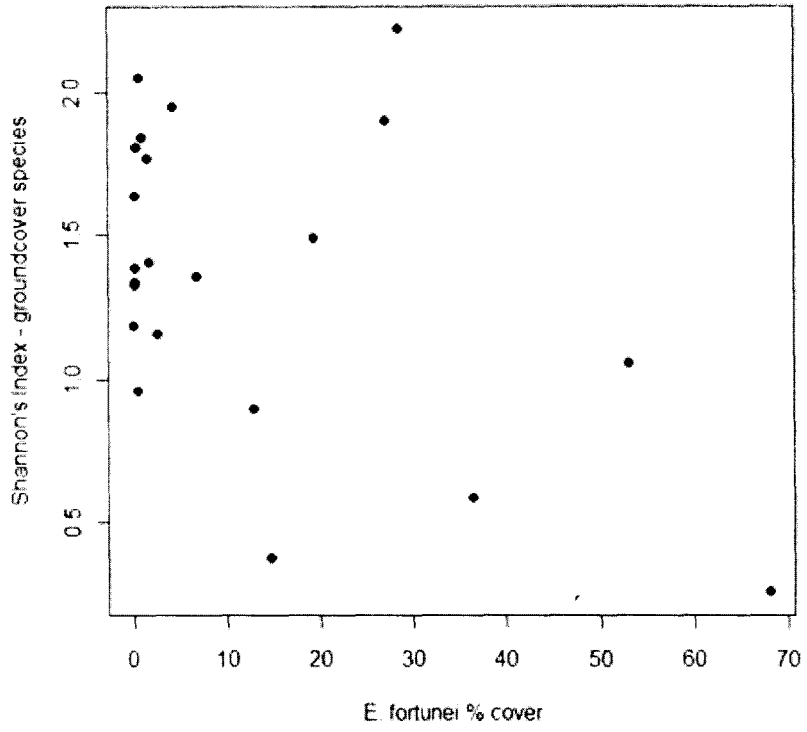
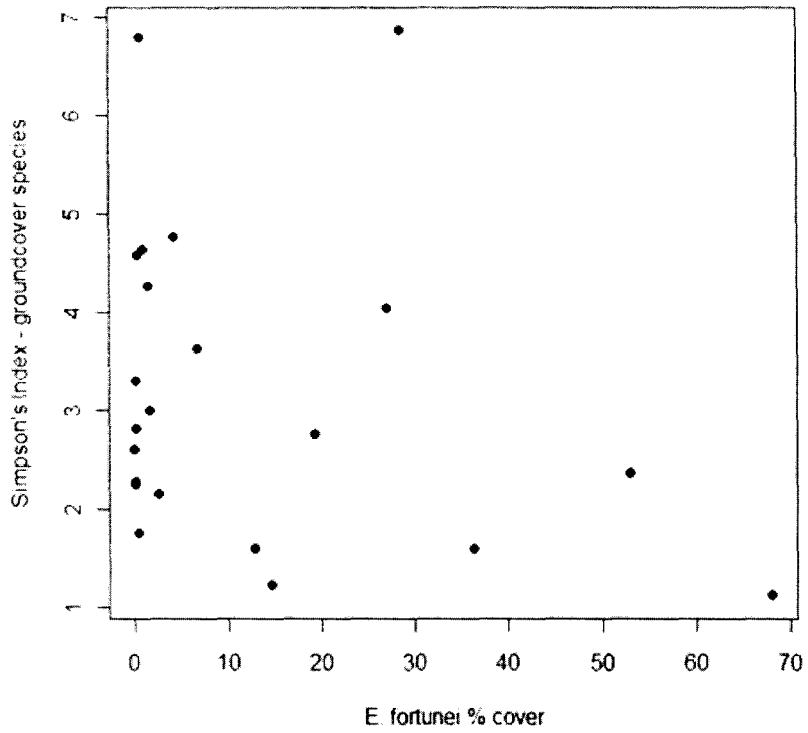


Figure 3-14: Shannon's equitability of groundcover species in nearstream plots in response to *Euonymus fortunei* percent cover. Only plots that contained *E. fortunei* were included. The adjusted coefficient of determination was $r^2 = 0.12$, and the p-value was $p = 0.06$.



(a)



(b)

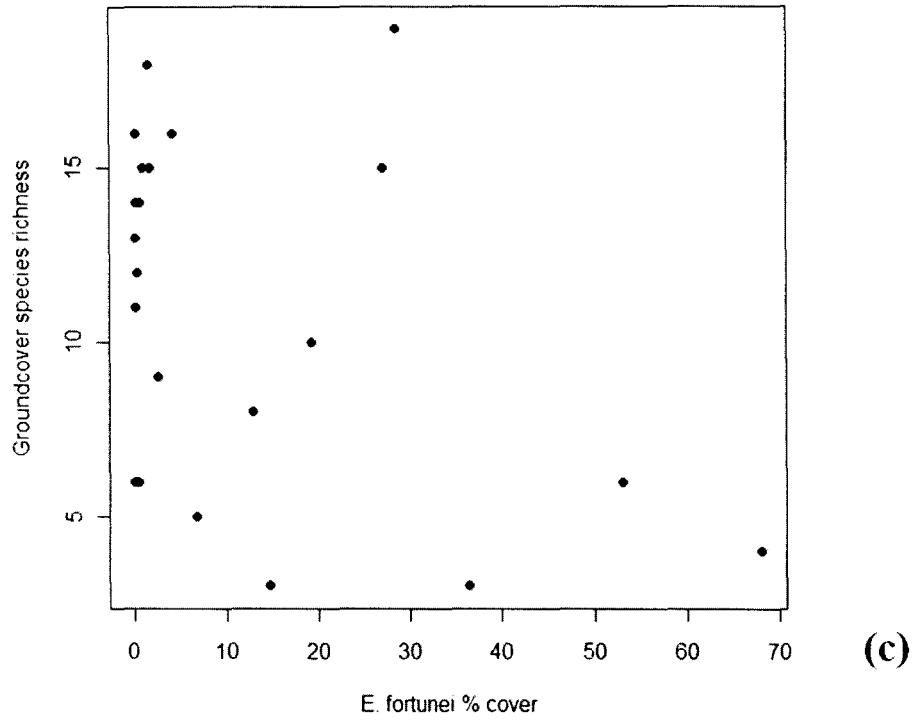


Figure 3-15a-c: Shannon's Index (a), Simpson's Index (b), and species richness (c) of groundcover species in nearstream plots in response to *Euonymus fortunei* percent cover. Only plots that contained *E. fortunei* were included.

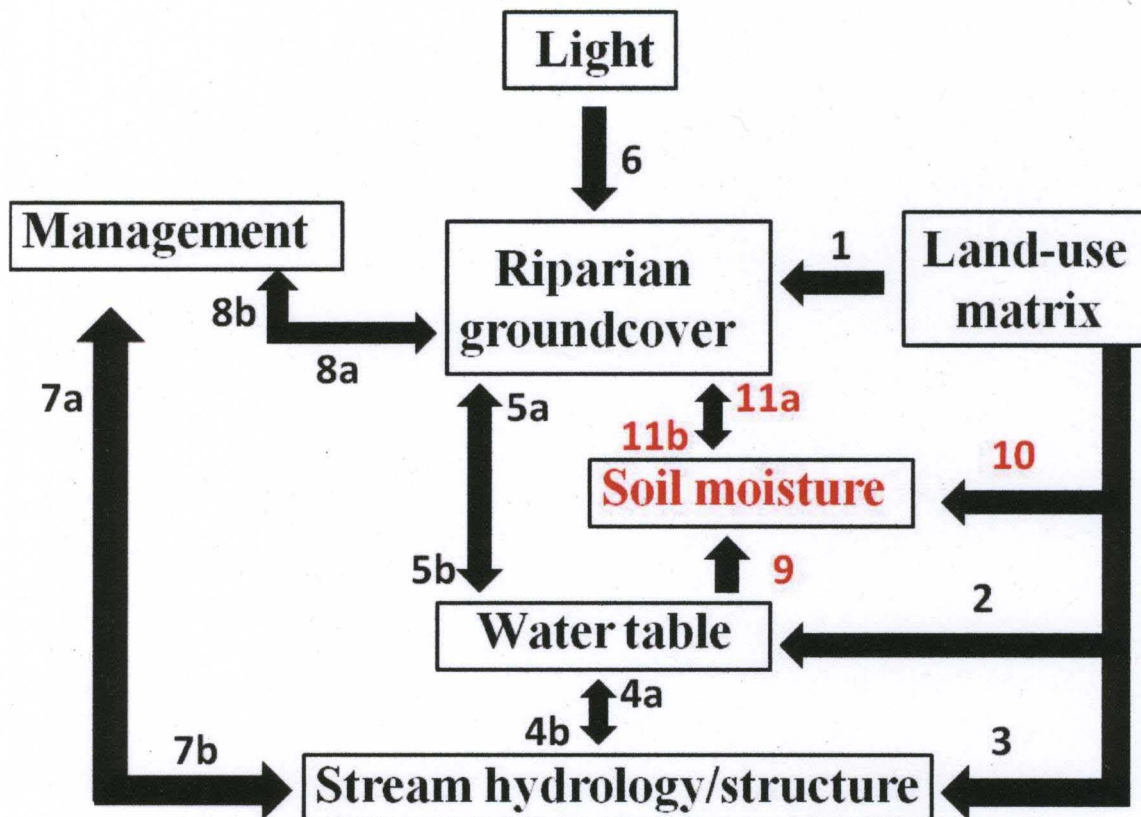


Figure 3-16: New conceptual model of variables that affect and are affected by vegetation within a riparian area modified from Figure 1. Additional arrows in need of further study are depicted in red. A direct link between groundwater levels and riparian vegetation was not found. Soil moisture was added as an intermediary to refocus attention on that link to the vegetation composition within riparian zones. The water table can still limit species composition if it is close enough to the soil surface (Arrow 5a). The water table along with surrounding land-use can also affect soil moisture (Arrow 9 & 10 respectively). Where species are not limited by saturated soil, soil moisture may play a larger role in species composition by favoring species that are better able to compete under existing moisture conditions (Arrow 11a). Riparian groundcover can feedback on soil moisture depending on proportion of the soil that is shaded by leaf cover and the transpiration rates of various plants (Arrow 11b). Descriptions of other arrows as in Figure 1-1.

CHAPTER 4: SEASONAL GREENHOUSE GAS FLUX IN RIPARIAN SOILS ALONG URBAN, SUBURBAN, AND RURAL RIPARIAN ZONES IN METRO LOUISVILLE, KENTUCKY

Introduction

Removal of nutrients before they reach the stream is a major function of the vegetation, soils and soil microbes in riparian zones (Lowrance et al. 1997, Groffman et al. 2003). In riparian zones the pathways and microbial transformations of nutrients are significantly controlled by the presence or absence of oxygen. Since the water table is closer to the surface in riparian areas than in upland areas, variation in depth to water table can affect the balance of aerobic vs. anaerobic microorganisms and gas transport from the soil to the atmosphere and *vice versa*. Under water-saturated soil conditions that occur in soils of forested riparian sites, aerobic microorganisms require elemental O₂ as the final electron acceptor during respiration and, therefore, cannot thrive under high soil moisture conditions of long duration. In contrast, anaerobically respiring microorganisms can use compounds other than O₂ for this process, and therefore flourish under low to no oxygen conditions.

Understanding how landscape level variation controls nutrient transformations within riparian zones is important, since they lie adjacent to bodies of water and filter and transform nutrients entering these aquatic systems. Riparian zones have been shown to be effective in removing sediment (Cooper et al. 1987, Lowrance et al. 1988), ammonium (NH₄⁺) (Jacobs and Gilliam 1985a, Puckett 2004), nitrogenous compounds (NO₃⁻), and P

(Cooper et al. 1987). Eutrophication of water bodies in urban and suburban areas is common and of concern to state and regulatory agencies that are responsible for water quality (Kolenbrander 1972, Gilliam 1994, Groffman et al. 2002). Nitrogen inputs to streams have been shown to occur to a greater extent from urban riparian areas than from forested reference riparian areas (Groffman et al. 2004). Studying the effects of urbanization on the soil processes that remove or supply these nutrients to water bodies is a worthwhile endeavor from both economic and water quality standpoints.

Gas effluxes (CO_2 , CH_4 , N_2 , N_2O) from soils are among the routes by which carbon and nitrogen inputs into streams are decreased. As compared to forested rural catchments, soil gas fluxes may differ within urban and suburban catchments via several mechanisms (Raich and Schlesinger 1992, Groffman et al. 2004, Groffman et al. 2006). Riparian soils along urban-to-rural gradients may differ in the quality and quantity of organic matter, inorganic nutrients, bulk density, soil moisture and depth to water table, all of which affect nutrient cycling rates and directions. In addition to variation in parent soil material, such urban-rural differences are dependent on the width of the forested buffer. Forests within ten meters of edges were found to be efficient traps for nutrients when compared to 200-m towards the forest interior (Draaijers et al. 1988). Degree of riparian colonization by different species of woody plants would also affect riparian nutrient sink functions. Species differences in growth rate, size, phenology, and morphology affects net primary productivity, which in turn affects the quantity and quality of carbon inputs (Ehrenfeld 2003) and nutrient uptake dynamics in riparian systems. For example, such changes in urban and suburban plant communities could be due to exotic plant invasion. Urban and suburban riparian areas are highly susceptible to exotic species invasions

(Burton et al. 2005, Burton and Samuelson 2008) due to proximity of exotic seed sources, management planting and disturbances, as well as altered stream hydrology. Variation in atmospheric nutrient deposition could also potentially affect nutrient cycling differentially along urban-to-rural gradients via direct inputs to soils and by affecting plant productivity and community composition. Bulk density differences can influence the anaerobic or aerobic nature of the soil. High bulk density results in more anoxic soils (Ball 1999). Stream channelization can cause more rapid losses of ground water and decreased soil moisture in urban riparian areas (Groffman et al. 2003). These factors alter soil oxygen concentrations, microbiological communities, and atmospheric-soil gaseous exchange rates.

In addition, since urban areas tend to be hotter than their rural counterparts due to the urban heat island effect. Given similar canopy coverage, riparian gas flux processes could also differ between urban and rural areas due to higher soil temperatures increasing the metabolic activity of plant roots and microbes. Higher atmospheric temperatures in urban areas can also potentially result in drier soils by accelerating evapotranspiration. Also, higher streambanks along piedmont streams coupled with lower water tables in urban areas (see Dissertation Chapter 2) (Groffman et al. 2003) exacerbate soil drying, because upper soil layers are not inundated as frequently with water. As cities expand and become more populous, understanding source-sink dynamics of trace gas fluxes in riparian areas and the soil properties that drive them will be crucial to modeling greenhouse gas accumulation within the atmosphere.

This study's first objective, therefore, was to measure the fluxes of three greenhouse gasses (CO_2 , CH_4 , N_2O) over three seasons of the year along streams located

in urban, suburban and rural catchments in and near the city of Louisville, Kentucky to determine if they varied predictably with increasing impervious surface area in these catchments. I predicted that CO₂ flux, due to aerobic respiration by roots and microbes, would be higher in urban riparian areas due to dryer soils and potentially warmer temperatures in urban than in suburban or rural riparian habitats. I also predicted that net methane uptake into soils via methanotrophy would be greatest within urban riparian sites since methanotrophs are obligate aerobes. Conversely, I predicted a net methane efflux (methanogenesis) from soils in rural forested sites, because the anoxic environment created by shallower water tables should favor methanogenic over methanotrophic activity. Predicting land-use relationships with N₂O flux is less straightforward, because it is the product of two processes, one aerobic (a by-product of nitrification), and the other anaerobic (denitrification).

The second objective was to determine if the urban, suburban and rural variation in flux rates could be related to variation in riparian characteristics expected to affect these rates. These included streambank height, depth to water table, and degree of plant community invasion by an exotic shrub, *Lonicera maackii*. I predicted that urban riparian water tables would be lower when compared to suburban and rural reference riparian sites because of higher urban streambanks, which I had demonstrated previously (see Dissertation Chapter 2). In addition, I reported in Chapter 2 (this dissertation) that urban and suburban riparian areas were densely colonized by the exotic shrub, *L. maackii*. I expected that this shrub could affect N₂O gas fluxes at some times of the year due to its highly nitrogenous litter, which mineralizes N rapidly (Trammell 2010). Litters with rapid mineralization often stimulate soil nitrification, providing substrate to support

denitrification under high moisture conditions. In addition, high soil nitrification rates would also lead to high rates of N₂O emissions under drier, aerobic conditions, as well.

The third objective was to evaluate whether greenhouse gas fluxes, particularly N₂O, could be related to the concentrations of dissolved organic carbon (DOC) and nitrate in groundwater collected at the same time as flux data. Because denitrification is dependent on nitrate (NO₃⁻) and DOC levels, I predicted higher concentrations of groundwater nitrates and DOC would be related to greater nitrous oxide flux rates from the soil under conditions of high soil moisture and depth to water table.

The fourth objective was to evaluate the influence of soil properties on greenhouse gas flux rates. I predicted % clay and soil bulk density would be negatively correlated with CH₄ uptake by the soil, since CH₄ uptake is highly dependent on CH₄ diffusion rates into the soil from the atmosphere. I also predicted fluxes of CO₂ and N₂O would follow patterns others have found with regards to bulk density (Xu and Qi 2001), with decreasing flux rates at higher bulk densities due to fewer soil pore spaces for microbial activity.

Methods

Study Area

The study area consisted of three individual watersheds located within and adjacent to Louisville, Kentucky, USA (Latitude 38°15'N, Longitude 85°46'W), which has a population of approximately 700,000 with a mean density of 695 people km⁻² (U.S. Census Bureau 2008). These were the Beargrass Creek (Middle Fork), Goose Creek, and Harrods Creek (South Fork) watersheds. The watersheds lie adjacent to each other with

all streams flowing in a primarily westerly direction and eventually emptying into the Ohio River. The three watersheds are of approximately equivalent sizes, containing varying proportions of impervious surface cover.

Beargrass Creek watershed, located in the north-northeast portion of Jefferson County, KY, is approximately 65-km² (Metropolitan Sewer District 1999b) and contains the greatest proportion of impervious surface cover (33% ISC). This Middle Fork of Beargrass Creek joins the South Fork before flowing into the Ohio River. Several stretches of this stream are featured on Kentucky's Division of Water 303d list, which list streams with impairment issues as well as the pollutant(s) causing the impairment (Kentucky Environmental and Public Protection Cabinet 2008). Since combined sewer overflows (CSOs) commonly occur in this stream, the presence of fecal coliform and sewage are stated as the primary problem pollutants within Beargrass Creek.

Goose Creek is a 50-km² watershed also in Jefferson County, in-between and adjacent to Beargrass and Harrods Creek watersheds (Metropolitan Sewer District 1999a). Of the three watersheds in this study, impervious surface cover is intermediate (20% ISC). The creek is composed of a pair of forks that meet before emptying into the Ohio River. Goose Creek is also listed on the 303d list, with cadmium and fecal coliform levels stated as impairment issues.

South Fork Harrods Creek watershed (60 km²), which has the lowest impervious surface cover of the three (10% ISC), is located primarily in neighboring Oldham County, but extends into northeast Jefferson County. South Fork Harrods Creek eventually joins North Fork Harrods Creek to become Harrods Creek before flowing into the Ohio River. Sections of the larger Harrods Creek network made the 303d list in years

past, specifically the portion passing through Jefferson County which is a more developed area. As in the other two watersheds, fecal coliform has traditionally been a problem (Metropolitan Sewer District 1999c). The current 303d list contains Harrods Creek with fecal coliform as an impairment issue, although this information is for the stream section downstream of where the North Fork and the South Fork Harrods Creek join.

Watershed classification

Watersheds were chosen from digitized catchment layers downloaded from the Louisville/Jefferson Information Consortium (LOJIC) database (LOJIC 2010). LOJIC is a multi-agency effort to maintain a geographic information systems (GIS) database to serve primarily Jefferson County, KY, although limited information is available for a portion of Oldham County, KY. After checking for topographical accuracy, I divided catchments into subcatchments based on stream order and topography from the original LOJIC layers. I further divided subcatchments of higher stream orders with longer reaches into two or three subcatchments. Data layers from the LOJIC database were clipped to each of these subcatchments. Beargrass Creek watershed contained thirteen subcatchments, Goose Creek watershed contained twelve subcatchments, and Harrods Creek watershed contained fourteen subcatchments.

Impervious land use classification was determined using three data layers from the LOJIC database digitized in 2001: buildings, miscellaneous transportation, and roads. The buildings-layer included all built structures, whether commercial or residential. The roads-layer included all paved road surfaces, excluding parking lots. The miscellaneous

transportation-layer included secondary transportation features such as driveways, parking lots, and sidewalks. The area of land covered by impervious surfaces for each subcatchment was determined using ArcGIS 9 (ESRI, 2010) and was divided by the total land area for each subcatchment to determine the proportion of impervious surfaces covering the landscape.

Categorization of subcatchments into urban, suburban, and rural land use categories was modified from Schuler (1994). For this study, urban subcatchments were defined as containing $\geq 30\%$ impervious surface cover. Rural subcatchments contained $\leq 10\%$ impervious surface cover and suburban subcatchments contained between ten and 30% impervious surface cover. Research sites were classified by land use based on impervious surface cover at three scales. The first and largest areal scale was the subcatchment level, where sites were classified as urban, suburban, or rural based on the subcatchment in which they were located. Impervious surface cover was calculated at two additional and smaller scales surrounding the sites. Site buffers were created using ArcGIS 9 at a radius of 1-km and 500-m from the site center. The proportion of impervious surface cover was determined for these buffers in the same way as for the subcatchments.

Site selection and gas chamber installation

Chosen sites were a subset of those used in previous vegetation studies (see Dissertation Chapters 2 and 3). Thirteen out of 41 sites were chosen based on bank height, land-use classification (insuring similar numbers of urban, suburban, rural catchments), and landowner permission for entry and for groundwater well installation.

Contingent upon landowner cooperation, I attempted to install a groundwater well at each site. If a well could not be installed due to excessively rocky soils, or if I encountered bedrock before groundwater, then the site was not used, with the following exception. I used one rural site with no water table above bedrock following multiple attempts at other sites and encountering excessively rocky soils before encountering groundwater or bedrock. Groundwater levels were measured in urban, suburban, and rural sites every two weeks from November 2006 to October 2008. Some of the well locations were changed in 2007 due to limited landowner permissions and to capture the variation in bank heights observed (see Dissertation Chapter 2) by land-use type, calculated at the subcatchment scale.

All wells and gas chamber bases were installed by October 2007, which was a month before any sampling took place. Two circular gas chamber bases (28.5-cm inside diameter) (Artmor Plastics, Cumberland, MD, USA) were installed per site parallel to the stream one to two meters from the streambank edge, and three meters apart from each other. PVC bases were hammered into the ground as far as possible but no more than six cm (maximum base height). Periodic measurements of chamber height were taken relative to the ground surface to ensure an accurate air volume determination before measurements were taken. I began sampling with five urban, four suburban, and four rural sites. Four urban and one suburban site were located along Beargrass Creek; one urban, three suburban, and two rural sites along Goose Creek; and one suburban and two rural sites along Harrods Creek. In February 2008, two gas chambers were taken from a suburban site within a state park along Goose Creek; therefore, I abandoned that site. I chose an additional urban site along a first order tributary of Beargrass Creek in July

2008. The site was located in a remnant woodland area, with the entire riparian zone (\leq 12-m wide throughout) upstream of and at the site being forested.

Groundwater wells

Nearstream water table depth was determined over a two-year period at these 13 sites by sampling piezometer wells. One piezometer well was installed within 0.5- to 1-meter of the streambank edge in each of six urban, three suburban, and four rural riparian sites. The shafts of the piezometer wells were constructed of poly-vinyl chloride (PVC) pipe with a 1.25-inch inner diameter to allow for insertion of a measuring tape for determination of groundwater depth from the ground surface. The base of each piezometer was constructed of slotted PVC pipe approximately 300-mm in length. When measurements were begun at the end of 2006, piezometer bases used for measurement were commercially manufactured using three-quarter inch PVC pipe. Throughout 2007 some of those were replaced with self-constructed bases measuring 3.2-cm in diameter to allow enough water to collect inside the well for sampling. Plastic PVC caps were placed at the bottom and top of the well to prevent water and debris from entering the well from above and also to allow water to drain from the well had the water table dropped below the well, as a hole was drilled into the bottom cap. In both the commercial and self-constructed bases, a mesh filter covered the slots to limit soil and debris entering the well. The piezometer base was attached to the shaft via a PVC collar. PVC cement was used sparingly to attach the collar and bottom cap to minimize potential contamination.

To place the piezometer into the ground I used a 10.2-cm diameter auger to drill a hole large enough in diameter to prevent the well from contacting the sides upon

placement. I also allowed the top portion of the piezometer to protrude at least 30-cm above-ground to prevent floodwaters from entering the well. Once in place, sand was poured to cover the base of the piezometer to allow for drainage and prevent clogging of the filter surrounding the base. A layer of bentonite clay pellets approximately 15.2-cm deep was poured on top of the sand layer. Water was poured over the pellets to allow for expansion, and native soil was used as the remainder of the fill. Depth to groundwater measurements in the well were made approximately every two weeks from November 2006 to November 2007 using a tape measure coated with Keson® ultra-fine marking chalk. The remainder of the measurements up to October 2008 followed the frequency of soil gas sampling described below.

Soil and site properties

Soil properties determined at each site visit included soil temperature and gravimetric soil moisture. Soil temperature was determined using a long-stem thermometer, and was measured 50-cm from each chamber during each site visit. Measurements were taken at 5-, 10-, and 15-cm depths to obtain a representative average through the soil column. Air temperature was measured between the two chambers. Gravimetric soil moisture was determined from a 2-cm diameter soil core taken one to 1.2 meters from each chamber, so as not to disturb the soil in and closer to the chamber. Two soil cores for each chamber were obtained to a depth of 15-cm each site visit. Soils were enclosed in plastic bags until later that same day when they were weighed and then dried in an oven at 105°C for up to 48 hours. Gravimetric soil moisture content was determined using the following formula:

Eq. 1

$$\text{Gravimetric soil moisture (\%)} = 100 * (\text{Soil wet wt. (g)} - \text{Soil dry wt. (g)}) / \text{Soil dry wt. (g)}$$

Additional soil characteristics were determined from a single soil sample taken inside the chambers in the first half of October 2009 after the last gas samples for the study were collected. Soil cores (10.2-cm diameter and 13-cm long) were taken from the middle of each trace gas chamber using a soil impact corer with polypropylene sleeves. Stones and roots were removed and soils dried at 105°C for at least 48 hours. Bulk density was determined for each core using the following formula:

Eq. 2

$$\text{Bulk density (g/cm}^3\text{)} = \text{Dry soil weight (g)} / \text{Soil core volume (cm}^3\text{)}$$

Soil C:N molar ratios for each chamber were determined via combustion in the EAL at the University of Louisville using a Perkin-Elmer Series II 2400 (CHNS/O) analyzer.

Additional soil properties (nutrients and texture) were determined using soil samples pooled from both chambers at each site. Soils were well mixed, and passed through a 2-mm sieve prior to analysis. Soils were analyzed at Brookside Laboratories (308 S. Main Street, New Knoxville, OH, USA; www.blinc.com) for texture (ASTM Standard D422-63 2002) and trace minerals (Bray and Kurtz 1945, Lindsay and Norvell 1978, Mehlich 1984), the full list of which can be found in Appendix 13.

Vegetation surrounding each chamber was quantified to use as correlates that could potentially explain some of the variation in gas flux. Percent bare ground and

herbaceous cover, and shrub and sapling counts were assessed within a 2-m radius of each chamber. Percent cover for herbs and bare ground was assessed using a seven category cover class system (Category # = % cover; 1=<1; 2=1-5; 3=6-25; 4=26-50; 5=51-75; 6=76-95; 7=>95) with the average midpoints reported. Tree counts were determined within five meter radius of each chamber. Counts are reported as the total within a 12.6-m² area for shrubs and saplings and within a 78.5-m² area for trees. Tree canopy cover within a 5-m radius of the chambers was assessed using a spherical densiometer (Lemmon 1956).

Sampling Well Water for Dissolved Organic Carbon and Nitrate+Nitrite-N

DOC samples were collected once per month from November 2007 to October 2008, except during June 2008 when two collections were made. Samples for determining nitrate+nitrite-N concentration (mg/L) were also collected once per month from November 2007 to February 2008, and in August and October 2008. However, the collection frequency was doubled from March to July and in September 2008 to follow the gas sampling frequency described below. Prior to well water collection, wells were emptied of water using a peristaltic pump and Nalgene tubing and allowed to fill for at least 30 minutes prior to sampling. This was done to obtain a recent water sample and also to rinse the inside of the tubing. Water samples were collected from wells into 100-ml acid-washed Nalgene bottles. Samples were stored on ice until returned to the lab. Samples were then vacuum-filtered through 0.45- μ m filters (glass fiber, type A/E). Forty ml aliquots were decanted into sterile glass bottles for determination of dissolved organic carbon (DOC) concentrations. Ten drops of 4N HCl were added to the DOC sample to

stop microbial activity, and samples were stored at 4°C. The remainder of the sample was decanted into a sterile 40-ml Falcon tube and either stored at 4°C or frozen at -20°C for nitrate+nitrite-N determination.

DOC concentrations were determined via combustion by the Environmental Analysis Laboratory (EAL) at the University of Louisville using a Shimadzu TOC Model 5050A analyzer. Water samples were assessed for nitrate+nitrite-N within seven to ten days following collection. Colorimetric analysis was used following a modified cadmium reduction method from Hach, Inc. (2004a, b). Two different methods were used depending on sample concentration. The low-range method was used for samples containing from 0- to 0.5-mg/L nitrate+nitrite-N. The high-range method was used for samples containing from 0.5- to 30-mg/L nitrate+nitrite-N. These methods involve reduction of nitrate to nitrite, followed by conversion to a diazonium salt in the presence of sulfanilic acid. The salt couples with either gentisic acid (high-range) or chromotropic acid (low-range) for color formation. Samples were divided into 5- (for high-range) or 15-ml (for low-range) aliquots. All samples were warmed to 25°C in a water bath prior to analysis. First, for the low-range procedure, the contents of a NitraVer 6 Reagent Powder Pillow were added to the sample, followed by vigorous shaking for 3-min. A 2-min reaction period followed. Ten mls of the sample was decanted into a separate container, followed by addition of a NitraVer 3 Nitrite Reagent Powder Pillow and repeated inversion for 30-sec. The sample was allowed to incubate for 15-min. One ml of sample was analyzed in a UV/Vis spectrophotometer with a 1-cm path length at 507-nm, with the resulting reading in mg nitrate+nitrite-N/L. Samples found to contain a greater than 0.5-mg nitrate+nitrite-N /L were subjected to the high-range test. The NitraVer 5 Reagent

was added to the 5-ml sample, followed by vigorous agitation for 1-min. A 5-min. reaction period followed. One ml of the sample was read using the same UV/Vis spectrophotometer with a 1-cm path length at a wavelength of 500-nm.

Measuring net CO₂, net CH₄ and net N₂O fluxes from riparian soils

Soil respiration and CH₄ flux was measured from January to October 2008. Measurements were taken once a month in January, February, August and October. Sampling occurred at two-week intervals from March to July and in September. These trace gas fluxes were measured using closed chambers (purchased from Artmor Plastics Corporation, Cumberland, Maryland, USA) each consisting of a cylindrical lid and base constructed of polyvinyl chloride (PVC). A rubber gasket on the lid formed a seal with the base when in place. The lid had an inner volume of 3.1125-L and the area of the base measured 638-cm². Bases were permanently installed in the ground in October 2007, and volume varied depending on depth of installation, which was measured for each chamber to determine the total air volume of each. During sampling, the chamber lids were placed on the base and fine-needle 50-ml polypropylene syringes were used to collect 20- to 30-ml gas samples from sampling ports located in the center of the chamber lids. Collection times occurred at 0, 10, 20, and 30 minutes after placement of the chamber lid on the base. Syringes were closed via stopcocks and samples were processed the same or next day. An SRI 8610 gas chromatograph (SRI, Redondo Beach, CA, USA) equipped with a thermal conductivity (CO₂) and flame ionization (CH₄) detector and a porapak Q column was used to process the samples. Gasses used for standardization of chromatogram peaks were 360 ppm CO₂ and 10 ppm CH₄. Helium was used as the carrier gas. These trace gas

fluxes were calculated from the linear rate of change in gas concentration multiplied by the internal volume of the chamber and normalized on a m^2 soil surface basis, as follows:

Eq. 3

$$\text{Flux rate} = ((V_{\text{lid}} + V_{\text{base}}) / (R * T)) * (\Delta \text{conc} / \text{hour}) * (C_{\text{am}}) * (24 \text{hours} / \text{day}) / (A_{\text{base}} - \text{m}^2)$$

where V_{lid} is the volume of the chamber lid (3.1125 L), V_{base} is the volume of the installed chamber base (L), R is the ideal gas constant (0.0821 $\text{L} * \text{atm} / (\text{mol} * \text{K})$), T is chamber air temperature (K), Δconc is the change in concentration of CO_2 or CH_4 ($\text{ppm} / 1000 \text{mg} / \text{g} * \text{hour}$), C_{am} is the atomic mass of carbon (12-g/mol), and $A_{\text{base}} - \text{m}^2$ is the area of the chamber base in square meters (0.0638- m^2). The unit for flux rate is $\text{mg CH}_4\text{-C}$ or $\text{CO}_2\text{-C} / \text{m}^2 * \text{d}$.

Samples were assessed for N_2O concentrations at a separate facility (University of New Hampshire Water Resources Research Center, Dr. William McDowell, Jeffrey Merriam), since there was no operational electron capture detector on the SRI 8610 gas chromatograph. Samples were collected once in May, June, July, and August 2008. Samples were collected in the same way as CO_2 and CH_4 , and immediately transferred to 10-ml evacuated glass vials and sealed with rubber septa for storage. Silicon grease was used to reduce leakage from the vials during storage and shipment. Samples were shipped approximately seven to nine days following collection. Five to ten ml of the air sample was injected into a Hewlett-Packard 5890 Series II (1-ml sample loop) equipped with an electron capture detector. Vials with N_2O gas controls (500 ppm) were shipped along with the samples to assess the probability of leakage from vials during shipment. The

decision was made not to include data from the May and June sampling dates since it was determined that leakage occurred in many samples.

Data Analysis

Principal components analysis (PC-Ord v. 4.41) (McCune and Mefford 1999) and multiple regression analysis (R v. 2.11) (R Development Core Team 2009) were used to assess whether monthly and seasonal differences in CH₄ and CO₂ fluxes were related to land use and site variables. Principal components analysis was used to assess whether there were patterns in CH₄ and CO₂ fluxes that could be related to site variables (%IS, woody species stem counts, % canopy cover, catchment area, bank height) and soil variables (% carbon and nitrogen, molar C:N ratios, texture, bulk density, % organic matter). Two sites were removed from these analyses, because one site was determined to be an outlier, as it occurred by itself and caused the other sites to cluster together in a separate corner of the PCA output. The other site was removed because flux data were only collected from July through October for that site. However, these sites were used in subsequent analyses if they did not contribute to heteroscedasticity.

Multiple regression was used to identify distinct soil properties that might affect seasonal differences in site fluxes. Seasons used were winter (January, February, and March), spring (April & May), summer (June-August), and fall (September & October). Monthly fluxes were averaged for each site by season and simple regressions were used to choose the strongest two or three main effects to use in each multiple regression model. Land-use differences in depth to groundwater were assessed using repeated-measures ANOVA.

Results

Site and soil properties

Soil temperature followed air temperature, steadily increasing from freezing and single digit temperatures in January and February and peaking in late August and early September 2008. Increases in soil temperature lagged behind air temperature during the year, with soil temperature rising to between 22 and 23°C, and air temperature rising to 26 to 28°C (Fig. 4-1). No differences in air or soil temperature among land-use categories (urban, suburban, rural - subcatchment scale) were observed. Mean monthly soil moisture was highest in spring and summer and declined from February to October in sites across all land-uses (Table 4-1, Fig. 4-2). Soil moisture was consistently lower throughout the year in urban soils than in either suburban or rural soils (Table 4-1, Fig. 4-2). However, small incremental increases in soil moisture generally followed rain events among all land-use categories. Vegetation coverage and density near the chambers varied between sites and land-use categories (Table 4-2). Urban sites had the highest mean proportion of bare ground within two meters of each chamber. Two suburban sites contained the highest number of trees.

Groundwater

Both data sets (2006 to 2007, and 2007 to 2008) showed similar patterns regarding bank height and depth to groundwater, with the mean water table depth becoming lower as bank height increased (Fig. 4-3). The slopes of the regression lines for mean water table depth were also similar between years, indicating chronically lower water tables with higher banks. Sites were chosen in part to represent the range of bank

heights found. As such, only the highest urban bank did not have a similarly paired rural reference (Fig. 4-3b), since none of the rural streambanks I measured in a previous study (see Dissertation Chapter 2) were as high as urban banks. Urban groundwater was also deeper than in either suburban or rural riparian zones due to the higher banks in urban areas (Fig. 4-4). However, no significant differences were detected between urban, suburban, and rural depths to groundwater ($p = 0.44$). Depths did, however, significantly change over time ($p = <0.0001$), increasing from winter to spring, and decreasing continuously into fall. Groundwater at all sites peaked in March 2008 with a heavy monthly rainfall approaching 46-cm. Depth to groundwater levels decreased gradually from the March peak until October. A small rain pulse in May helped maintain groundwater levels during that month. The scant rainfall that occurred in subsequent months in 2008 was not sufficient to prevent groundwater levels from dropping. Greater recharge of rural groundwater levels occurred during spring rains with average groundwater levels rising 60-cm from November 2007 levels. In contrast, urban groundwater levels rose by 20-cm during the same time period, while suburban groundwater levels were intermediate, rising by approximately 40-cm.

Groundwater nitrate+nitrite-N and DOC

Over the 11 month sampling period (Table 4-3), nitrate+nitrite-N concentrations in groundwater were higher in urban (Mean = 2.8-mg/L; SE = 0.8) and rural (Mean = 2.6-mg/L; SE = 0.9) plots than in suburban plots (Mean = 0.09-mg/L; SE = 0.02). The highest nitrate+nitrite-N values in urban areas occurred from January through June 2008, often being at least two orders of magnitude above suburban and rural values during this

period. In contrast, the highest values in rural areas occurred from May through October, with values equal to those in urban areas from May to July, and exceeding urban values in August through October. Suburban nitrate+nitrite-N values remained relatively low and stable over the year-long period, rarely exceeding 0.1-mg/L. As far as could be determined, rainfall did not vary appreciably within the area by land-use type (Table 4-3), and therefore could not explain the order of magnitude differences in nitrate+nitrite-N concentration among land uses.

Within a land-use type, however, nitrate+nitrite concentrations followed average rainfall and groundwater levels. Urban mean nitrate+nitrite-N concentrations and variation were highest during periods of the greatest rainfall and highest groundwater levels in March (Fig. 4-5). Concentrations gradually diminished, falling below 1-mg/L in July. Rural groundwater levels did not start to increase until the relatively wet month of May, and levels peaked following a September pulse of rain. Suburban nitrate+nitrite-N concentrations in groundwater consistently remained below 1-mg/L for the entire sampling period despite variable precipitation inputs.

Dissolved organic carbon (DOC) showed a more stable trend across land cover categories (Table 4-4), usually ranging between 4- and 7-mg/L. The highest values across land-use categories occurred during winter and summer months. DOC concentrations in urban areas peaked in January and June 2008, with values between 8- and 9-mg/L. Mean suburban values were highest in November 2007 and June 2008, while mean rural values were highest in November 2007 and July 2008.

Carbon dioxide flux

Carbon dioxide flux varied about 5-fold from January through October 2008 (Fig. 4-6), generally following changes in soil temperature (Fig. 4-1b). PCA indicated that annual mean CO₂ flux was related positively with increasing % impervious surface around the sites from the 0.5 km to the subcatchment scales (Fig. 4-7). Differences among sites grouped by land use were first distinguishable in June, with mean annual fluxes being consistently highest in urban sites from June through October (Appendix 14). A multiple regression model was used to determine the extent to which impervious surface, calculated at three scales around each site (0.5- and 1-km radii and at the subcatchment scale), and upstream cumulative catchment area could explain the variation in CO₂ fluxes among sites. I grouped the data by season due to the effects of seasonal temperatures on gas fluxes. Cumulative catchment area (ha) upstream of the sites ($p = 0.002$) and % impervious surface within 1-km of the sites (as opposed to the subcatchment scale; $p = 0.002$) explained most of the variation in CO₂ flux during summer, with higher %IS resulting in greater flux rates and increasing catchment area resulting in lower flux rates (two-factor model adjusted $r^2 = 0.72$, $p = 0.001$; Table 4-5). One outlier with the highest IS within a 1-km radius (47%) and mid-range CO₂ flux rate was left out of the analysis to better meet assumptions of homoscedasticity. This site was located in a small city park (Browns Park) along Middle Fork Beargrass Creek, which may have served to reduce the otherwise positive relationships associated with impervious surface on CO₂ flux. Fall CO₂ flux rates (Table 4-5) were related to %IS and catchment area in the same way as during summer months, increasing with %IS ($p = 0.003$) and decreasing with increasing cumulative catchment area ($p = 0.01$).

Multiple regressions revealed relationships between CO₂ flux and plot-level environmental variables that varied by season. Winter fluxes (mean flux rates for January, February, and March) positively correlated with % soil organic matter (%SOM; $p = 0.004$, adj. $r^2 = 0.58$; Fig. 4-8, Table 4-6). One rural site was not used in this analysis due to its negative CO₂ flux value, making it an outlier among the sites. It was the only site with no groundwater above bedrock. Spring (April and May mean flux rates) flux differences (natural log transformed) among sites were found to increase with increasing soil bulk density (Fig. 4-9, Table 4-6), indicating a weak trend among spring CO₂ fluxes and within the range of bulk density, which was relatively low. More proximate variables related to CO₂ flux in the summer were soil temperature ($p = 0.01$) and soil % organic matter (OM) ($p=0.0496$). Together they explained 43% of the variation in CO₂ flux rates among sites (Table 4-6). Fall CO₂ flux rates also increased with soil temperature ($p = 0.004$, adj. $r^2 = 0.55$; Table 4-6, Fig. 4-10). The same outlier was removed from the regression model for the fall season as for the summer season. No correlations were found between seasonal CO₂ flux and either CH₄ or N₂O flux.

Methane flux

Methane uptake into the soil (methanotrophy) occurred in suburban and urban riparian zones from January to October 2008. Net CH₄ emissions from the soil (methanogenesis) primarily occurred in rural riparian zones from January to July 2008. Net methanotrophy occurred in all riparian zones regardless of surrounding land use from July to October 2008 (Fig. 4-11). The range of flux values varied widely among rural sites. Net efflux usually occurred at a single rural site (RG7), with a second rural site

(RH13) exhibiting net efflux on four of six sampling dates from April through June. These sites were responsible for the large rural peak in March and April. RG7 was typically an outlier because of frequent and sometimes substantial methane release from the soil. Net methane uptake occurred at the other two rural sites on all dates sampled. Net efflux also occurred within some suburban and urban sites, but most sites exhibited net methane uptake on most sampling dates (Appendix 15).

A multiple regression model was used to determine the extent to which impervious surface, calculated at three scales around each site (0.5- and 1-km radii and at the subcatchment scale), and upstream cumulative catchment area could explain the variation in CH₄ fluxes among sites. I grouped the data by season due to the effect of temperature on microbial activity and because PCA revealed a methane flux gradient among sites that appeared strongest during fall months (Fig. 4-12 & 4-13). At the landscape scale, soil uptake of methane increased with increasing subcatchment %IS (as opposed to within a 1-km radius) during summer months (Table 4-5, Fig. 4-14). Methane uptake rates at sites with less than 15% IS were between zero and 0.5-mg CH₄-C/m²*d. Both sites that exhibited net methanogenesis contained less than 15% IS in the surrounding subcatchment. Conversely, six of eight sites with greater than 15% IS had net methane uptake rates greater than 0.5-mg CH₄-C/m²*d. No significant correlations were found between spring and fall methane flux and landscape-scale variables (Table 4-5).

Multiple regression analysis revealed relationships between CH₄ flux and plot-level environmental variables. Differences in spring methane fluxes were most strongly correlated with soil temperature, as cooler temperatures resulted in greater methane

uptake from the atmosphere (Fig. 4-15). Summer flux differences across sites were most strongly correlated with soil % carbon (Fig. 4-16). Methane uptake decreased linearly as soil % carbon increased from 2 to 7%. Methane efflux (methanogenesis) became the dominant CH₄-related process at the highest soil moisture values (> 0.4-g water/g dry mass soil) during the summer months (Fig. 4-17). Fall fluxes were found to significantly correlate with soil C:N ratios and soil moisture (Table 4-6, Fig. 4-18). For the overall multiple regression model, the adjusted coefficient of determination was 0.75 and the p-value was 0.0004.

N₂O flux

Nitrous oxide (N₂O) fluxes showed a high degree of variability both among and within sites from August to October 2008, so no land-use level trends could be discerned. August and October mean flux rates (mg N₂O-N/m²*d) were 0.17 and 0.10 in rural, 0.17 and 0.12 in suburban, and 0.16 and 0.14 in urban riparian zones. Site fluxes decreased dramatically at some sites (RH13, SG13, and UB3) from August/September to October, while fluxes at other sites decreased slightly, remained stable or increased over that interval (Fig. 4-19). Flux rates decreased by at least one-half at four sites and by one-quarter to one-third at three other sites from August to October 2008. Rates increased by one-quarter to one-half at four other sites. No significant correlations were detected between N₂O flux and site variables.

Discussion

Overall, this project adds to the growing body of literature focused on riparian ecosystem functioning in urban areas. Urban riparian functioning has thus far been

concentrated on nutrient and pollutant removal (Stander and Ehrenfeld 2010). My study contributes not only to this area, but also to understanding the dynamics of groundwater levels, groundwater DOC and nitrate+nitrite-N, and greenhouse gas in riparian zones in urban and urbanizing catchments.

Groundwater levels

As expected annual changes in depth to groundwater in these riparian sites reflected rainfall inputs, with rain pulses resulting in higher water tables. Water table levels were highest in rural sites during periods of high rainfall. Urban riparian water levels also rose after rainfall, but not to the same levels as rural water tables. These differences in groundwater recharge between urban, suburban and rural riparian zones are likely a direct consequence of urban features, such as impervious surfaces and drainage structures. These features create a barrier to groundwater flow after precipitation events and allow water to bypass exposed soil by funneling it into drainage features that empty into streams. They also indirectly affect depth to groundwater by causing stream channelization and elevated stream banks, which is part of the urban stream syndrome (Walsh et al. 2005).

Lower water tables were observed among sites with higher streambanks, regardless of land-use in the catchment. However, the highest streambanks and the lowest average water tables occurred in urban areas, which was consistent with my prediction. High banks and lower groundwater recharge rates can create what Groffman et al. (2003) termed “urban hydrological drought,” whereby the upper soil layers become dryer, changing soil microbiological processes, including those involved in greenhouse gas flux.

Water table data coupled with soil moisture data (Table 4-1, Fig. 4-2) indicated that urban hydrological drought does exist in riparian zones within the Louisville Metro area, since urban riparian soils were generally dryer than those in suburban and rural riparian soils.

Groundwater nitrate+nitrite-N and DOC

Nitrate+nitrite-N occurred in greater concentrations in urban and rural plots, peaking at different times of the year. The highest nitrate+nitrite-N concentrations (>0.5-mg/L) generally occurred at a single urban site (UB3) and a single rural site (RG7). Both sites were located adjacent to housing developments with a house within 50 m of the sites, and both were at the end of a cul-de-sac. Since nitrate levels peaked coinciding with rainfall events, runoff from fertilized lawns could have contributed to the high levels of nitrate+nitrite-N within the groundwater at these two sites.

Other studies have found that riparian vegetation dynamics influence soil nitrogen dynamics and groundwater concentrations of nitrate. Nearstream groundwater NO_3^- concentrations were found to increase four-fold with uprooted canopy tree disturbance in a mountainous region of North Carolina, although elimination of shrubs at other sites had no significant effect (Yeakley et al. 2003). This suggested that canopy trees, especially at the root-soil interface, were primarily responsible for control of nitrates in groundwater. Exotic species invasions have variable effects on soil nitrate concentrations, with effects primarily expressed on N mineralization and nitrification rates, both increasing with plant species invasions (Ehrenfeld 2003). My suburban sites had the greatest number of trees (> 1-m high, \geq 2.54-cm diameter at breast height) within a 78.5-m² area surrounding the groundwater wells and the lowest groundwater nitrate concentrations. The absence of

trees at urban and rural sites could have contributed to the relatively higher nitrate+nitrite-N concentrations. In addition, the two urban sites with the highest groundwater nitrate+nitrite-N concentrations (> 1-mg/L) from January to June 2008 (UB3 and UB5) also had the greatest densities of the exotic shrub *L. maackii* (see Appendix 9 and Dissertation Chapter 2 for sampling methods). However, the rural site (RG7) with the highest groundwater nitrate+nitrite-N concentrations (> 1-mg/L) from May to October 2008 did not contain any exotic shrubs (see Appendix 9 and Dissertation Chapter 2 for sampling methods). These conflicting findings make drawing inferences regarding increased groundwater nitrate+nitrite-N concentrations and exotic shrub invasions more complicated.

The only discernable pattern regarding nearstream groundwater DOC concentrations were related to seasonal peaks. Table 4-4 shows winter and summer peaks among all land-use types in 2008. Average rural DOC seasonal peaks were the highest overall, reaching 9.89-mg/L in November and 10.51-mg/L in July. Suburban areas had the lowest average seasonal peaks, with concentrations between 7- and 7.5-mg/L. Urban areas had seasonal peaks between 8- and 9-mg/L. Summer DOC peaks were observed in a coastal ecosystem in Sweden, with the increases attributed to low phosphate concentrations inhibiting breakdown of DOC by limiting microbial activity (Zweifel et al. 1995). Other possible explanations for the summer and winter peaks include root exudates and leachable organic matter from the litter layer (Wright and Coleman 2002, Yeakley et al. 2003), which have both been found to increase soil carbon concentrations.

CO₂ flux

At the landscape scale, % impervious surface within a 1-km site radius was positively correlated with CO₂ flux rates during both summer and fall (Table 4-5), supporting my prediction. Cumulative catchment area, another landscape scale variable, was negatively correlated with CO₂ flux rates (Table 4-5), with decreasing flux rates occurring with increasing catchment area (ha). Explanations for this trend are less easily made. However, Jones and Mulholland (1998) found lower in-stream partial pressures of CO₂ with downstream distance in a stream system in the Great Smoky Mountains National Park, which they attributed to the declining influence of groundwater on stream chemistry from upstream to downstream. Similar mechanisms may be responsible for this decline in CO₂ in both situations, and further study may be warranted if similar findings can be replicated in other systems.

Carbon dioxide flux demonstrated seasonal variations related to plot-level environmental variation as well. As expected, winter CO₂ flux was the lowest because of cold temperatures, while CO₂ flux and soil temps peaked in mid-summer (Fig. 4-1b & 4-6). Many of the soils were frozen early in the season (January), and began to thaw later in the season (March). Summer and fall CO₂ flux rate differences among sites were positively correlated with variation among sites in soil temperature (Table 4-6, Fig. 4-10). This correlation between soil CO₂ fluxes with temperature is expected and consistent with many other findings (Edwards 1975, Crill 1991, Davidson et al. 1998, Rustad et al. 2000, Xu and Qi 2001). Percent soil organic matter (SOM) was positively correlated with winter and summer CO₂ flux rates (Table 4-6, Fig. 4-8), which was also found by Xu and Qi (2001) during summer months under a ponderosa pine forest. Spring CO₂ flux rates

were positively, though not significantly, correlated with increasing soil bulk density (Fig. 4-9). This was surprising, and opposite of my prediction, as a higher soil density would indicate a more compact soil with smaller pore spaces for gaseous diffusion. However, these soils were not highly compacted, relatively speaking, with the highest bulk density being 0.73-g soil/cm³. In the same study referenced above, Xu and Qi (2001) found a negative correlation between CO₂ efflux and bulk density, attributing the finding to limited pore space for microbial activity. Therefore, the range of bulk densities at my research sites may be too low to detect a similar pattern.

CH₄ flux

Only one of my predictions regarding CH₄ was supported by the data within this study. I found a weak relationship between CH₄ flux and land-use context (Table 4-5, Fig. 4-14), where the greatest methane uptake rates occurred in more urban sites (determined at the subcatchment scale), whose soils were typically driest in the summer and fall. Groffman et al. (2006) found CH₄ uptake rates correlated with land-use context in Baltimore forests. Land-use context was related to high-fertility and low-fertility sites, with high fertility sites occurring primarily in urban environments and low-fertility sites in rural environments. Rates of methane uptake were lower in high fertility forested urban sites than in low fertility rural sites. I found the opposite trend at my sites, with increasing rates of CH₄ uptake with increasing %IS, which may have been due to more proximal scale environmental variation.

Higher spring temperatures resulted in lower rates of CH₄ uptake within these riparian sites. Castro (1995) revealed methane uptake rates are dependent on temperature

within the range of 0 and 10°C, increasing as temperature increases. Above 10°C methane uptake becomes independent of temperature in temperate forest soils. Incubation of forest soil cores has also revealed no relationship between soil temperature above 10°C and methane efflux (Crill 1991). This was attributed to a shift from biological to diffusional controls over methane uptake. During spring thaw, uptake rates responded to temperature since substrate (CH₄ for methanotrophs) was plentiful and microbes increased their activity in response to the temperature increase. Into summer, substrate became limiting as microbial activity increased in response to the soil temperature, and diffusion of CH₄ into the soil became the limiting factor controlling rates of CH₄ uptake. The sites with higher spring temperatures and lower net CH₄ uptake rates suggest substrate-limitation of methanotrophy. Average spring temperatures were above 10°C at my sites, which potentially explains the weak trend between temperature and CH₄ uptake rates.

Patterns in CH₄ flux rates during summer and fall were related to soil moisture and soil carbon and nitrogen concentrations. Soil moisture acted as a switch by determining whether the soil was a net consumer or emitter of CH₄ (Fig. 4-17 & 4-18b, Table 4-6), as methanogenesis is an anaerobic process and soil methane uptake is an aerobic process. Ambus and Christensen (1995) found similar results regarding CH₄ flux and soil moisture in a riparian site where static chambers, arranged along a topographic gradient from upslope to downslope, were used to record methane efflux in the flood plain. They found a significant increase in methane efflux from soils with higher % water-filled pore space (WFPS) and higher % soil organic matter (SOM). Castro et al. (1995) found experimental evidence for negative control of methane consumption by soils at levels of

soil moisture (measured as % WFPS) between 60 and 100% in Harvard Forest in Massachusetts. Whalen et al. (1990) found the lowest methane consumption rates occurred above 40% and below 5% soil moisture by weight. Reported optimum values for gravimetric soil moisture (g soil water/g dry soil) in relation to methane uptake occur between the ranges of 10-30% (Whalen et al. 1990, Torn and Harte 1996). In my riparian systems soil moisture values above ~0.35-g water/g dry soil, slightly higher moisture values than these other studies, promoted net methane efflux from the soil (Fig. 4-17). One site in particular (RG7) exhibited CH₄ efflux at nearly every date measured. This site also had some of the highest monthly soil moisture values (Table 4-1) and an average depth to groundwater of 91-cm, and the highest groundwater nitrate concentrations from May to October 2008. Net methane uptake occurred at lower soil moisture levels; however, these soil moisture values did not explain variation in my data set.

In summer, net CH₄ uptake increased with decreasing soil % carbon (Fig. 4-16, Table 4-6). This could partially explain increasing methane uptake rates with increasing subcatchment %IS mentioned previously, since soil % carbon was negatively correlated with %IS (Pearson's $r = -0.69$). However, soil % carbon was determined from fall soil collects and they may have been slightly different had they been taken during summer months. In fall, net CH₄ uptake increased with decreasing soil C:N ratios (Fig. 4-18a, Table 4-6). The relationship with C:N ratios was consistent with responses others have found regarding flux rates and soil nitrogen concentrations. Steudler et al. (1989) found CH₄ uptake increased with concentrations of soil nitrogen, particularly ammonium, via fertilization in southeastern temperate forests. This was believed to occur due to increased nitrifier activity, particularly *Nitrosomonas europaea*, which also possess the

ability to oxidize CH₄ (Jones and Morita 1983). Since higher soil nitrogen concentrations would lead to lower C:N ratios (high fertility), and greater methane uptake, nitrifier activity may have been responsible for this pattern within these sites. From a soil fertility perspective, my results are opposite those found by Groffman et al. (2006), who found lower rates of CH₄ uptake at high fertility (low C:N) urban sites. However, in my study, no link between land-use and fall CH₄ flux rates was found (Table 4-5).

No patterns were found regarding winter CH₄ flux between sites or with site or soil variables, which may have been due to the frozen soils. Gullledge and Schimel (2000) found no relationship between climatic factors and methane consumption within taiga forests in Alaska, likely due to the semi-arid nature of the area's climate. In conclusion, net methane flux rates within my riparian sites appear to be controlled by seasonal and local site variables rather than factors operating at the land-use scale.

N₂O flux

Nitrous oxide flux rate differences, while not correlating with any environmental variables, did show a dramatic seasonal decrease at many sites from August to October. Average soil temperature during this period decreased from 23°C to approximately 16°C. This likely contributed to the decrease in N₂O flux at some of the sites, although it does not explain steady or increased flux at other sites. As for CH₄ flux, interpreting the dynamics of nitrous oxide fluxes is complicated by the fact that it is formed by both aerobic (nitrification) and anaerobic (denitrification) microbial processes. Davidson et al. (1993) found nitrification to be the dominant source of N₂O production in a tropical forest soil following soil wetting after the dry season. This could not explain variation in

my data however, as no large rain pulses occurred prior to my sampling dates, and no consistent responses were observed among my sites either by land use or proximity to one another. Others have found N₂O emissions varied with soil NO₃⁻ concentrations, soil organic matter (SOM), and pH among a range of habitat types (Ambus and Christensen 1995). Arable and coastal grasslands exhibited the highest annual mean N₂O emissions from among seven different habitat types, including forested and riparian sites. While none of my predictions were supported, further research should also include measures of *in situ* nitrification rates and soil moisture monitoring to determine potential associations with N₂O flux in riparian sites, and modeling a mechanistic partitioning between denitrification and nitrification as explanations for seasonal and land-use scale variation in emissions of this important greenhouse gas.

Conclusions

Depth to water table was lower in urban areas, as expected, since urban areas had the highest stream banks (Fig. 4-3 & 4-4). In addition, the degree of groundwater recharge in urban areas during spring rains was much less than in rural areas, demonstrating the effect of impervious surface runoff and structural bypasses on riparian groundwater levels. The absence of recharge means the upper soil layers are never as saturated as in rural areas and soil moisture remained lower in the urban riparian areas as well. Therefore I found evidence to support the notion that urban hydrological drought exists in riparian areas in urban and urbanizing catchments in Louisville.

Carbon dioxide flux rates increased with increasing soil temperatures among all sites (Fig. 4-1 & 4-6). Site flux difference and potential explanatory variables, however,

changed in response to season, with winter fluxes influenced by % SOM (Fig. 4-8). As temperatures increased in the spring, the degree of CO₂ release was better explained by soil bulk density (Fig. 4-9). However, the trend was opposite of what I expected, with greater flux rates occurring in the denser soils. However, soil bulk density in these systems ranged only from 0.49- to 0.73-g/cm³, and were not considered to be compacted. Soil temperatures played the greatest role in explaining land-use patterns of CO₂ release during the hottest months and into fall, with urban riparian sites releasing more CO₂ than suburban and rural sites.

Explaining patterns of net soil CH₄ flux rates is more complex since net flux is controlled by both anaerobic and aerobic processes. Soil temperature weakly correlated with site flux rates during the spring thaw, with lower net CH₄ uptake rates at higher spring temperatures attributed to substrate limitation (atmospheric CH₄ diffusion into the soil) of microbial activity (Fig. 4-14). As soil temperatures and soil microbial activity increased, other site differences began to affect patterns of CH₄ uptake and efflux. Summer net CH₄ uptake rates were negatively correlated with soil % carbon (Fig. 4-16). Differences in fall CH₄ uptake rates were negatively correlated with soil C:N ratios (Fig. 4-18a). Soils with C:N values below 23 are associated with higher net annual nitrification potential. The latter half of the summer of 2008 was one of prolonged drought. Precipitation events in the fall could have stimulated the activity of nitrifying bacteria, which are also known to take up CH₄ (Jones and Morita 1983, Bedard and Knowles 1989). This offers one potential explanation for the negative correlation with soils C:N ratios. Potential effects of high soil moisture on the net balance between methanogenic and methanotrophic activity were evidenced during summer and fall seasons. Higher soil

moisture values (greater than ~ 0.35 -g soil water/g dry soil) resulted in a shift from net methanotrophy to net methanogenesis. Patterns of net methane uptake, however, were not explained by variation in soil moisture below ~ 0.35 -g soil water/g dry soil.

N_2O efflux rates showed a strong seasonal response between August and October, decreasing from hot and dry August to cooler October. No measured environmental site differences could explain the flux differences across sites. Groffman et al. (2006) found natural soil factors had greater control over soil nitrogen cycling dynamics, with higher N_2O flux rates attributed to finer textured soils. The differences could also be a function of the dual processes in which N_2O is released from the soil, one process being aerobic (nitrification) and one being anaerobic (denitrification).

Table 4-1: Gravimetric soil water content (g water/g oven dry soil) in the upper 15-cm horizons within two meters of streambanks in urban, suburban, and rural riparian sites. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover. Site names beginning with R, S, and U are located in rural, suburban, and urban subcatchments, respectively. The second letter in the site name indicates whether the site is located in Harrods (H), Goose (G), or Beargrass (B) creek watershed. N/A indicates the soils were frozen.

Site	Jan.	Feb.	March	April	May	June	July	Aug.	Sept.	Oct.
RG4	N/A	0.44	0.36	0.34	0.32	0.32	0.31	0.22	0.21	0.21
RG7	N/A	0.41	0.56	0.53	0.52	0.56	0.54	0.47	0.48	0.45
RH13	0.04	0.39	0.38	0.38	0.34	0.33	0.23	0.12	0.12	0.12
RH5	0.05	N/A	0.37	0.37	0.35	0.38	0.32	0.17	0.21	0.18
Mean:	0.02	0.31	0.42	0.40	0.38	0.40	0.35	0.25	0.26	0.24
SB12	N/A	0.37	0.42	0.36	0.35	0.33	0.25	0.15	0.17	0.18
SG13	N/A	0.61	0.46	0.45	0.42	0.42	0.42	0.36	0.37	0.30
SG9	N/A	0.42	0.41	0.38	0.35	0.33	0.28	0.20	0.21	0.21
Mean:	N/A	0.47	0.43	0.40	0.38	0.36	0.32	0.24	0.25	0.23
UB1	N/A	N/A	0.33	0.30	0.29	0.27	0.25	0.16	0.17	0.17
UB14	N/A	N/A	NA	NA	NA	NA	0.40	0.26	0.23	0.26
UB2	0.36	0.38	0.33	0.32	0.31	0.27	0.22	0.16	0.17	0.15
UB3	0.36	0.39	0.37	0.36	0.35	0.33	0.30	0.20	0.18	0.15
UB5	0.31	N/A	0.36	0.31	0.30	0.21	0.23	0.14	0.13	0.11
UG8	0.00	0.33	0.35	0.30	0.33	0.29	0.26	0.18	0.15	0.17
Mean:	0.21	0.22	0.35	0.32	0.32	0.28	0.28	0.18	0.17	0.17

Table 4-2: Site mean vegetation data surrounding the static chambers at riparian sites. Percent bare ground and herbaceous cover, as well as shrub and sapling counts were assessed within a 12.6 m² area (2-m radius) surrounding each chamber. Tree counts were assessed within a 78.5 m² area (5-m radius) surrounding each chamber. Canopy cover was determined directly over each chamber. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover. Site names beginning with R, S, and U are located in rural, suburban, and urban subcatchments, respectively. The second letter in the site name indicates whether the site is located in Harrods (H), Goose (G), or Beargrass (B) creek watershed.

Site	% cover			Counts		
	Canopy	Bare ground	Herbaceous	Shrubs	Saplings	Trees
RG4	48	37.5	62.5	0.5	0	1
RG7	61.5	37.5	37.5	2.5	6	1
RH13	38	37.5	37.5	23	0.5	1.5
RH5	12.5	85	15	0	2	1
Mean:	40	49	38	7	2	1
SB12	53	62.5	37.5	58.5	0	4.5
SG13	39.5	62.5	37.5	0	0	1
SG9	48	15	73.75	2	0	5
Mean:	47	47	50	20	0	4
UB1	37	85	15	0	0	2
UB14	39.5	37.5	62.5	1.5	2.5	1.5
UB2	19	62.5	37.5	27.5	0.5	3
UB3	45.5	62.5	37.5	3	6.5	1.5
UB5	44	62.5	37.5	30.5	0	1.5
UG8	17.5	85	15	8.5	0	2.5
Mean:	34	66	34	12	2	2

Table 4-3: Riparian groundwater nitrate+nitrite-N monthly means (\pm S.E.) for subcatchment land-use categories and total monthly rainfall from November 2007 to October 2008. Nitrate+nitrite-N values are means from five or six urban, 3 suburban, and 3 rural sites. Rainfall was determined from a single location monitored by the Louisville Metropolitan Sewer District within Goose (suburban/rural) and Beargrass Creek (Urban) watersheds. Average rainfall is the mean of both stations. Urban subcatchments contain \geq 30% impervious surface cover, rural subcatchments contain \leq 10% impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

Month	Nitrate+nitrite-N - mg/L (SE)			Rainfall (cm)		
	Urban	Suburban	Rural	Urban	Suburban/ Rural	Avg. Rain
Nov.	0.80 (.29)	0.33 (0.16)	0.35 (0.07)	7.1	8.3	7.7
Jan.	13.17 (8.51)	0.082 (0.01)	0.23 (0.09)	8.3	8.2	8.25
March	13.84 (8.34)	0.12 (0.01)	0.14 (0.06)	26.7	27.1	26.9
April	6.60 (6.53)	0.10 (0.02)	0.087 (0.02)	15.7	13	14.4
May	6.66 (4.83)	0.049 (0.01)	7.29 (4.71)	15.2	15.6	15.4
June	14.50 (8.09)	0.068 (0.02)	12.76 (9.35)	7.6	10.3	8.9
July	0.10 (0.01)	0.068 (0.01)	0.16 (0.07)	14.6	9.3	12
August	0.10 (0.02)	0.12 (0.05)	4.42 (N/A)	1.4	1.2	1.3
Sept.	0.13 (0.04)	0.057 (0.01)	13.57 (4.10)	3.4	5.2	4.3
Oct.	0.15 (0.06)	0.046 (0.003)	8.20 (8.09)	2.1	2.2	2.1

Table 4-4: Riparian groundwater dissolved organic carbon (DOC) monthly means (\pm S.E.) for subcatchment land-use categories and total monthly rainfall. DOC values are means from five or six urban, three suburban, and three rural sites. Rainfall was determined from a single location monitored by the Louisville Metropolitan Sewer District within Goose (suburban/rural) and Beargrass Creek (Urban) watersheds. Average rainfall is the mean of both stations. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

Month	DOC - mg/L (SE)			Rainfall (cm)		
	Urban	Suburban	Rural	Urban	Sub/Rural	Avg. Rain
November	6.23 (1.07)	7.31 (1.53)	9.89 (1.39)	7.1	8.3	7.7
January	8.06 (1.94)	5.48 (0.48)	4.58 (0.21)	8.3	8.2	8.25
March	4.17 (0.60)	4.58 (1.18)	4.69 (0.85)	26.7	27.1	26.9
April	5.75 (0.44)	6.74 (0.53)	6.08(1.40)	15.7	13	14.4
May	4.28 (1.10)	5.27 (0.81)	6.29 (1.38)	15.2	15.6	15.4
June	8.76 (0.88)	7.43 (0.27)	6.59 (0.83)	7.6	10.3	8.9
July	6.60(1.12)	5.47 (0.66)	10.51 (6.33)	14.6	9.3	12
August	5.81 (0.57)	5.54 (0.30)	5.62 (NA)	1.4	1.2	1.3
September	N/A	N/A	N/A	3.4	5.2	4.3
October	5.85 (0.66)	5.85 (0.12)	6.92 (0.87)	2.1	2.2	2.1

Table 4-5: Relationships between landscape-level soil factors and CO₂ and CH₄ fluxes in riparian areas across an urban-to-rural land-use gradient.

Greenhouse Gas	Season	Variables	Ind p-value	Model p-value	Adj. r ²	Model Equation
Carbon dioxide (mg CO ₂ -C/m ² /day)	Spring	%IS - 1 km Catch. area	0.39 0.33	0.46	-0.03	
	Summer	%IS - 1 km Catch. area	0.002 0.002	0.001	0.72	y = 43(%IS) - 32(Catch. area) + 2449
	Fall	%IS - 1 km Catch. area	0.003 0.01	0.002	0.65	y = 43(%IS) - 26(Catch. area) + 1618
Methane (mg CH ₄ -C/m ² /day)	Spring	%IS - SC Catch. area	0.31 0.24	0.37	0.02	
	Summer	%IS - SC	-	0.06	0.21	y = -0.02(% IS) - 0.03
	Fall	%IS - SC Catch. area	0.26 0.34	0.3	0.06	

%IS - 1 km = % impervious surface within 1 km of the research sites

%IS - SC = % impervious surface within the subcatchment in which the site is located.

Catch. area = cumulative catchment area (ha) within and upstream of the subcatchment in which the site is located.

Ind. p-value = p-values for individual explanatory variables in a multiple regression model.

Table 4-6: Relationships between plot-level soil factors and CO₂ and CH₄ fluxes in riparian areas across an urban-to-rural land-use gradient.

Greenhouse Gas	Season	Proximal variables	Ind. p-value	Model		Model Equation
				p-value	Adj. r ²	
Carbon dioxide (mg CO ₂ -C/m ² /day)	Winter	Soil % OM		0.004	0.58	y = 187(OM) - 438
	Spring	Soil BD		0.08	0.2	Ln(y) = 2(BD) + 6.3
	Summer	Soil temp.	0.011	0.024	0.43	y = 715(temp.) + 262(OM) - 13746
		Soil % OM	0.0496			
Fall	Soil temp.		0.004	0.55	y = 748 (temp.) - 11888	
Methane (mg CH ₄ -C/m ² /day)	Spring	Soil temp.		0.067	0.28	y = 0.15(temp.) - 2.45
	Summer	Soil % Carbon		0.003	0.54	y = 0.19 (%carbon) - 1.2
	Fall	Soil molar C:N	0.001	0.004	0.75	y = 0.03(C:N) + 1.9(moisture) - 1.8
Soil moisture		0.005				

BD = bulk density (g/m³); Soil moisture = g water/g dry soil; Soil temperature = °C

Ind. P-value = p-values for individual explanatory variables in a multiple regression model.

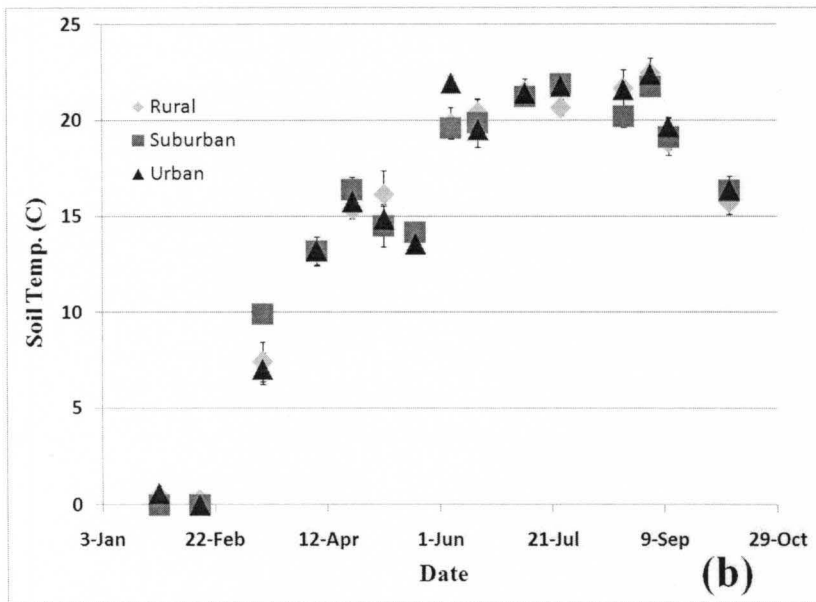
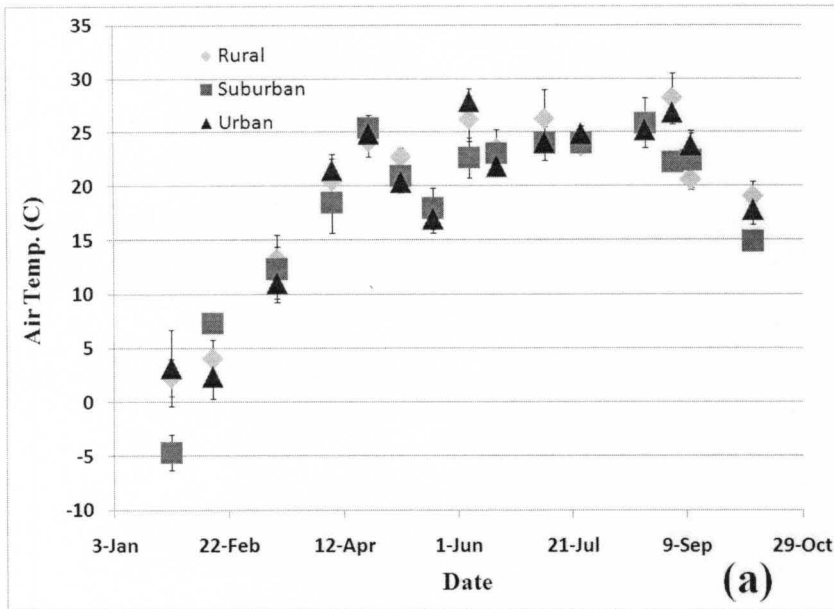


Figure 4-1a-b: Air temperature and soil temperature taken during twice-monthly gas sampling visits in urban, suburban, and rural riparian sites in 2008. Soil temperature was taken in two locations within each site to five, ten, and fifteen cm depths and averaged to obtain a representative soil temperature through the soil column. Air temperature was taken between the two soil temperature locations. Points are means of five urban, three suburban, and four rural sites with standard error bars. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

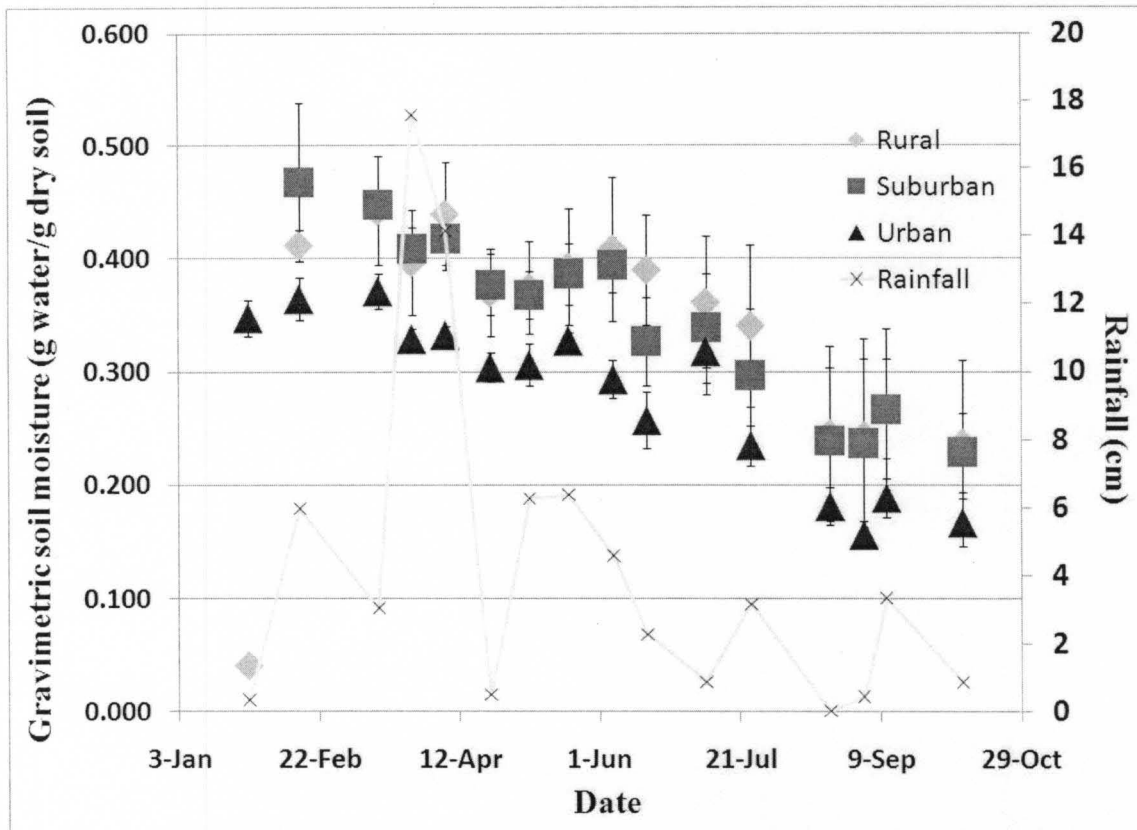
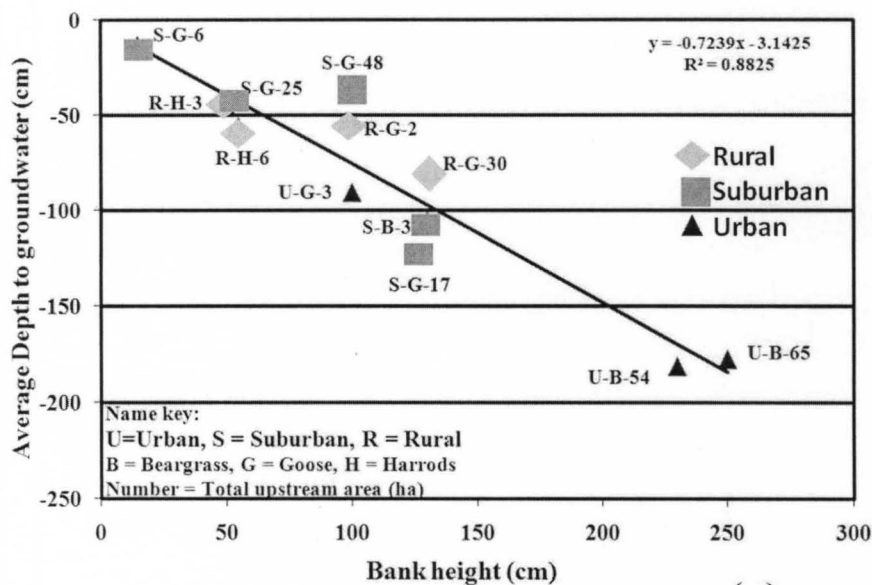
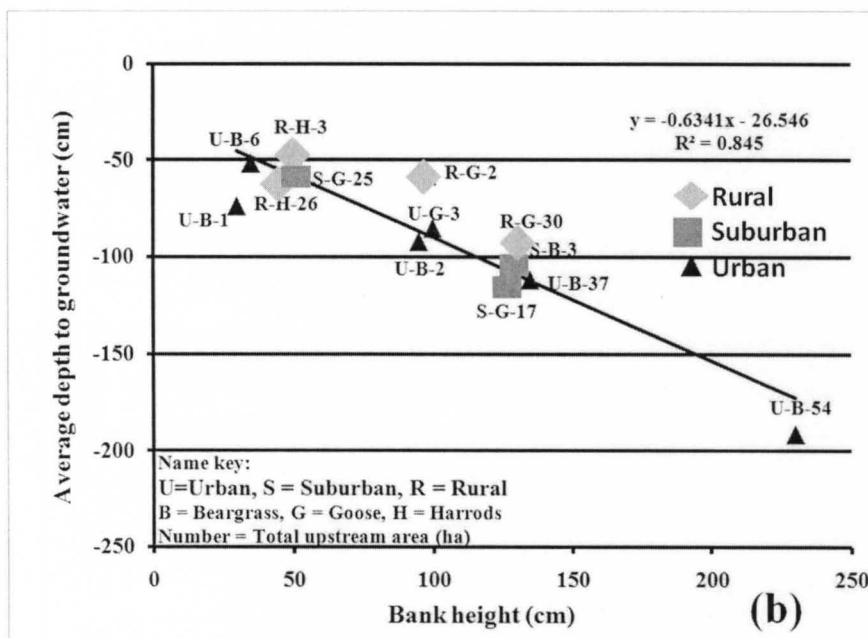


Figure 4-2: Gravimetric soil moisture in the upper 15-cm soil horizon in urban, suburban, and rural riparian sites in 2008. Points are means of five urban, three suburban, and four rural sites with standard error bars. Rainfall was determined from a single location monitored by the Louisville Metropolitan Sewer District within Goose and Beargrass creek watersheds and represents total ppt. monthly in January and February and every two weeks from March to October. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.



(a)



(b)

Figure 4-3a-b: Nearstream depth to groundwater (water table) in relation to bank height in years 2007 (a) and 2008 (b). Depth to groundwater was measured twice a month in riparian groundwater wells in urban, suburban, and rural sites from November 2006 to October 2007 (a) and from November 2007 to October 2008 (b). Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

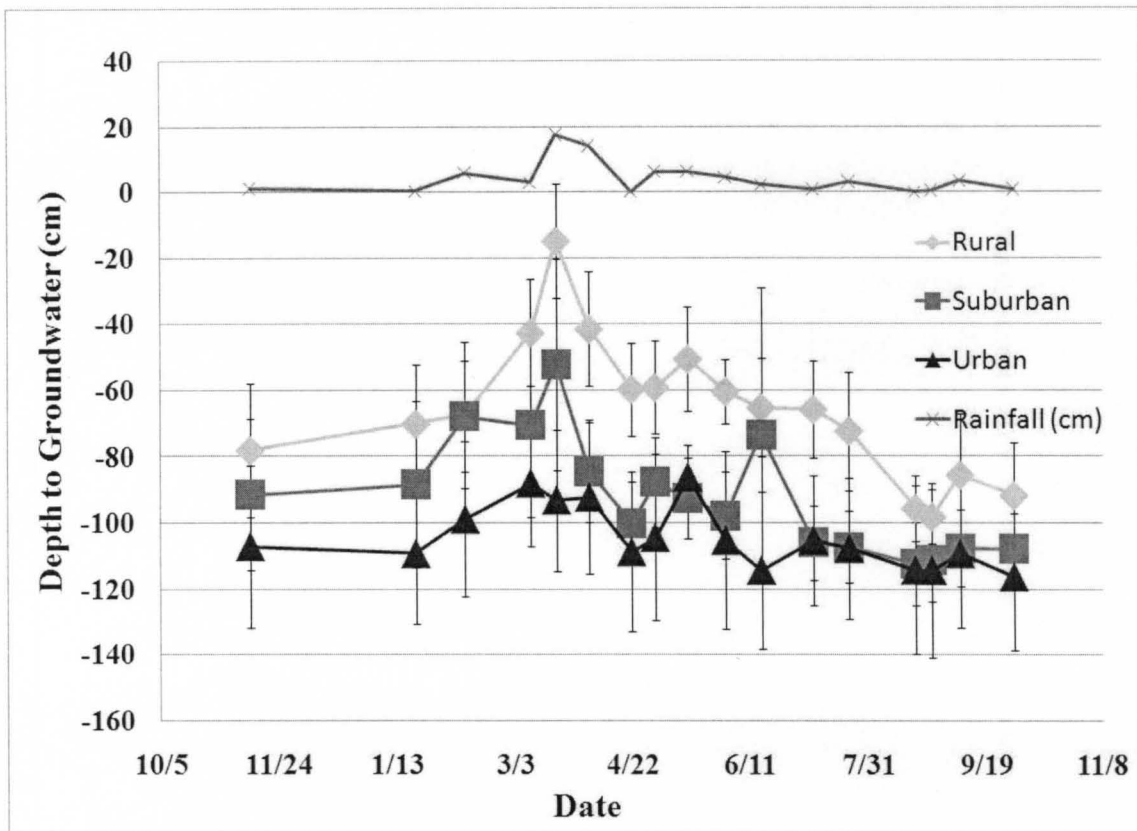


Figure 4-4: Nearstream depth to groundwater (water table) from November 2007 to October 2008. Depth to groundwater was measured once or twice a month in riparian groundwater wells in urban, suburban, and rural sites based on subcatchment % impervious surface. Points are means of five urban, three suburban, and three rural sites with standard error bars. Rainfall was determined from a single location monitored by the Louisville Metropolitan Sewer District within Goose and Beargrass creek watersheds and represents total ppt. monthly in January and February and every two weeks from March to October. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover. Repeated-measures ANOVA revealed a significant time effect within groups ($p = <0.0001$), but no significant between-group effects were observed ($p = 0.44$).

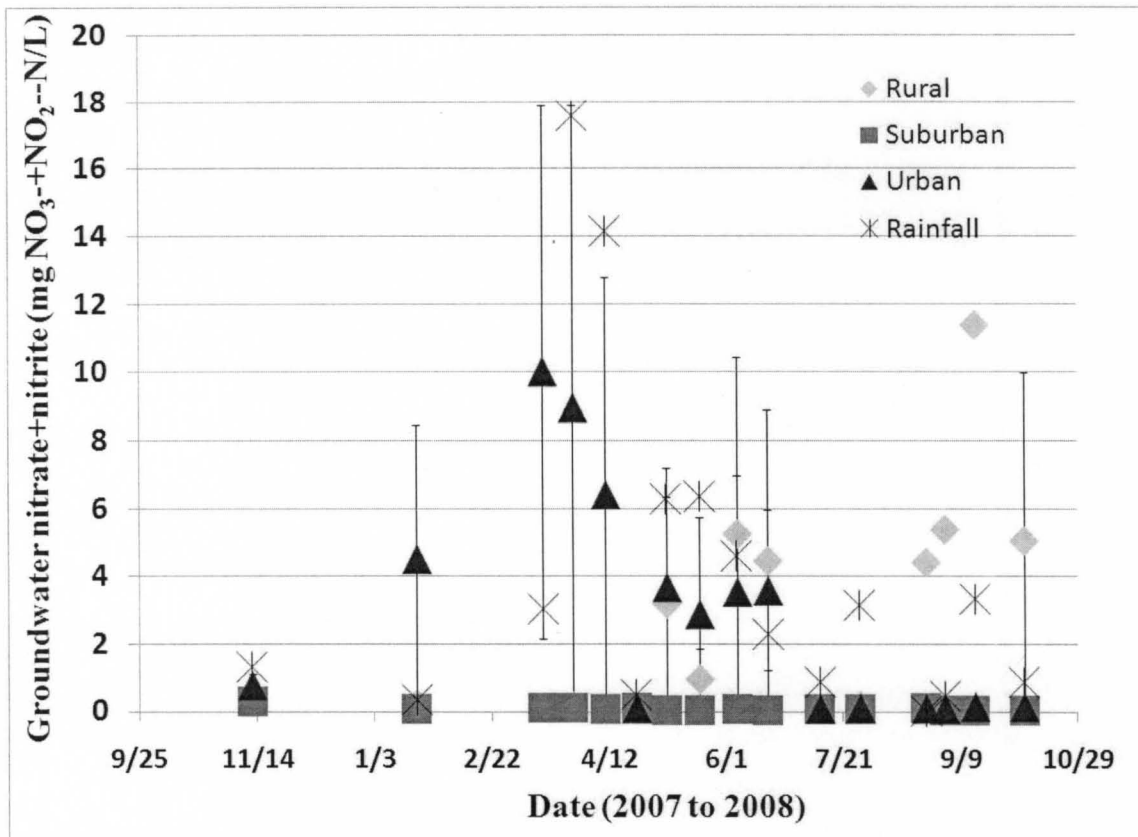


Figure 4-5: Mean nitrate+nitrite-N nearstream riparian groundwater concentrations in urban, suburban, and rural subcatchments. Nitrate+nitrite-N values are means from five or six urban, 3 suburban, and 3 rural sites. Rainfall was determined from a single location monitored by the Louisville Metropolitan Sewer District within Goose and Beargrass creek watersheds and represents total ppt. monthly in January and February and every two weeks from March to October. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

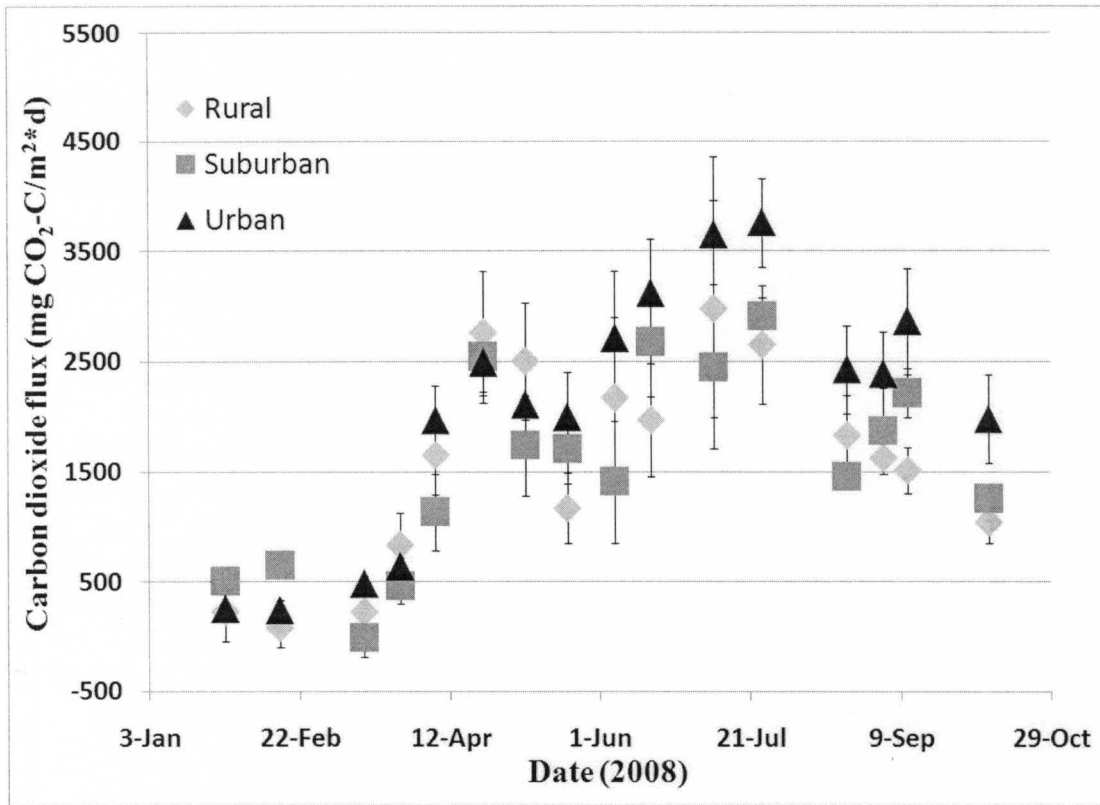


Figure 4-6: Mean (\pm SE) riparian CO₂ flux between January and October 2008 averaged for sites in rural, suburban, and urban subcatchments. Points are means of five or six urban, three suburban, and four rural values. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

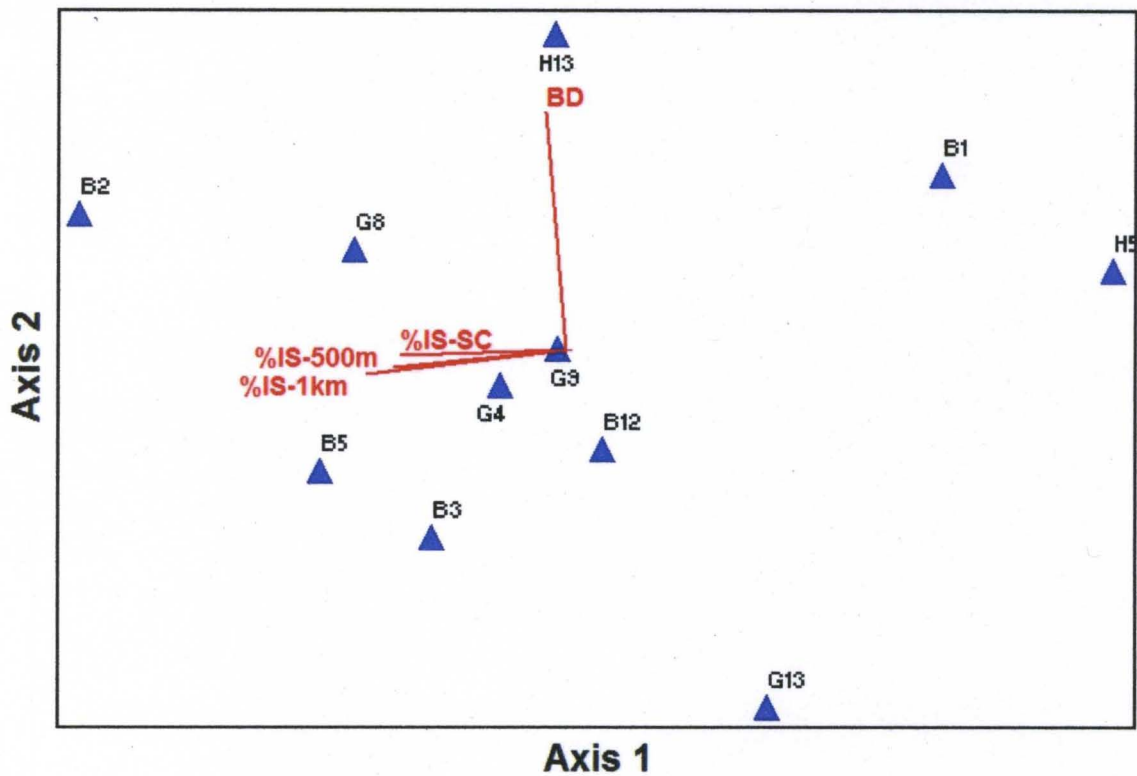


Figure 4-7: Principal components analysis (PCA) of sites and monthly CO₂ fluxes. Sites are arranged according to mean annual CO₂ flux along axis 1, with increasing CO₂ emissions from right to left. Axis 2 separation among sites was primarily explained by soil bulk density, which increases from bottom to top of the graph. Vectors represent site and soil properties that have a coefficient of determination ≥ 0.3 with at least one axis. Visible site and soil variables are BD = bulk density, %IS = percent impervious surface at two site radius buffers – 500-meters (500m) and 1-kilometer (1km) - and within the subcatchment (SC) in which the site is located.

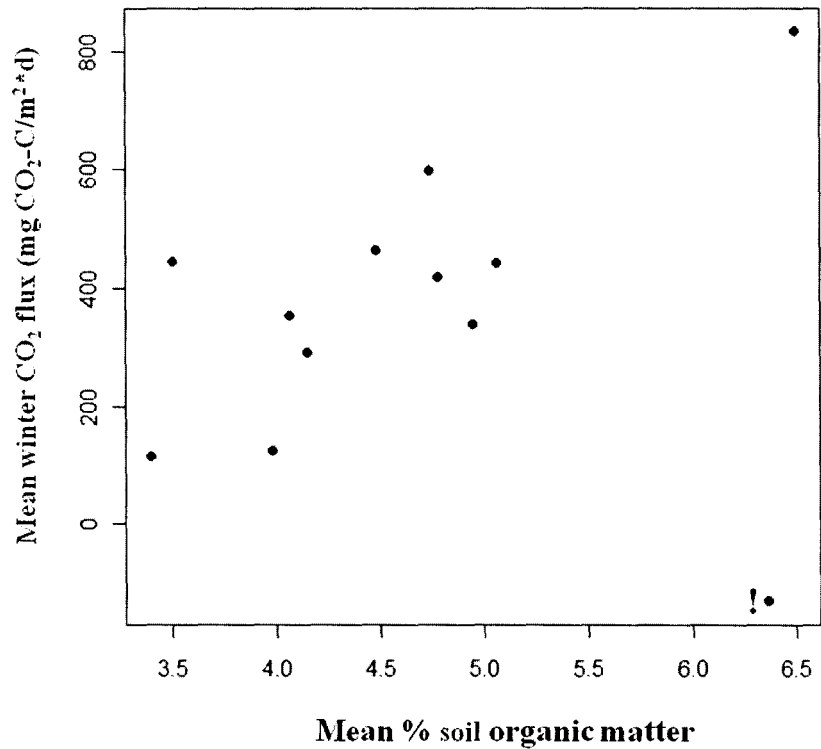


Figure 4-8: Relationship between riparian winter CO₂ flux and % soil organic matter (OM). Carbon dioxide fluxes and % soil OM were averaged for each site for cold weather months (January, February, and March) and entered into a linear regression model ($p = 0.004$, $\text{adj. } r^2 = 0.58$). The site marked with an “!” was determined to be an outlier as described in Methods (this chapter).

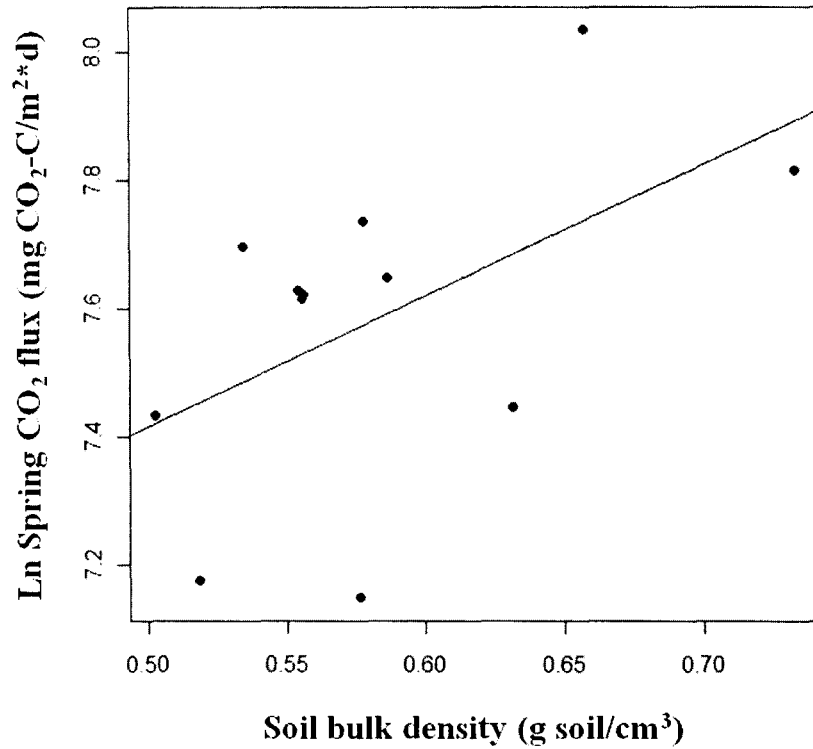


Figure 4-9: Relationship between riparian spring CO₂ flux and soil bulk density (BD). Carbon dioxide flux was averaged for each site for spring months (April and May). BD was determined from October 2008 soil samples. BD in this range exhibited a positive relationship with CO₂ flux ($p=0.081$; $r^2 = 0.20$).

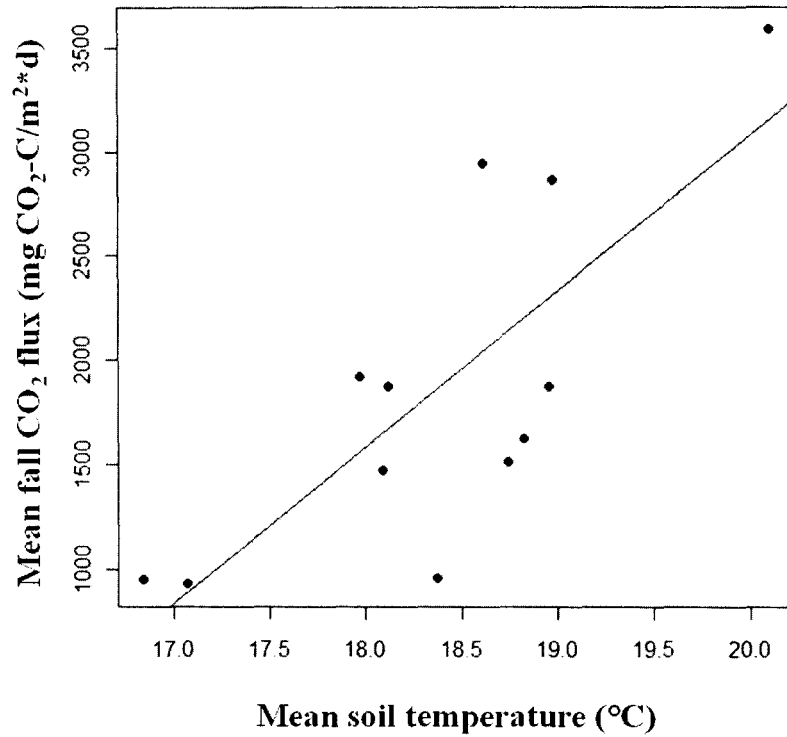


Figure 4-10: Increasing mean fall CO₂ flux rates with increasing mean soil temperature among riparian sites. Each point is the average of three site measurements for each variable taken during September and October 2008. The adjusted r^2 was 0.56 and the p-value was 0.004.

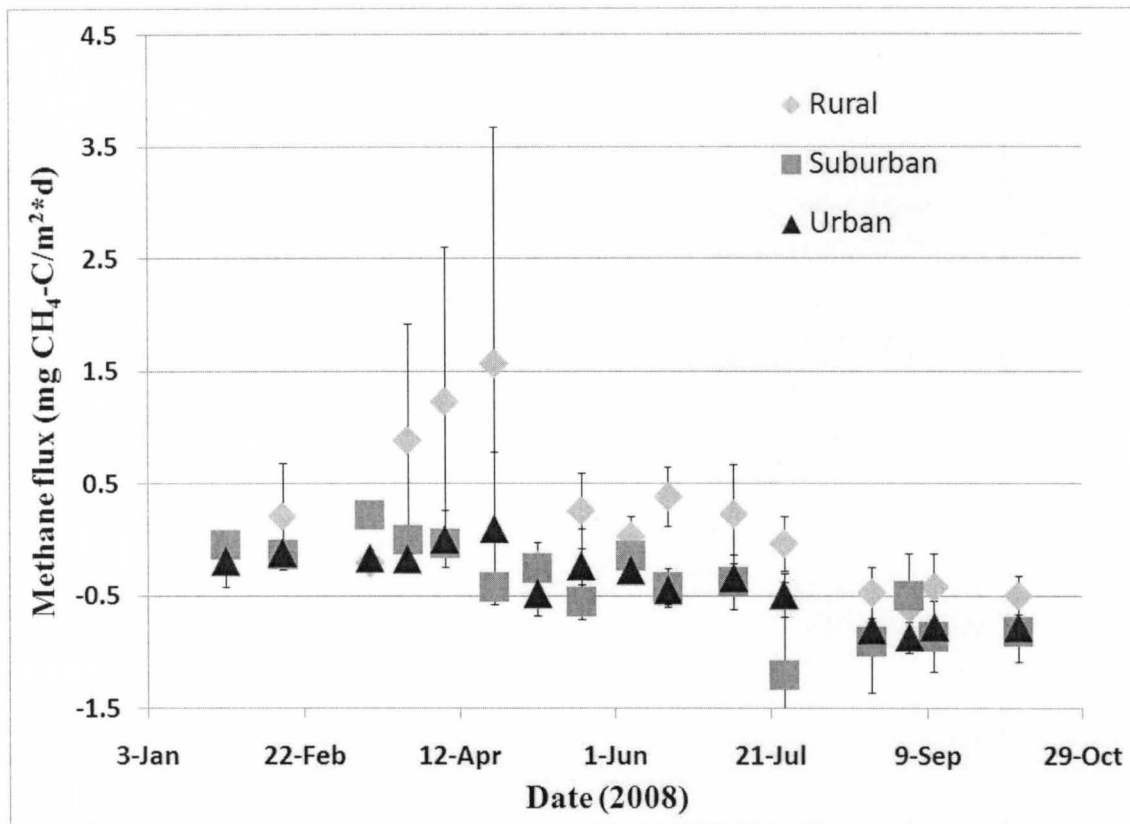


Figure 4-11: Mean (\pm SE) riparian CH₄ fluxes from January to October 2008 averaged for sites in rural, suburban, and urban subcatchments. Points are means of five or six urban, three suburban, and four rural values. Values above zero indicate net CH₄ efflux from the soil. Values below zero indicate net CH₄ uptake. Urban subcatchments contain \geq 30% impervious surface cover, rural subcatchments contain \leq 10% impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

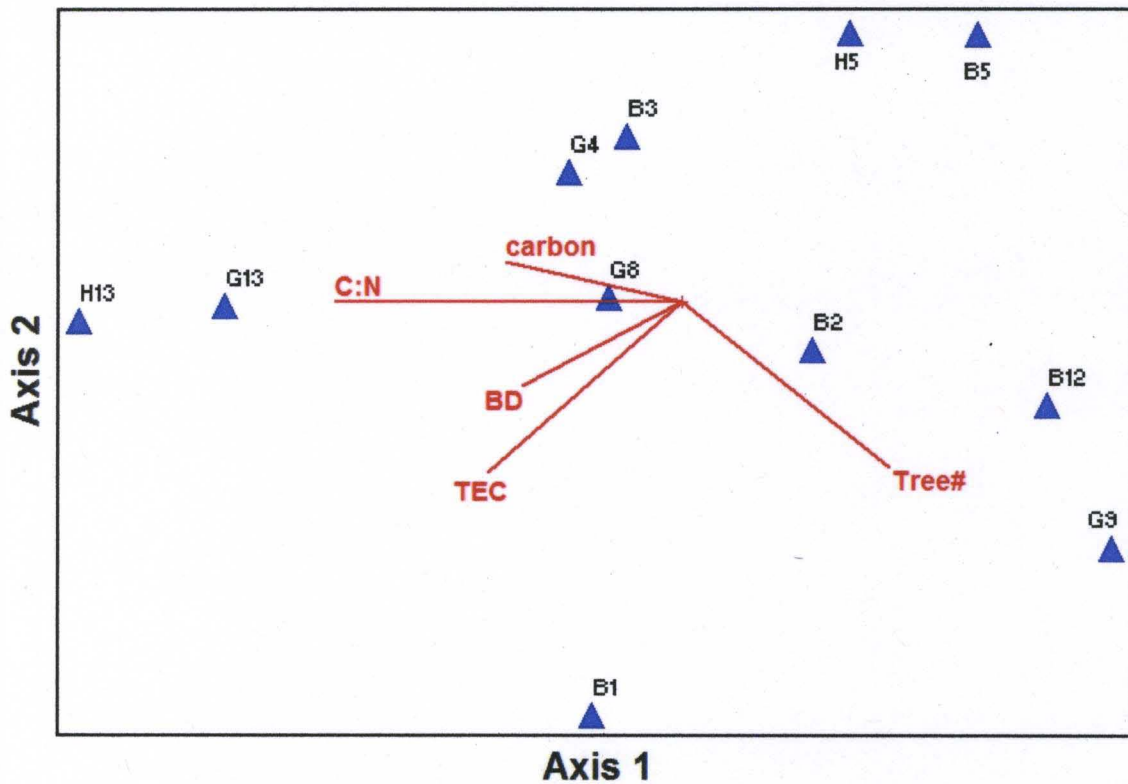
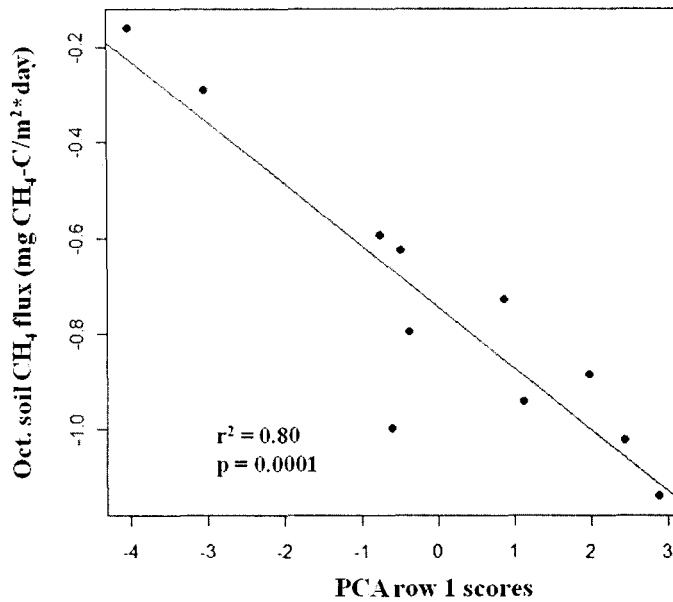
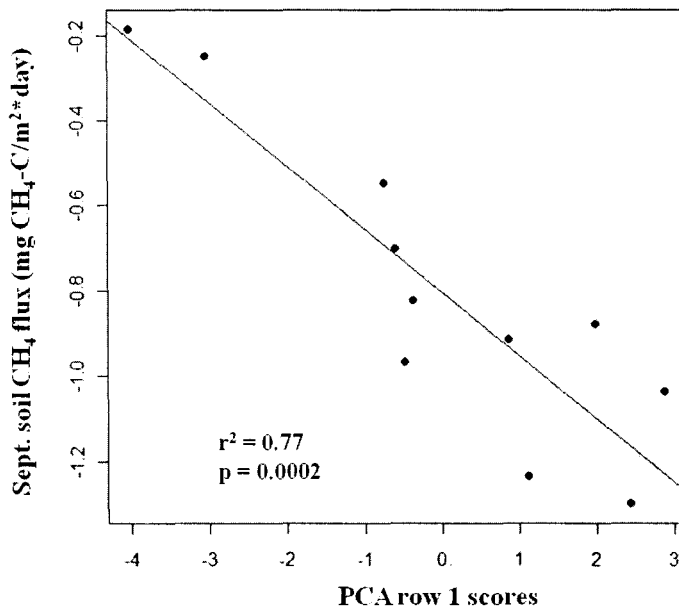


Figure 4-12: Principal components analysis (PCA) of sites and mean monthly CH₄ fluxes from January to October 2008. Sites are arranged according to monthly average CH₄ flux along axis 1, with increasing CH₄ uptake from left to right. Vectors represent site and soil properties that have a coefficient of determination ≥ 0.3 with at least one axis. Soil variables are TEC = total exchange capacity, BD = bulk density, carbon = % carbon, C:N = molar C:N ratio. The site variable Tree# represents average number of trees > 1-m high and ≥ 2.54 -cm diameter breast height within five meters of the area of CH₄ flux determination.



(a)



(b)

Figure 4-13a-b: Mean monthly methane flux rates at each site from 2008 in relation to PCA row 1 scores from Fig. 4-12 for the months October (a) and September (b). Site CH₄ fluxes occur along a linear gradient from left to right on x-axis from low methane uptake to high methane uptake in September and October. The linear pattern began to dissipate in August ($r^2 = 0.55$) and the gradient becomes non-linear in July as well as in months prior through January (not shown).

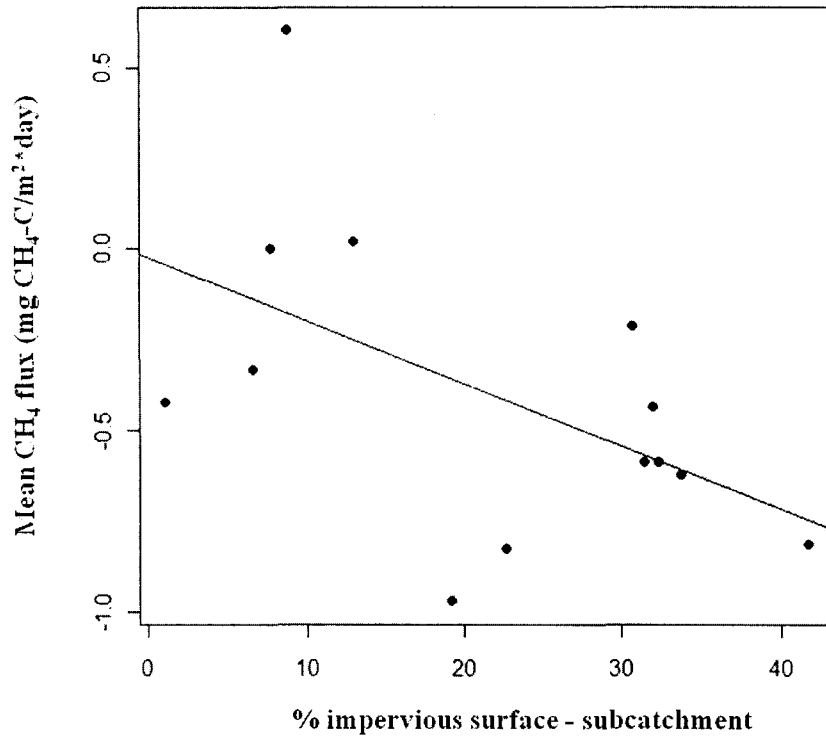


Figure 4-14: Changes in mean summer CH₄ flux in relation to subcatchment %IS. Site methane fluxes and soil moisture values were averaged over June, July, and August 2008. Soil uptake of CH₄ increased with increasing %IS ($p = 0.06$, $\text{adj. } r^2 = 0.21$, $y = 0.02x - 0.03$). Values below zero indicate mean CH₄ uptake of by soils.

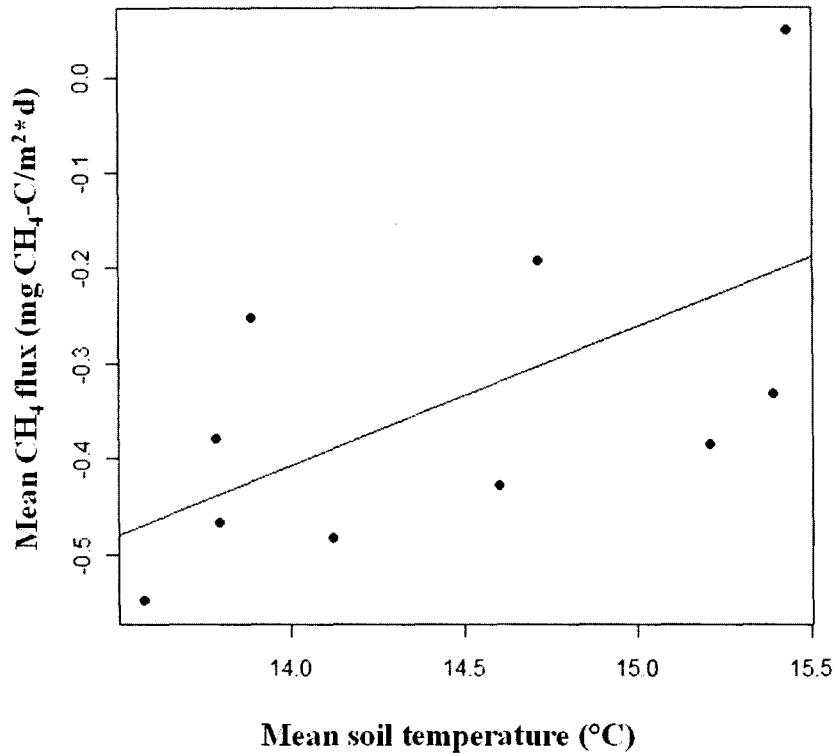


Figure 4-15: Changes in mean spring CH₄ flux with increasing mean soil temperature among sites averaged over April and May 2008 ($p = 0.067$; adj. $r^2 = 0.28$; $y = 0.15x - 2.45$). Methane fluxes and soil temperature were taken on two separate dates each month. Negative fluxes indicate soil uptake from the atmosphere. Two sites (UB1 & RG7) were removed from this analysis because they exhibited high levels of net CH₄ efflux.

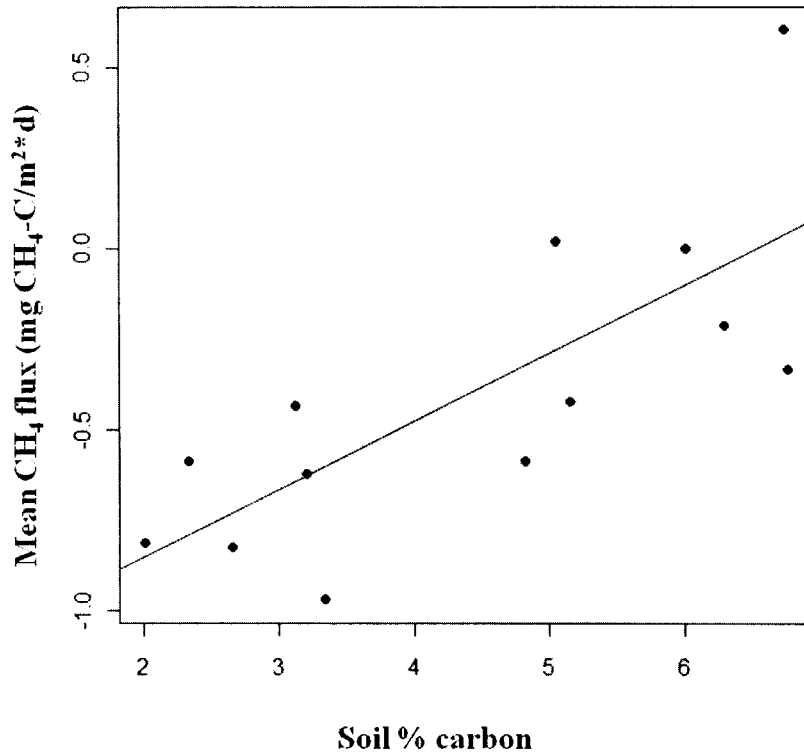


Figure 4-16: Summer mean net CH₄ uptake (values less than zero) increases (values from left to right) with decreasing soil % carbon. Five flux measurements, taken in June, July, and August, 2008, were averaged to obtain mean flux. Soil % carbon was determined for each site from a single sample taken in October 2008. The p-value was 0.003 and the adjusted r² was 0.5 ($y = 0.19x - 1.2$).

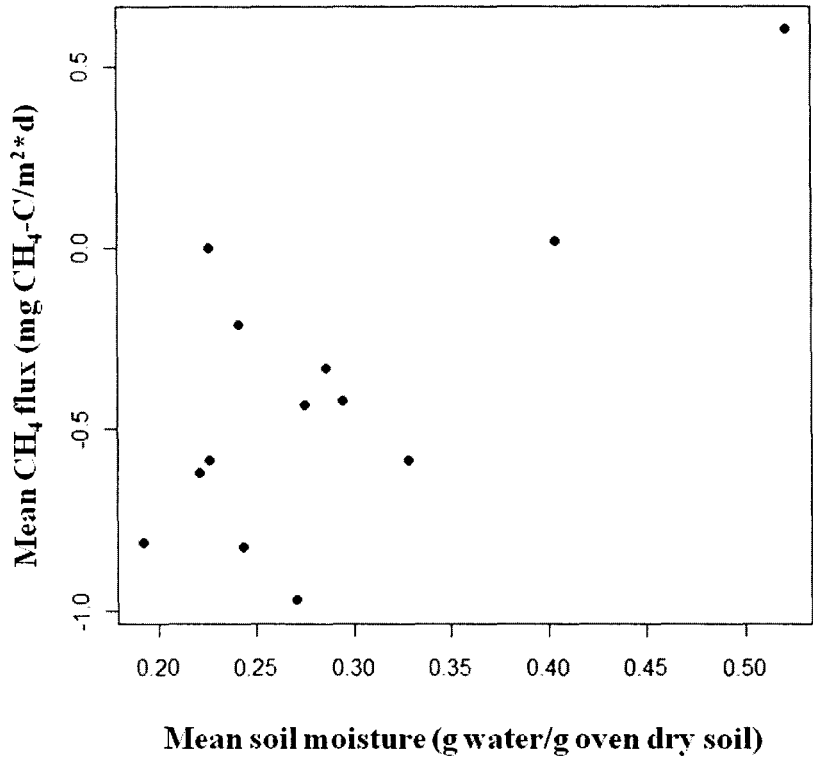


Figure 4-17: Variation in site CH₄ flux with soil moisture. Site methane fluxes and soil moisture values were averaged over June, July, and August 2008. Methane fluxes and soil moisture were measured twice monthly except August, when one sample was collected.

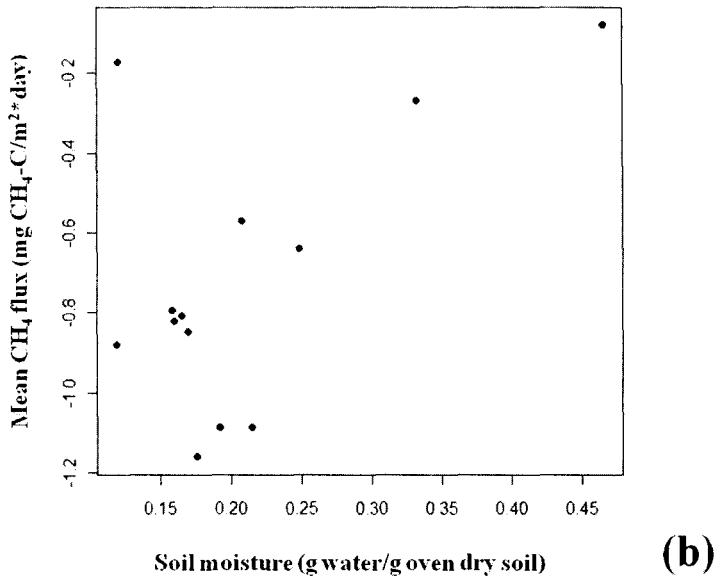
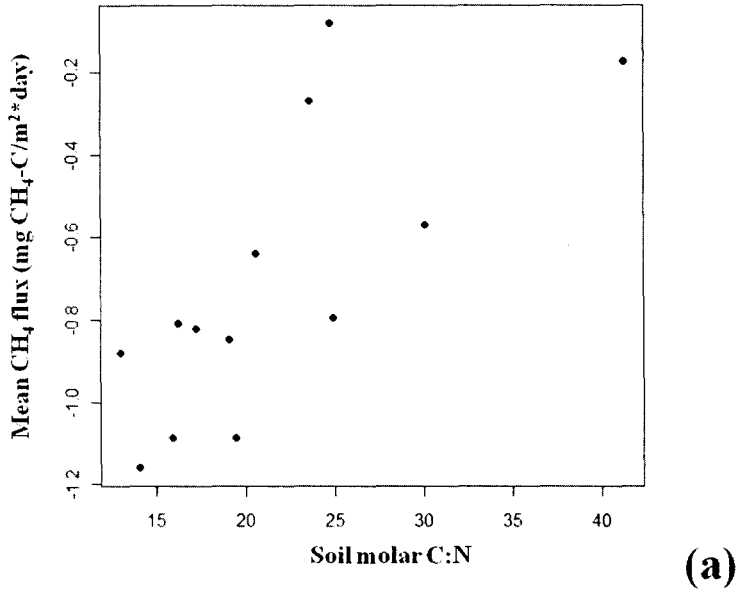


Figure 4-18a-b: Relationships between mean fall 2008 CH₄ flux and soil molar C:N ratios (a) and gravimetric soil moisture (b). Methane fluxes from September and October were averaged for each site. Soil C:N ratios were determined from soil samples collected in October. Gravimetric soil moisture was determined to a depth of 15-cm adjacent to each gas sampling chamber and averaged for the site each time samples were collected. A multiple regression was used to assess relationships of these two variables with CH₄ flux. The overall adjusted coefficient of determination for the model was 0.75, with a p-value = 0.0004. Individual p-values for each variable and the equation can be found in Table 4-6.

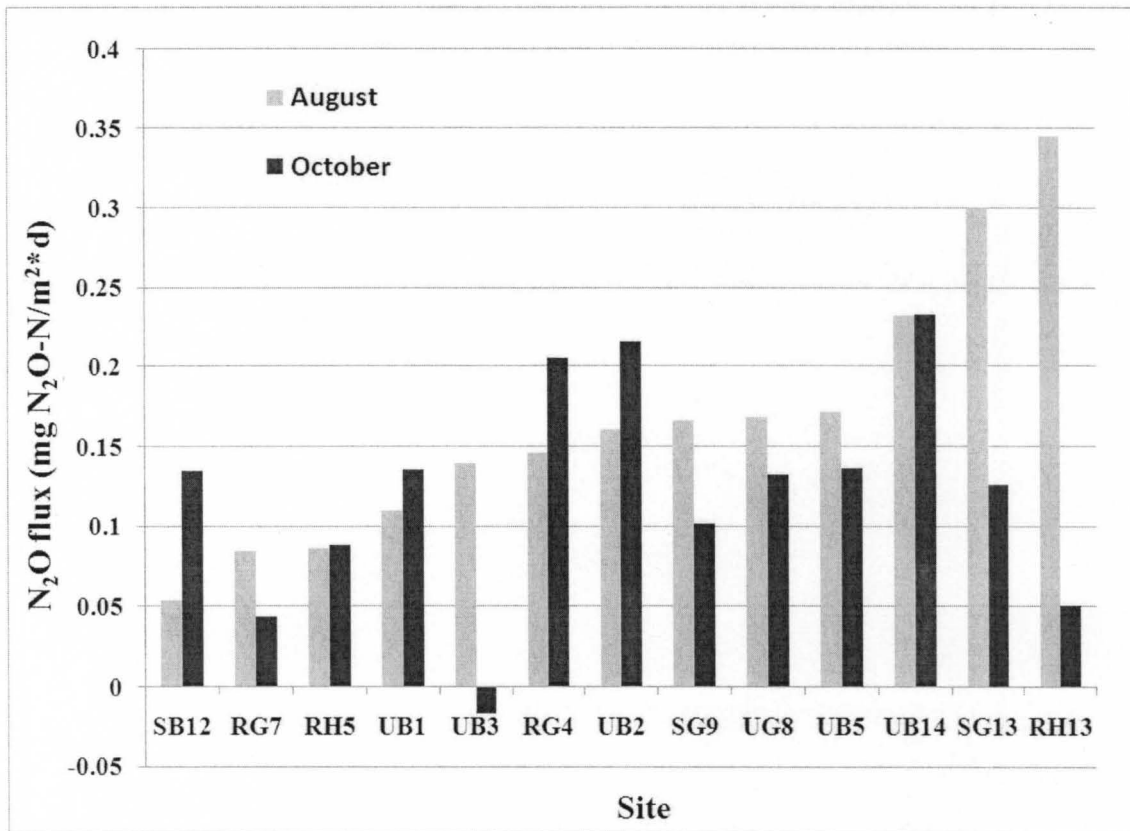


Figure 4-19: Nitrous oxide flux from static chambers along riparian sites in three watersheds spanning an urban-to-rural gradient. Sites were ordered from low to high N₂O flux from left to right. Site names reflect the urban-to-rural gradient and the watershed in which they are located. The first letter in the site name reflects the urban-to-rural gradient, where R=rural, S=suburban, and U=urban based on impervious surface at the subcatchment scale. The second letter reflects the watershed in which the site is located, where B=Beargrass, G=Goose, and H=South Fork Harrods creek watershed. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover.

CHAPTER 5: CONCLUSIONS

Woody plant and groundcover (herbs and woody vines) assemblages in riparian areas in Metro Louisville were both affected by land use (urban, suburban, rural) measured using degree of impervious surface cover at catchment and smaller scales. Urban riparian plant assemblages were dominated by exotic species more so than those alongside rural streams where forests were a more dominant land-cover type. The exotic, invasive shrub *L. maackii* was the most prevalent woody species, and the most abundant species in urban woody plant communities. *E. fortunei*, an exotic woody vine, dominated the herb and woody vine plant communities in urban riparian areas. Not surprisingly, species densities of *L. maackii* and % cover of *E. fortunei* were highly correlated ($r^2 = 0.66$). Conversely, native species were more likely found in riparian assemblages surrounded by forest cover at the catchment scale. The most common native woody species in forested riparian areas were *A. triloba* (understory tree) and *L. benzoin* (shrub). Wetland species (woody and herbaceous groundcover species) also demonstrated a greater affinity for forested riparian sites than for urban ones. As these species are more commonly associated with wetter environmental conditions, a sustained moisture gradient is implied along this urban-to-rural gradient of riparian sites, with urban being drier and rural sites being lower. This pattern of drier soils is expected and was found in 2008 when soils in urban sites were found to be drier in summer and fall than in suburban and rural sites.

However, surprisingly, given these wetland species associations with rural site

groupings, streambank height adjacent to the sites displayed no obvious associations with site assemblage composition even though the highest streambanks (and presumably less flooded and drier soils) were found in urban areas. This demonstrates that other factors, including composition of the regional species pool and arrival at sites, and human riparian management and other anthropogenic factors also affect riparian plant composition. Variation in species diversity indices with land use was greater for groundcover herbs and woody vines as opposed to woody tree and shrub species. I found that riparian groundcover species diversity decreased with increasing percent impervious surface coverage within a 1-km site radius. Groundcover species diversity also decreased with increasing lateral distance from the stream, and increased with increasing upstream catchment area in suburban riparian zones. The presence of exotic invasive species was also found to contribute to the reduction in plant species diversity for the groundcover community in more urban sites.

Since soil water levels differed among urban, suburban, and rural riparian zones, I expected soil gas fluxes of three important greenhouse gases (CO_2 , CH_4 and N_2O) to differ with land use as well. Carbon dioxide (CO_2) flux rates were related to proportion of impervious surface within a 1-km proximity, but the strength of this relationship varied by season. Carbon dioxide flux rates increased with increasing soil temperatures across all sites (Fig. 4-1b & 4-6), but site differences in soil temperatures played the greatest role in explaining differences in CO_2 release during the hottest months and into fall, which is also when the effects of land-use manifested itself more clearly. Greater proportions of impervious surface within a 1-km radius and higher soil temperatures resulted in greater CO_2 release from the soil (Tables 4-5 & 4-6).

Methane and nitrous oxide flux patterns were more complex to explain, since net flux of each is determined by both aerobic and anaerobic processes. Differences in methane flux among sites in relation to their soil and site differences were more obvious in summer and fall. Fall net methane uptake rates were negatively correlated with C:N ratios (Fig. 4-18), which may be attributable to greater methane-oxidizing nitrifier activity (Jones and Morita 1983, Bedard and Knowles 1989). Soil moisture levels also affect microbial activity by affecting the oxygenation levels of soil pores. Across these riparian sites, higher soil moisture values ($> \sim 0.35$ -g soil water/g dry soil) were correlated with a shift from net methanotrophy (net methane uptake by soils) to net methanogenesis (net methane release from soil), because anaerobic, wet soils likely increased methanogen activity. Methane uptake, however, did not seem to be affected in a linear fashion by the range of soil moisture levels below ~ 0.35 -g soil water/g dry soil (Fig. 4-17 & 4-18b). Nitrous oxide efflux rates showed a strong seasonal, and variable, response between hot, dry August and cooler October (Fig. 4-19). No measured environmental site differences could explain the N_2O flux variation observed among sites. However, release of N_2O is the product or by-product of two different soil processes: anaerobic denitrification and aerobic nitrification. The variable response of N_2O release from the soil is likely due to the synergism between these two processes.

The greenhouse gas response in urban, suburban, and rural riparian zones is variable (Fig. 4-6, 4-11, & 4-19). While urban riparian soils exhibited some of the highest CO_2 fluxes from the soil during summer, some of that debt was paid back by way of net CH_4 uptake. Conversely, rural riparian soils exhibited lower CO_2 flux rates; however, net methane emissions into the atmosphere occurred throughout spring and early summer.

Carbon dioxide flux from suburban riparian soils was also relatively lower during summer, but forested suburban riparian zones were the least wide among the three land-use types, effectively reducing the carbon-savings. These findings complicate the determination of greenhouse gas budgets; however, considerations based on land-use are essential for accurate determination of climate alterations and formulating the appropriate response.

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Appendix Table 1: Tree species within 41 riparian sites in Jefferson and Oldham counties in Kentucky. Site names refer to the watershed in which the site was located, where B = Middle Fork Beargrass, G = Goose, and H = South Fork Harrods Creek watersheds. Relative importance values (IV) were calculated from relative density and basal area by land cover category. Land use categories for each site were determined by the proportion of impervious surface (IS) within the subcatchment in which the site was located. Urban $\geq 30\%$ IS, Rural $\leq 10\%$ IS, and Suburban is between 10 and 30% IS.

Site	Land use category	Tree Species Names	Density (stems/ha)	Basal area (m ² /ha)	IV
G2	Rural	<i>Acer saccharum</i> Marsh.	267	18.1	86
G2	Rural	<i>Fraxinus americana</i> L.	67	0.0	9
G2	Rural	<i>Maclura pomifera</i> (Raf.) Schneid.	400	18.4	105
G7	Rural	<i>Acer negundo</i> L.	200	1.1	200
H1	Rural	<i>Acer negundo</i> L.	200	10.7	48
H1	Rural	<i>Asimina triloba</i> (L.) Dunal	100	0.2	13
H1	Rural	<i>Cercis canadensis</i> L.	200	1.5	28
H1	Rural	<i>Juglans nigra</i> L.	200	34.6	98
H1	Rural	<i>Ulmus rubra</i> Muhl.	100	0.3	13
H11	Rural	<i>Acer negundo</i> L.	100	4.9	30
H11	Rural	<i>Fraxinus americana</i> L.	600	9.1	108
H11	Rural	<i>Platanus occidentalis</i> L.	100	13.5	62
H12	Rural	<i>Acer negundo</i> L.	50	0.1	26
H12	Rural	<i>Fraxinus americana</i> L.	50	0.4	31
H12	Rural	<i>Juglans nigra</i> L.	50	5.4	115
H12	Rural	<i>Prunus serotina</i> Ehrh.	50	0.2	28
H14	Rural	<i>Acer negundo</i> L.	350	3.4	111
H14	Rural	<i>Juglans nigra</i> L.	50	10.9	89
H5	Rural	<i>Aesculus glabra</i> Willd.	100	1.0	44
H5	Rural	<i>Asimina triloba</i> (L.) Dunal	100	0.1	23
H5	Rural	<i>Celtis occidentalis</i> L.	100	0.3	27
H5	Rural	<i>Maclura pomifera</i> (Raf.) Schneid.	200	2.7	106
H6	Rural	<i>Acer saccharum</i> Marsh.	277	2.2	161
H6	Rural	<i>Cornus florida</i> L.	139	0.1	39
H7	Rural	<i>Asimina triloba</i> (L.) Dunal	1211	1.6	153
H7	Rural	<i>Carpinus caroliniana</i> Walter	173	0.4	29
H7	Rural	<i>Cornus florida</i> L.	173	0.1	18
H8	Rural	<i>Acer negundo</i> L.	50	1.3	13
H8	Rural	<i>Acer saccharum</i> Marsh.	50	1.5	14

Appendix Table 1 (cont.)					
H8	Rural	<i>Aesculus glabra</i> Willd.	300	0.6	39
H8	Rural	<i>Celtis occidentalis</i> L.	100	1.8	22
H8	Rural	<i>Juglans nigra</i> L.	50	8.9	57
H8	Rural	<i>Quercus alba</i> L.	50	2.8	22
H8	Rural	<i>Ulmus rubra</i> Muhl.	250	0.7	33
H9	Rural	<i>Juniperus virginiana</i> L.	1000	38.6	200
B11	Suburban	<i>Celtis occidentalis</i> L.	358	0.2	63
B11	Suburban	<i>Staphylea trifolia</i> L.	717	0.5	137
B12	Suburban	<i>Acer negundo</i> L.	400	8.4	88
B12	Suburban	<i>Tilia americana</i> L.	500	10.7	112
G1	Suburban	<i>Celtis occidentalis</i> L.	100	0.1	22
G1	Suburban	<i>Juglans nigra</i> L.	100	1.7	89
G1	Suburban	<i>Platanus occidentalis</i> L.	100	0.4	36
G1	Suburban	<i>Ulmus rubra</i> Muhl.	200	0.3	52
G10	Suburban	<i>Acer negundo</i> L.	400	23.2	200
G11	Suburban	<i>Acer negundo</i> L.	600	2.9	105
G11	Suburban	<i>Celtis occidentalis</i> L.	100	0.9	24
G11	Suburban	<i>Fraxinus americana</i> L.	300	1.9	60
G11	Suburban	<i>Juglans nigra</i> L.	100	0.1	10
G12	Suburban	<i>Acer saccharum</i> Marsh.	300	0.6	53
G12	Suburban	<i>Fraxinus americana</i> L.	300	19.3	147
G13	Suburban	<i>Acer negundo</i> L.	100	5.6	130
G13	Suburban	<i>Ulmus rubra</i> Muhl.	100	1.4	70
G3	Suburban	<i>Acer saccharinum</i> L.	400	12.0	54
G3	Suburban	<i>Morus alba</i> L.	300	8.0	38
G3	Suburban	<i>Ulmus rubra</i> Muhl.	1000	19.0	108
G6	Suburban	<i>Acer negundo</i> L.	100	2.8	81
G6	Suburban	<i>Aesculus glabra</i> Willd.	100	0.2	25
G6	Suburban	<i>Asimina triloba</i> (L.) Dunal	200	0.7	56
G6	Suburban	<i>Ulmus rubra</i> Muhl.	100	0.8	38
G8	Suburban	<i>Acer negundo</i> L.	145	0.8	23
G8	Suburban	<i>Acer saccharum</i> Marsh.	291	5.0	57
G9	Suburban	<i>Acer negundo</i> L.	1000	9.1	200
H10	Suburban	<i>Acer saccharum</i> Marsh.	300	6.9	111
H10	Suburban	<i>Fraxinus americana</i> L.	100	6.6	69
H10	Suburban	<i>Ulmus rubra</i> Muhl.	100	0.1	21
H2	Suburban	<i>Acer negundo</i> L.	200	13.0	81
H2	Suburban	<i>Acer saccharum</i> Marsh.	400	8.8	84
H2	Suburban	<i>Asimina triloba</i> (L.) Dunal	100	0.2	12
H2	Suburban	<i>Celtis occidentalis</i> L.	100	0.1	12

Appendix Table 1 (cont.)					
H2	Suburban	<i>Tilia americana</i> L.	100	0.1	11
H4	Suburban	<i>Celtis occidentalis</i> L.	263	0.2	24
H4	Suburban	<i>Cornus alternifolia</i> L. f.	526	2.8	114
H4	Suburban	<i>Fraxinus pennsylvanica</i> Marsh.	263	0.2	26
H4	Suburban	<i>Ulmus rubra</i> Muhl.	263	0.6	35
B1	Urban	<i>Acer negundo</i> L.	1000	40.6	190
B1	Urban	<i>Morus rubra</i> L.	50	0.1	5
B1	Urban	<i>Quercus rubra</i> L.	50	0.1	5
B13	Urban	<i>Morus alba</i> L.	654	11.8	200
B14	Urban	<i>Acer negundo</i> L.	400	8.7	58
B14	Urban	<i>Aesculus glabra</i> Willd.	100	6.7	21
B14	Urban	<i>Fraxinus americana</i> L.	200	10.2	38
B14	Urban	<i>Platanus occidentalis</i> L.	100	40.7	72
B14	Urban	<i>Ulmus rubra</i> Muhl.	100	0.2	11
B2	Urban	<i>Celtis occidentalis</i> L.	500	32.6	116
B2	Urban	<i>Platanus occidentalis</i> L.	100	67.9	84
B3	Urban	<i>Acer saccharum</i> Marsh.	400	0.7	33
B3	Urban	<i>Carya cordiformis</i> (Wangenh.) K. Koch	300	17.5	72
B3	Urban	<i>Celtis occidentalis</i> L.	500	10.8	68
B3	Urban	<i>Juglans nigra</i> L.	100	7.1	27
B4	Urban	<i>Acer negundo</i> L.	700	5.0	107
B4	Urban	<i>Acer saccharum</i> Marsh.	100	8.0	58
B4	Urban	<i>Morus rubra</i> L.	100	4.0	34
B5	Urban	<i>Acer negundo</i> L.	200	9.0	200
B6	Urban	<i>Fraxinus pennsylvanica</i> Marsh.	515	1.5	19
B6	Urban	<i>Morus alba</i> L.	1804	20.3	92
B6	Urban	<i>Platanus occidentalis</i> L.	258	19.6	41
B6	Urban	<i>Prunus cerasifera</i> Ehrh.	258	3.1	13
B6	Urban	<i>Ulmus rubra</i> Muhl.	258	15.9	35
B7	Urban	<i>Betula populifolia</i> Marsh.	33	0.0	6
B7	Urban	<i>Fraxinus americana</i> L.	167	1.3	55
B7	Urban	<i>Juglans nigra</i> L.	133	3.0	83
B7	Urban	<i>Maclura pomifera</i> (Raf.) Schneid.	67	0.5	22
B7	Urban	<i>Thuja occidentalis</i> L.	33	0.0	6
B7	Urban	<i>Ulmus rubra</i> Muhl.	133	0.2	27
B8	Urban	<i>Juglans nigra</i> L.	200	11.5	200
B9	Urban	<i>Picea pungens</i> Engelm.	100	0.3	200
G8	Urban	<i>Celtis occidentalis</i> L.	291	24.0	120

Appendix Table 2: Wetland indicator status and species provenance codes used in this study and definitions (U. S. Department of Agriculture and Natural Resources Conservation Service 2009, 2010).

*Est. prob. – estimated probability.

Code	Term	Meaning
OBL	Obligate	Occurs almost always (est. prob*. 99%) under natural conditions in wetlands
FACW	Facultative wetland	Usually occurs in wetlands (est. prob. 67%-99%)
FAC	Facultative	Equally likely to occur in wetlands or non-wetlands (est. prob. 34%-66%)
FACU	Facultative upland	Usually occurs in non-wetlands (est. prob. 67%-99%)
UPL	Upland	Occurs in wetlands in another region, but occurs almost always (est. prob. 99%) in non-wetlands under natural conditions
NI	No indicator	Insufficient information available to determine an indicator status
N	Native	Plants that have developed over hundreds of years in an area – pre-European settlement
E	Exotic	Plants that have been introduced with human assistance into an area in which it was not found pre-European-settlement
Z	Naturalized	Exotic species that do not need human assistance to reproduce, thereby maintaining themselves over an unspecified period of time

Appendix Table 3: National Land Cover Database (U. S. Department of the Interior and U. S. Geological Survey 2008) land-cover classification categories, definitions, and examples. Table reproduced from U. S. Dept. of Interior and U.S. Geological Survey (2008)

Classification	Definition
Developed, Open Space	Includes areas with a mixture of some constructed structures, but mostly vegetation in the form of lawn grasses. Impervious surfaces account for less than 20% of total cover (e. g. park, golf course).
Developed, Low Intensity	Includes areas with a mixture of constructed structures and vegetation. Impervious surfaces account for 20-49% of total cover (e. g. single family housing).
Developed, Medium Intensity	Includes areas with a mixture of constructed structures and vegetation. Impervious surfaces account for 50-79% of the total cover (e. g. single family housing)
Developed, High Intensity	Includes highly developed areas where people reside or work in high numbers. Impervious surfaces account for 80-100 percent of the total cover (Apartments, Commercial/Industrial).
Deciduous Forest	Areas dominated by trees generally greater than 5 meters tall, and greater than 20% of total vegetation cover. More than 75% of the tree species shed foliage simultaneously in response to seasonal change.
Mixed Forest	Areas dominated by trees generally greater than 5 meters tall, and greater than 20% of total vegetation cover. Neither deciduous nor evergreen species are greater than 75% of total tree cover.
Herbaceous	Areas dominated by graminoid or herbaceous vegetation, generally greater than 80% of total vegetation. These areas are not subject to intensive management such as tilling, but can be utilized for grazing.
Pasture	Areas of grasses, legumes, or grass-legume mixtures planted for livestock grazing or the production of seed or hay crops, typically on a perennial cycle. Pasture/hay vegetation accounts for greater than 20% of total vegetation.
Cultivated Crops	Areas used for the production of annual and perennial woody crops. Crop vegetation accounts for greater than 20% of total vegetation (e. g. corn, orchard)
Wetlands	Areas where soil or substrate is periodically saturated with or covered with water.

Appendix Table 4: Tree, sapling, tree seedling, and shrub species within 41 riparian sites in Jefferson and Oldham counties in Kentucky. Sites were located in Middle Fork Beargrass, Goose, and South Fork Harrods Creek watersheds. Species codes and wetland indicator status were obtained from USDA PLANTS Database (2010).

Species sampled across all sites (n=41)

Trees	Code	Wetland Indicator Status
<i>Acer negundo</i> L.	ACNE2	FAC
<i>Acer saccharinum</i> L.	ACSA2	FACW
<i>Acer saccharum</i> Marsh.	ACSA3	FACU
<i>Aesculus glabra</i> Willd.	A EGL	FACU
<i>Asimina triloba</i> (L.) Dunal	ASTR	FACU
<i>Betula populifolia</i> Marsh.	BEPO	FAC
<i>Carpinus caroliniana</i> Walter	CACA18	FAC
<i>Carya cordiformis</i> (Wangenh.) K. Koch	CACO15	FACU
<i>Celtis occidentalis</i> L.	CEOC	FACU
<i>Cercis canadensis</i> L.	CECA4	FACU
<i>Cornus alternifolia</i> L. f.	COAL2	NI
<i>Cornus florida</i> L.	COFL2	FACU
<i>Fraxinus americana</i> L.	FRAM2	FACU
<i>Fraxinus pennsylvanica</i> Marsh.	FRPE	FACW
<i>Juglans nigra</i> L.	JUNI	FACU
<i>Juniperus virginiana</i> L.	JUVI	FACU
<i>Maclura pomifera</i> (Raf.) Schneid.	MAPO	UPL
<i>Morus alba</i> L.*	MOAL	UPL
<i>Morus rubra</i> L.	MORU2	FACU
<i>Picea pungens</i> Engelm.	PIPU	NI
<i>Platanus occidentalis</i> L.	PLOC	FACW
<i>Prunus cerasifera</i> Ehrh.**	PRCE2	NI
<i>Prunus serotina</i> Ehrh.	PRSE2	FACU
<i>Quercus alba</i> L.	QUAL	FACU
<i>Quercus rubra</i> L.	QURU	FACU
<i>Staphylea trifolia</i> L.	STTR	FAC
<i>Thuja occidentalis</i> L.	THOC2	FACW
<i>Tilia americana</i> L.	TIAM	FACU
<i>Ulmus rubra</i> Muhl.	ULRU	FAC

**Appendix Table
4 (cont.)**

Saplings	Code	Wetland Indicator Status
<i>Acer negundo</i> L.	ACNE2	FAC
<i>Acer saccharum</i> Marsh.	ACSA3	FACU
<i>Aesculus glabra</i> Willd.	AEGL	FACU
<i>Asimina triloba</i> (L.) Dunal	ASTR	FACU
<i>Carya cordiformis</i> (Wangenh.) K. Koch	CACO15	FACU
<i>Celtis occidentalis</i> L.	CEOC	FACU
<i>Cercis canadensis</i> L.	CECA4	FACU
<i>Cornus sericea</i> L. ssp. <i>sericea</i>	COSES	FACW
<i>Cornus</i> sp.	CORNU	
<i>Fraxinus americana</i> L.	FRAM2	FACU
<i>Fraxinus pennsylvanica</i> Marsh.	FRPE	FACW
<i>Fraxinus quadrangulata</i> Michx.	FRQU	NI
<i>Juglans nigra</i> L.	JUNI	FACU
<i>Maclura pomifera</i> (Raf.) Schneid.	MAPO	UPL
<i>Morus rubra</i> L.	MORU2	FACU
<i>Prunus serotina</i> Ehrh.	PRSE2	FACU
<i>Quercus rubra</i> L.	QURU	FACU
<i>Robinia pseudoacacia</i> L.	ROPS	FAC
<i>Staphylea trifolia</i> L.	STTR	FAC
<i>Thuja occidentalis</i>	THOC2	FACW
<i>Tilia americana</i> L.	TIAM	FACU
<i>Ulmus rubra</i> Muhl.	ULRU	FAC

Tree seedlings	Code	Wetland Indicator Status
<i>Acer negundo</i> L.	ACNE2	FAC
<i>Acer saccharum</i> Marsh.	ACSA3	FACU
<i>Acer</i> sp.	ACER	
<i>Aesculus glabra</i> Willd.	AEGL	FACU
<i>Asimina triloba</i> (L.) Dunal	ASTR	FACU
<i>Betula alleghaniensis</i> Britt.	BEAL2	FAC
<i>Carya cordiformis</i> (Wangenh.) K. Koch	CACO15	FACU
<i>Cercis canadensis</i> L.	CECA4	FACU
<i>Celtis occidentalis</i> L.	CEOC	FACU
<i>Cornus drummondii</i> C.A. Mey.	CODR	FAC

**Appendix Table
4 (cont.)**

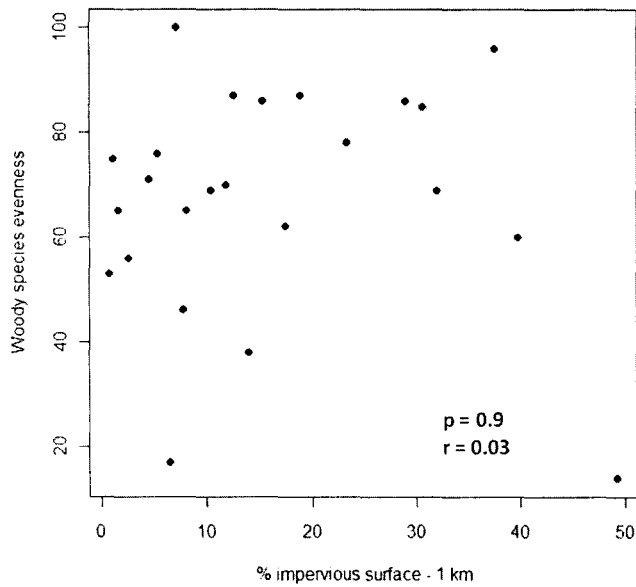
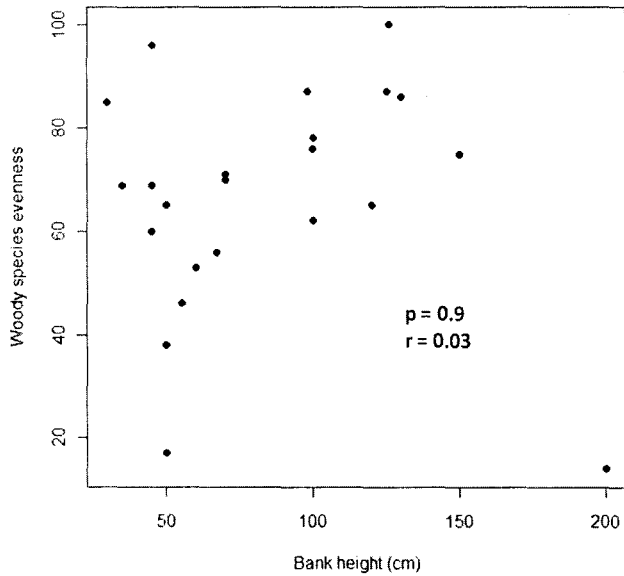
<i>Fraxinus americana</i> L.	FRAM	FACU
<i>Fraxinus pennsylvanica</i> Marsh.	FRPE	FACW
<i>Fraxinus quadrangulata</i> Michx.	FRQU	NI
<i>Fraxinus</i> sp.	FRAXI	
<i>Gleditsia tricanthos</i> L.	GLTR	FAC
<i>Juglans cinerea</i> L.	JUCI	FACU
<i>Liriodendron tulipifera</i> L.	LITU	FACU
<i>Morus rubra</i> L.	MORU2	FACU
<i>Platanus occidentalis</i> L.	PLOC	FACW
<i>Prunus serotina</i> Ehrh.	PRSE2	FACU
<i>Quercus rubra</i> L.	QURU	FACU
<i>Quercus shumardii</i> Buckl.	QUSH	FAC
<i>Robinia pseudoacacia</i> L.	ROPS	FAC
<i>Tilia americana</i> L.	TIAM	FACU
<i>Ulmus rubra</i> Muhl.	ULRU	FAC

Shrubs	Code	Wetland Indicator Status
<i>Amelanchier arborea</i> (Michx. f.) Fern.	AMAR3	FAC
<i>Cornus racemosa</i> Lam.	CORA6	NI
<i>Cornus</i> sp.	CORNU	
<i>Euonymus alatus</i> (Thunb.) Sieb. *	EUAL13	NI
<i>Hibiscus syriacus</i> L.**	HISY	NI
<i>Hydrangea arborescens</i> L.	HYAR	FACU
<i>Hypericum prolificum</i> L.	HYPR	FACU
<i>Lagerstroemia indica</i> L.**	LAIN	NI
<i>Ligustrum sinense</i> Lour.*	LISI	FACU
<i>Lindera benzoin</i> (L.) Blume	LIBE3	FACW
<i>Lonicera maackii</i> (Rupr.) Herder *	LOMA6	NI
<i>Rosa multiflora</i> Thunb. ex Murr.*	ROMU	FACU
<i>Rosa palustris</i> Marsh.	ROPA	OBL
<i>Rubus</i> spp.	RUBUS	
<i>Symphoricarpos orbiculatus</i> Moench	SYOR	UPL
<i>Viburnum acerifolium</i> L.	VIAC	UPL

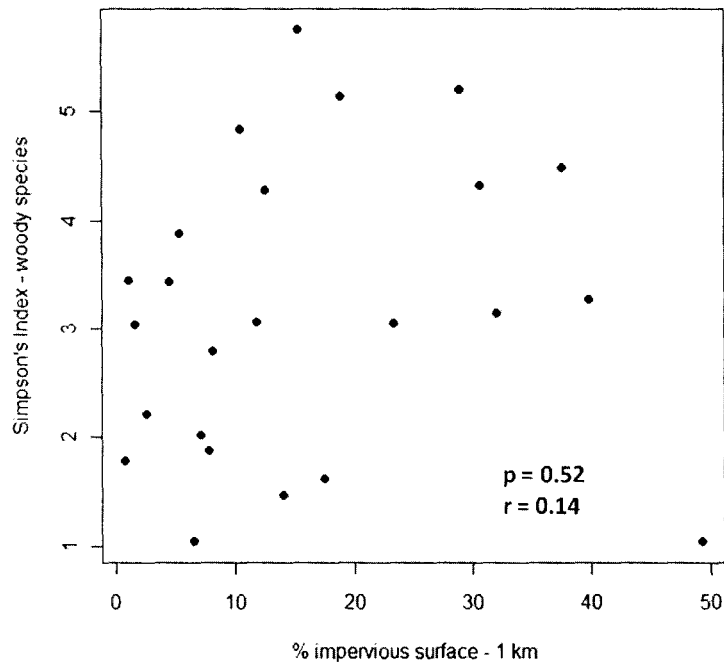
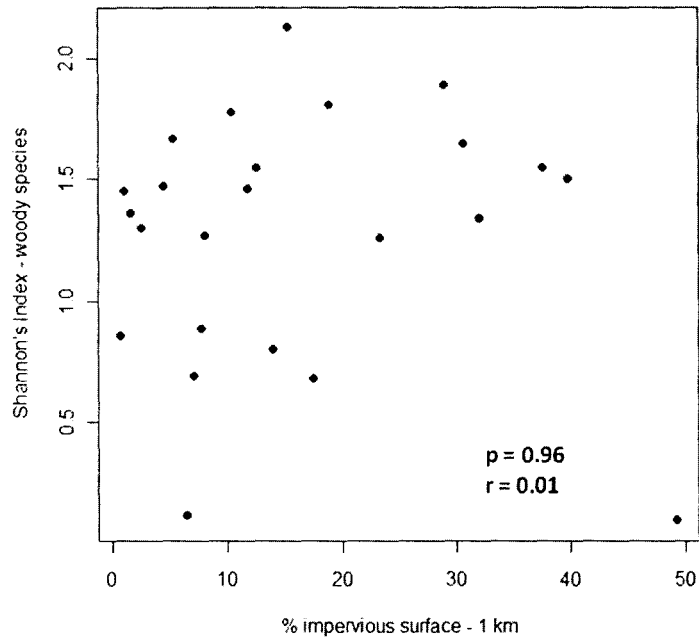
Non-native species indicated with a (*) if naturalized, and a (**) if non-naturalized. Naturalized, exotic species successfully reproduce and maintain viable populations without human intervention.

Plants were identified to genus group if species could not be determined.

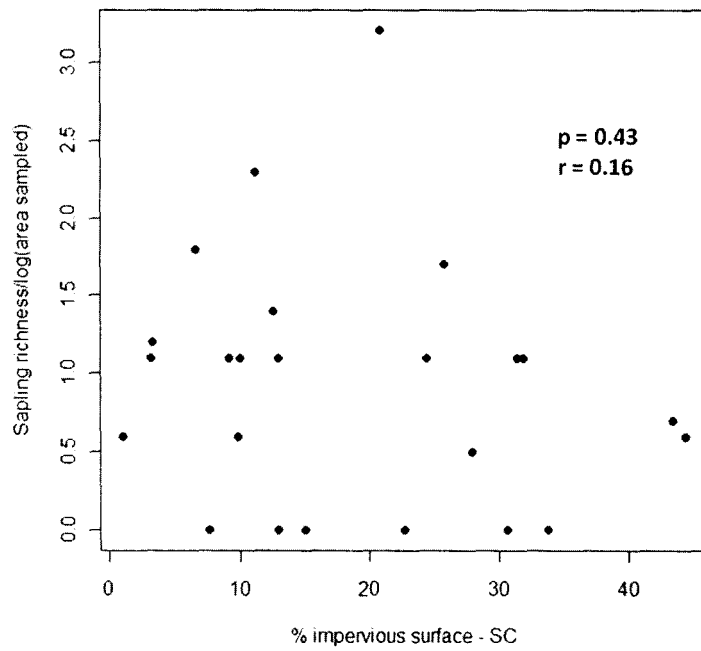
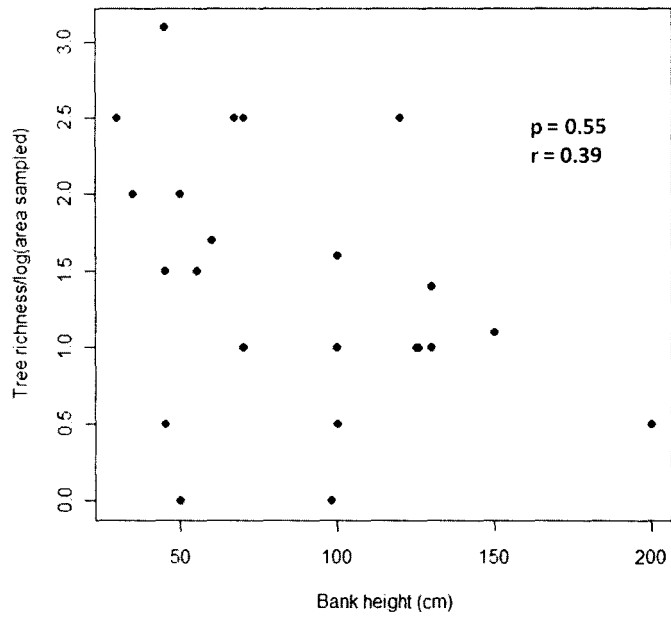
Appendix Figure 5: Woody species abundance and diversity plotted against % impervious surface at the subcatchment (SC) scale and within 1-km of the site as well as streambank height for all sites along first order streams.



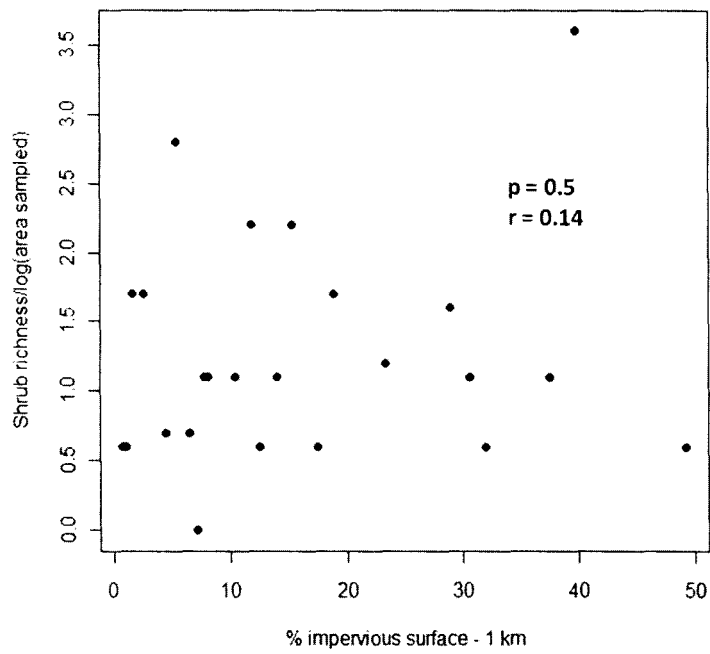
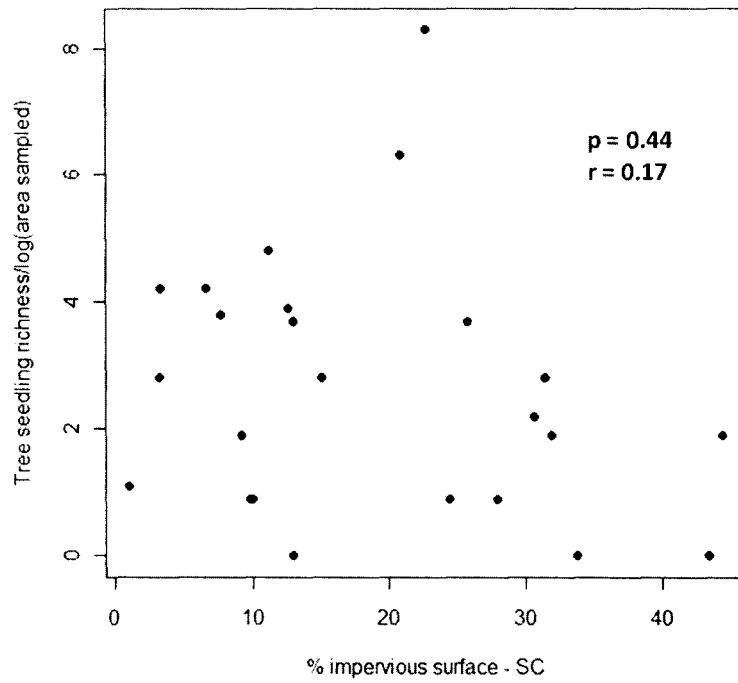
Appendix Figure 5 (continued)



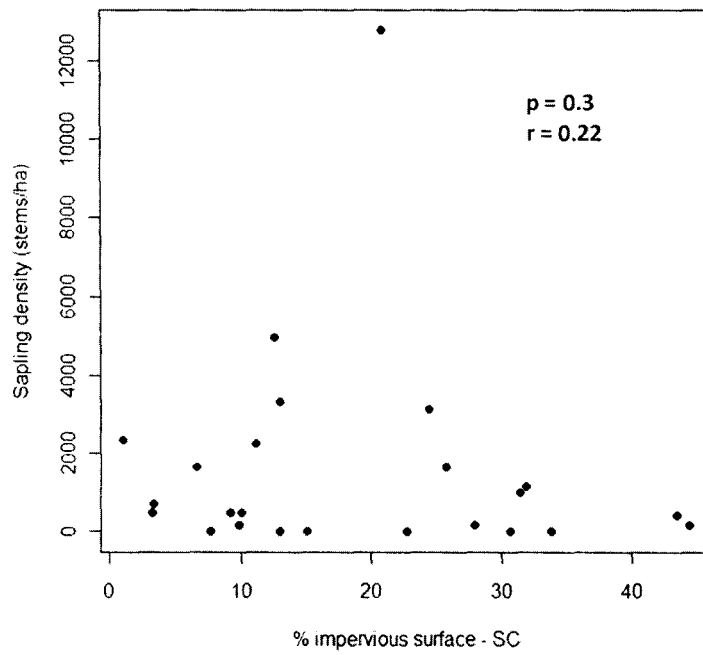
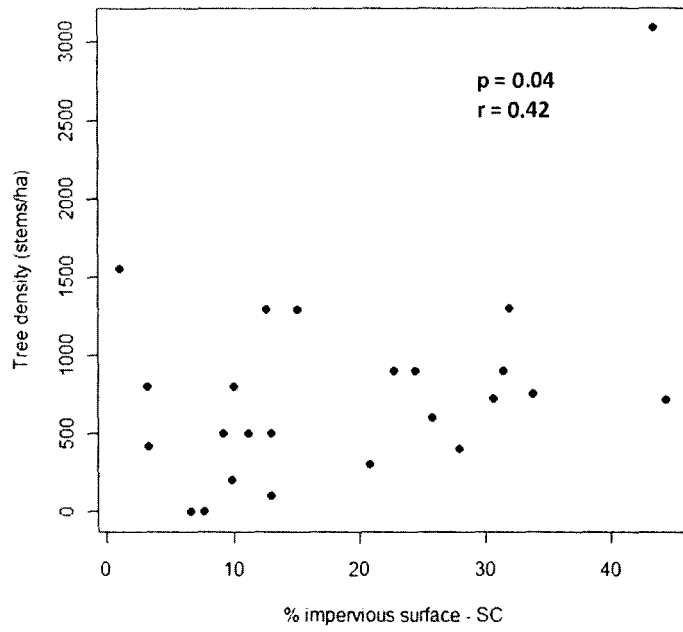
Appendix Figure 5 (continued)



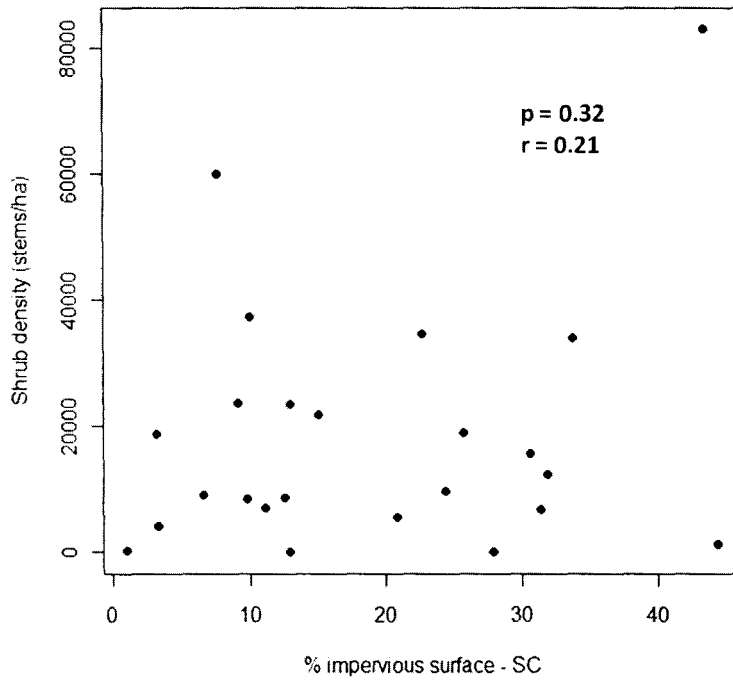
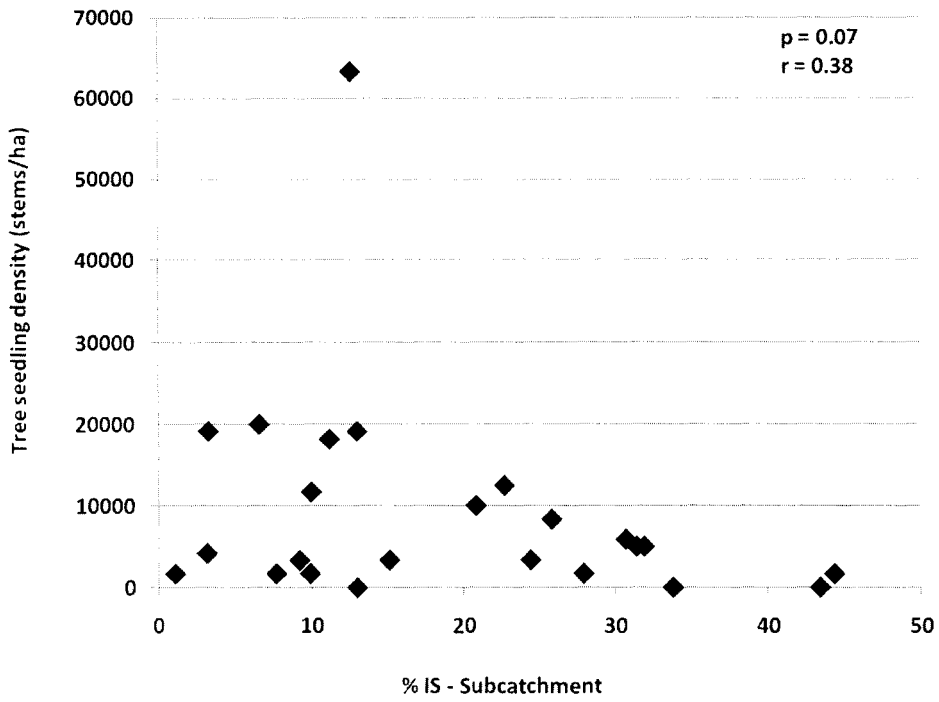
Appendix Figure 5 (continued)



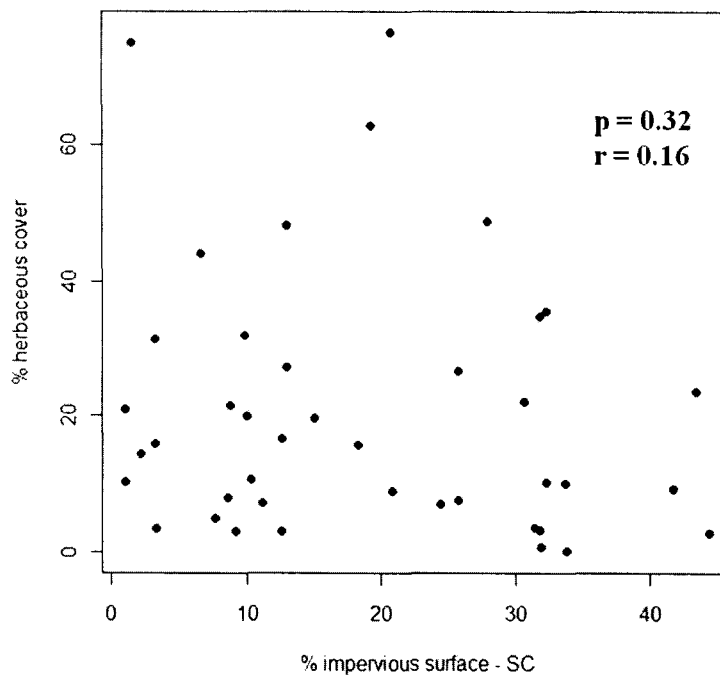
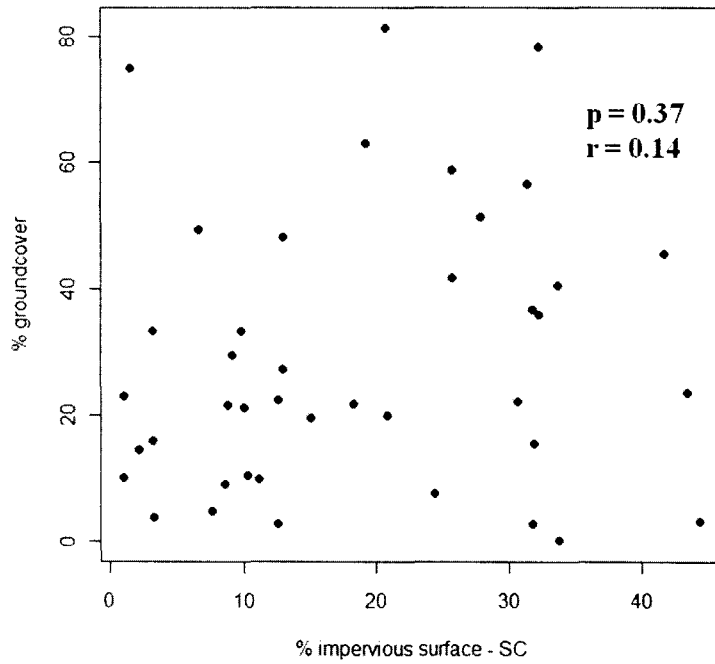
Appendix Figure 5 (continued)



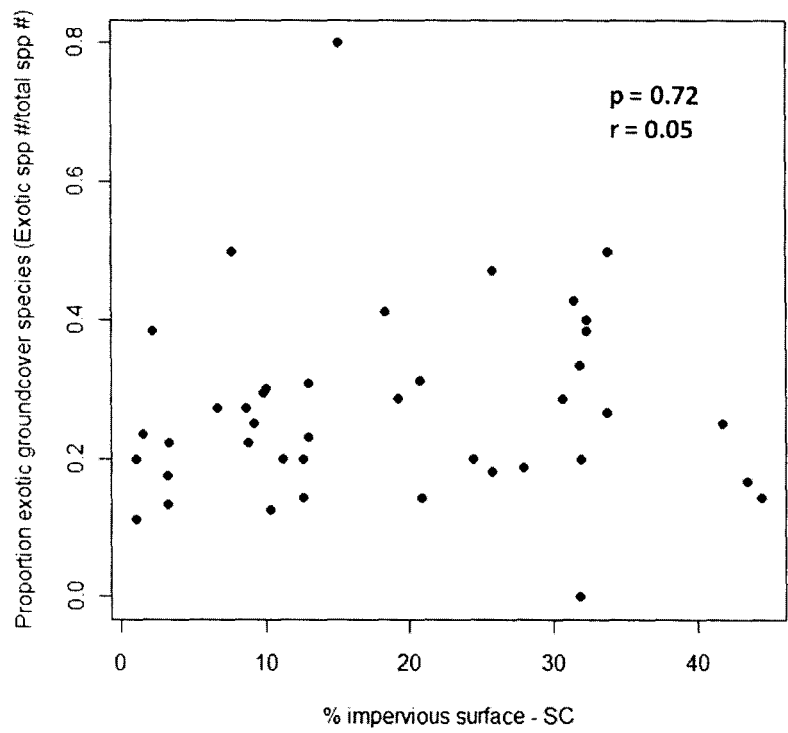
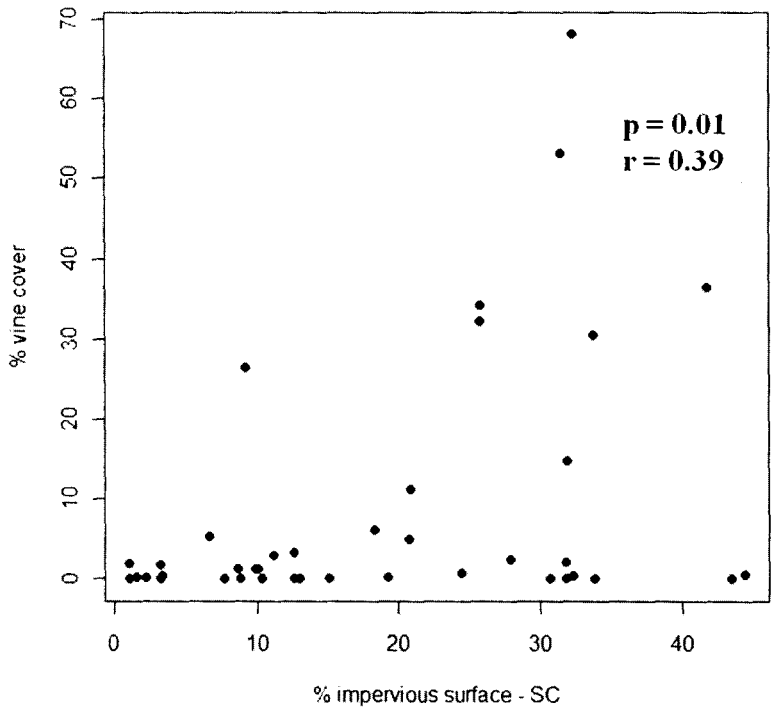
Appendix Figure 5 (continued)



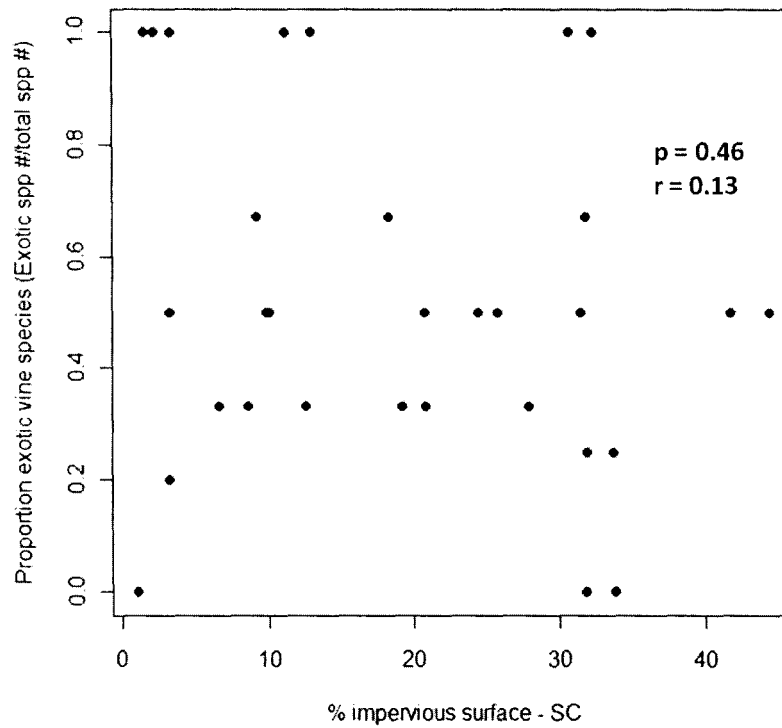
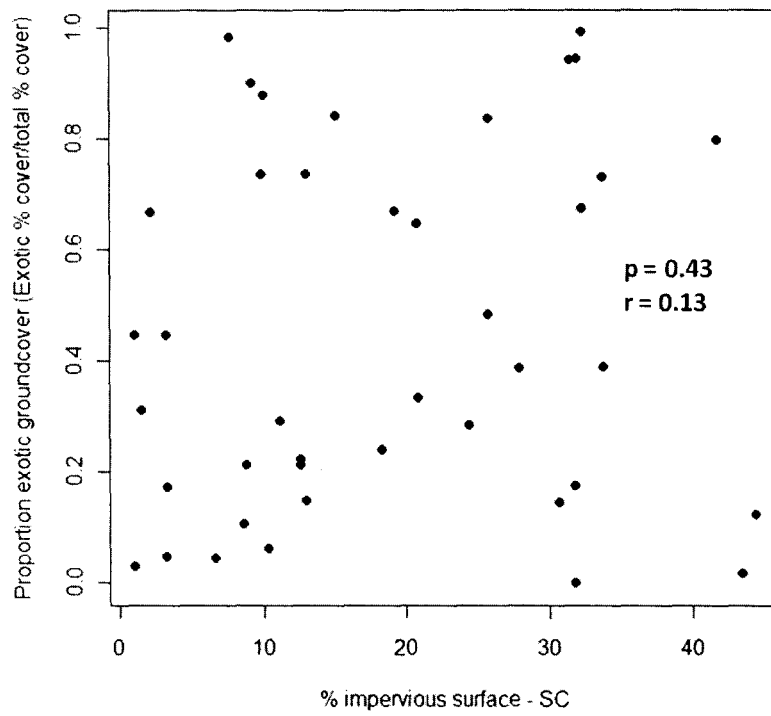
Appendix Figure 6: Total groundcover, herb, and woody vine abundance (% cover), as well as proportion exotic herb and woody vine species richness and abundance, vs. % impervious surface at the subcatchment scale.



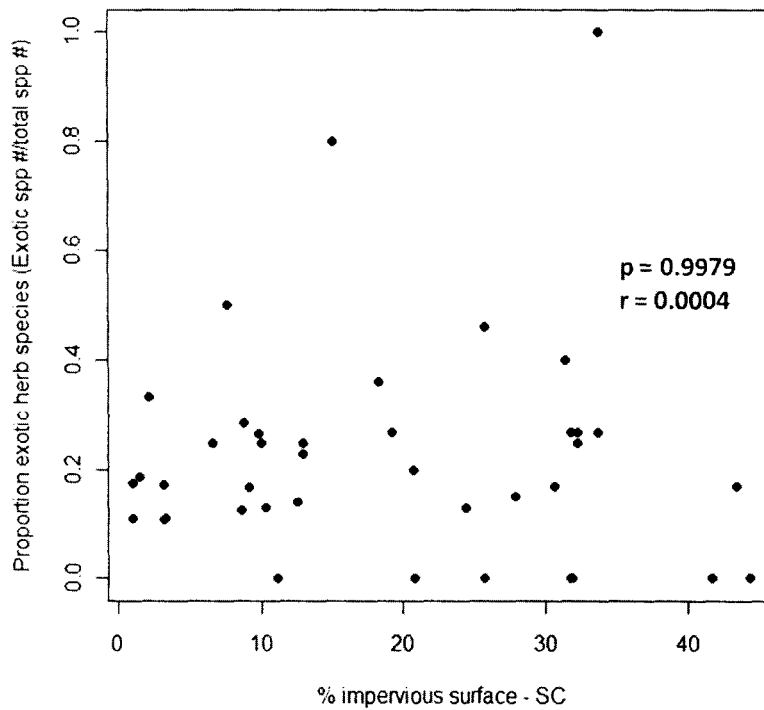
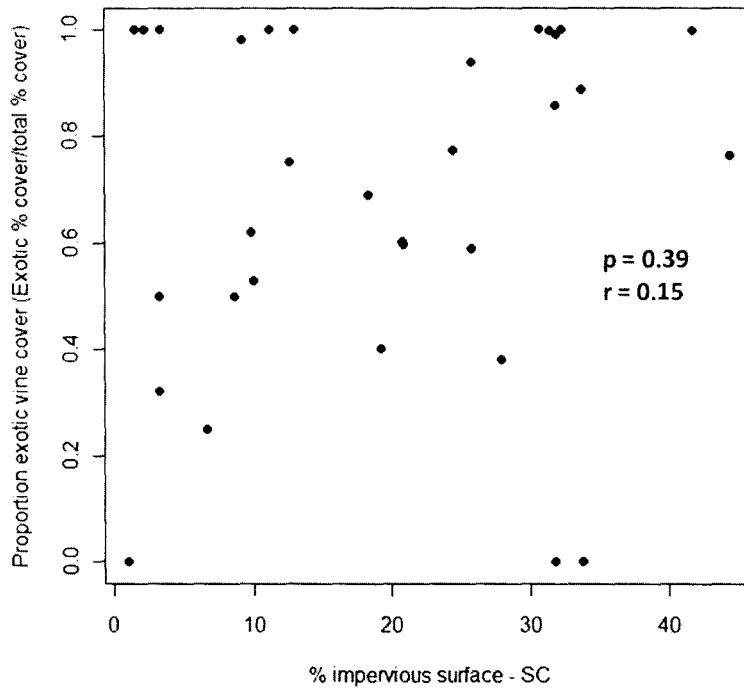
Appendix Figure 6 (continued)



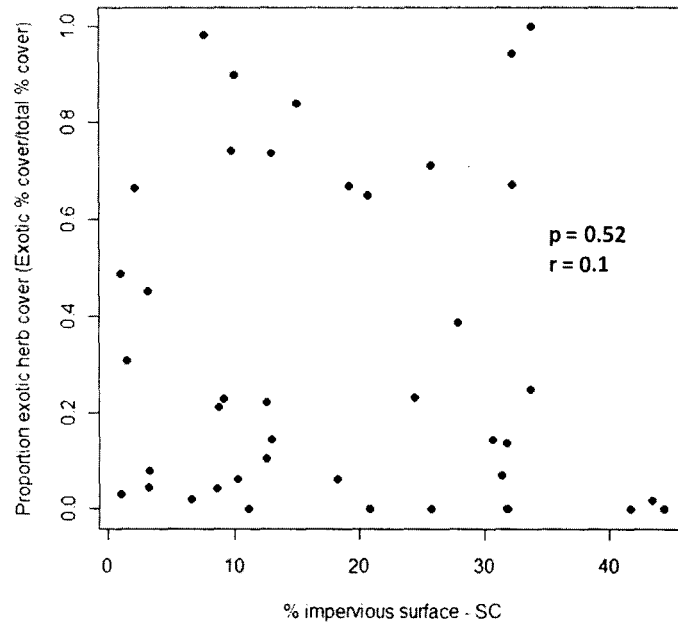
Appendix Figure 6 (continued)



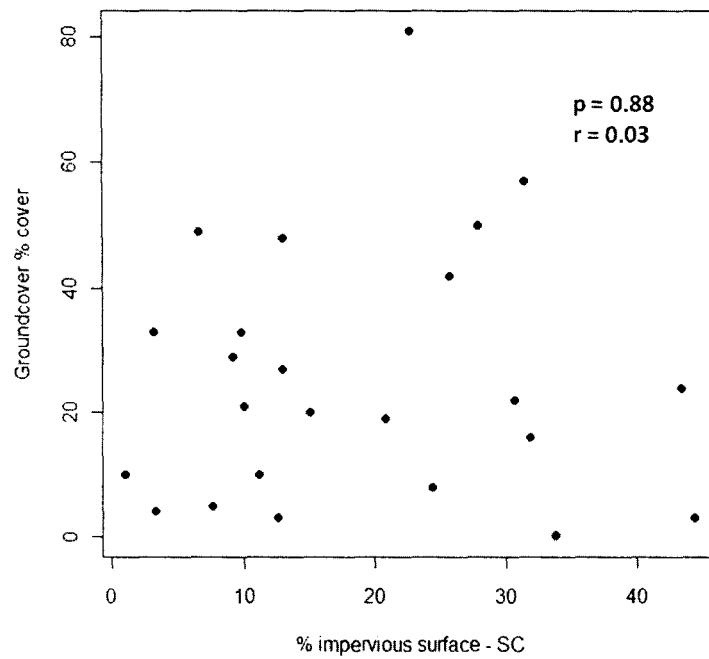
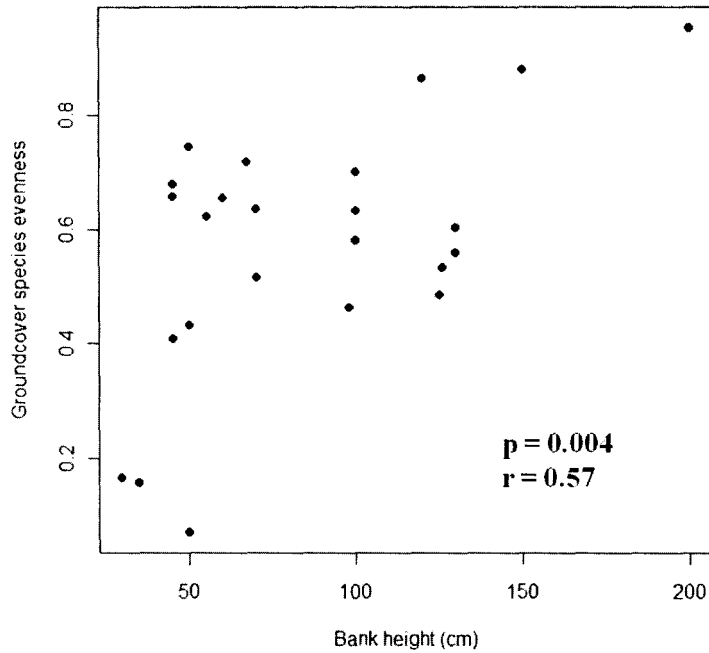
Appendix Figure 6 (continued)



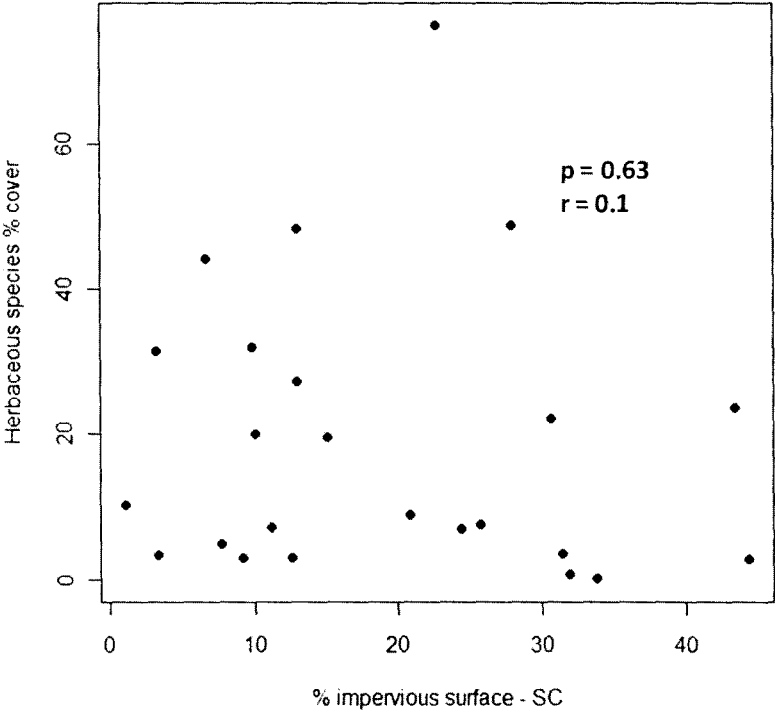
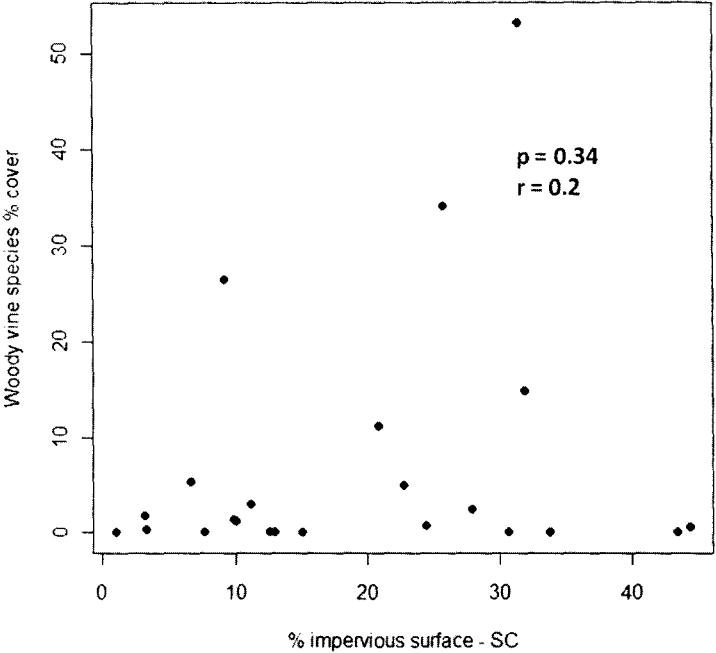
Appendix Figure 6 (continued)



Appendix Figure 7: Groundcover (herb and woody vine) species evenness vs. bank height and groundcover abundances (% cover) vs. % impervious surface at the subcatchment scale within riparian zones along first order streams.



Appendix Figure 7 (cont.)



Appendix Table 8: Sapling species within 41 riparian sites in Jefferson and Oldham counties in Kentucky. Site names refer to the watershed in which the site was located, where B = Middle Fork Beargrass, G = Goose, and H = South Fork Harrods Creek watersheds. Relative importance values (IV) were calculated from relative density and basal area by land cover category. Land use categories for each site were determined by the proportion of impervious surface (IS) within the subcatchment in which the site was located. Urban $\geq 30\%$ IS, Rural $\leq 10\%$ IS, and Suburban is between 10 and 30% IS.

Site	Land-cover category	Sapling Species Names	Freq.	Density (stems/ha)	IV
G2	Rural	<i>Celtis occidentalis</i> L.	0.33	222	133
G2	Rural	<i>Fraxinus americana</i> L.	0.17	111	67
G4	Rural	<i>Acer negundo</i> L.	0.33	208	38
G4	Rural	<i>Celtis occidentalis</i> L.	0.67	625	88
G4	Rural	<i>Robinia pseudoacacia</i> L.	0.33	833	75
H1	Rural	<i>Asimina triloba</i> (L.) Dunal	0.33	333	117
H1	Rural	<i>Ulmus rubra</i> Muhl.	0.33	167	83
H11	Rural	<i>Acer negundo</i> L.	0.33	333	117
H11	Rural	<i>Ulmus rubra</i> Muhl.	0.33	167	83
H12	Rural	<i>Acer negundo</i> L.	0.17	83	100
H12	Rural	<i>Juglans nigra</i> L.	0.17	83	100
H14	Rural	<i>Asimina triloba</i> (L.) Dunal	0.33	500	200
H5	Rural	<i>Aesculus glabra</i> Willd.	0.17	83	50
H5	Rural	<i>Asimina triloba</i> (L.) Dunal	0.50	250	150
H6	Rural	<i>Asimina triloba</i> (L.) Dunal	0.33	476	117
H6	Rural	<i>Fraxinus americana</i> L.	0.33	238	83
H7	Rural	<i>Asimina triloba</i> (L.) Dunal	1.00	2368	200
H8	Rural	<i>Aesculus glabra</i> Willd.	0.33	500	125
H8	Rural	<i>Asimina triloba</i> (L.) Dunal	0.17	83	38
H8	Rural	<i>Ulmus rubra</i> Muhl.	0.17	83	38
B11	Suburban	<i>Carya cordiformis</i> (Wangenh.) K. Koch	0.33	556	29
B11	Suburban	<i>Cornus</i> sp.	0.33	1667	38
B11	Suburban	<i>Fraxinus americana</i> L.	0.33	556	29
B11	Suburban	<i>Staphylea trifolia</i> L.	0.33	10000	103
G10	Suburban	<i>Celtis occidentalis</i> L.	0.33	167	200
G11	Suburban	<i>Acer negundo</i> L.	1.00	2333	148
G11	Suburban	<i>Prunus serotina</i> Ehrh.	0.33	167	26
G12	Suburban	<i>Acer saccharum</i> Marsh.	1.00	1167	130
G12	Suburban	<i>Celtis occidentalis</i> L.	0.33	333	40
G12	Suburban	<i>Fraxinus americana</i> L.	0.33	167	30

Appendix Table 8 (cont.)

G3	Suburban	<i>Fraxinus americana</i> L.	0.33	167	83
G3	Suburban	<i>Ulmus rubra</i> Muhl.	0.33	333	117
G6	Suburban	<i>Aesculus glabra</i> Willd.	0.33	167	30
G6	Suburban	<i>Asimina triloba</i> (L.) Dunal	1.00	3167	170
H10	Suburban	<i>Carya cordiformis</i> (Wangenh.) K. Koch	0.33	175	18
H10	Suburban	<i>Celtis occidentalis</i> L.	0.33	175	18
H10	Suburban	<i>Cercis canadensis</i> L.	0.33	175	18
H10	Suburban	<i>Fraxinus quadrangulata</i> Michx.	0.67	351	35
H10	Suburban	<i>Quercus rubra</i> L.	0.33	175	18
H10	Suburban	<i>Tilia americana</i> L.	0.33	175	18
H10	Suburban	<i>Ulmus rubra</i> Muhl.	1.00	1053	76
H2	Suburban	<i>Acer negundo</i> L.	0.33	167	55
H2	Suburban	<i>Asimina triloba</i> (L.) Dunal	1.00	3000	145
H4	Suburban	<i>Asimina triloba</i> (L.) Dunal	0.67	2917	108
H4	Suburban	<i>Fraxinus pennsylvanica</i> Marsh.	0.67	2083	92
B1	Urban	<i>Acer negundo</i> L.	0.50	833	143
B1	Urban	<i>Morus rubra</i> L.	0.33	167	57
B13	Urban	<i>Acer negundo</i> L.	0.33	278	200
B14	Urban	<i>Celtis occidentalis</i> L.	0.33	667	117
B14	Urban	<i>Fraxinus americana</i> L.	0.33	333	83
B2	Urban	<i>Celtis occidentalis</i> L.	1.00	1500	200
B3	Urban	<i>Acer saccharum</i> Marsh.	0.67	1000	53
B3	Urban	<i>Carya cordiformis</i> (Wangenh.) K. Koch	0.33	167	20
B3	Urban	<i>Celtis occidentalis</i> L.	1.00	4000	127
B6	Urban	<i>Fraxinus quadrangulata</i> Michx.	0.33	417	200
B7	Urban	<i>Acer negundo</i> L.	0.11	56	22
B7	Urban	<i>Celtis occidentalis</i> L.	0.11	56	22
B7	Urban	<i>Cornus sericea</i> L. ssp. <i>sericea</i>	0.11	222	45
B7	Urban	<i>Fraxinus pennsylvanica</i> Marsh.	0.22	278	67
B7	Urban	<i>Maclura pomifera</i> (Raf.) Schneid.	0.11	56	22
B7	Urban	<i>Thuja occidentalis</i>	0.11	56	22
B8	Urban	<i>Morus rubra</i> L.	0.33	167	200

Appendix Table 9: Shrub species within 41 riparian sites in Jefferson and Oldham counties in Kentucky. Site names refer to the watershed in which the site was located, where B = Middle Fork Beargrass, G = Goose, and H = South Fork Harrods Creek watersheds. Relative importance values (IV) were calculated from relative density and basal area by land cover category. Land use categories for each site were determined by the proportion of impervious surface (IS) within the subcatchment in which the site was located. Urban $\geq 30\%$ IS, Rural $\leq 10\%$ IS, and Suburban is between 10 and 30% IS.

Site	Land use category	Shrub Species Names	Freq.	Density (stems/ha)	IV
G2	Rural	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.67	444	36
G2	Rural	<i>Ligustrum sinense</i> Lour.	0.67	2667	53
G2	Rural	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	7778	94
G4	Rural	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	6875	200
G7	Rural	<i>Hydrangea arborescens</i> L.	0.17	1667	45
G7	Rural	<i>Lindera benzoin</i> (L.) Blume	0.67	5000	155
H1	Rural	<i>Lindera benzoin</i> (L.) Blume	1.00	7667	129
H1	Rural	<i>Lonicera maackii</i> (Rupr.) Herder	0.33	1167	29
H11	Rural	<i>Ligustrum sinense</i> Lour.	1.00	16333	174
H11	Rural	<i>Lindera benzoin</i> (L.) Blume	0.33	167	26
H12	Rural	<i>Cornus racemosa</i> Lam.	0.33	583	17
H12	Rural	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	1167	34
H12	Rural	<i>Rubus</i> spp.	0.42	8333	72
H12	Rural	<i>Symphoricarpos orbiculatus</i> Moench	0.33	3333	35
H12	Rural	<i>Viburnum acerifolium</i> L.	0.08	833	9
H13	Rural	<i>Symphoricarpos orbiculatus</i> Moench	1.00	60000	200
H14	Rural	<i>Lindera benzoin</i> (L.) Blume	0.33	667	200
H3	Rural	<i>Symphoricarpos orbiculatus</i> Moench	0.33	167	200
H5	Rural	<i>Hydrangea arborescens</i> L.	0.08	833	13
H5	Rural	<i>Ligustrum sinense</i> Lour.	0.08	1667	22
H5	Rural	<i>Lindera benzoin</i> (L.) Blume	1.00	2167	81
H5	Rural	<i>Lonicera maackii</i> (Rupr.) Herder	0.25	1667	32
H6	Rural	<i>Lindera benzoin</i> (L.) Blume	1.00	4286	200
H7	Rural	<i>Lindera benzoin</i> (L.) Blume	0.33	263	200
H8	Rural	<i>Ligustrum sinense</i> Lour.	0.67	833	33
H8	Rural	<i>Lindera benzoin</i> (L.) Blume	0.83	3750	69
H8	Rural	<i>Lonicera maackii</i> (Rupr.) Herder	0.83	5000	82
H9	Rural	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	7333	200
B11	Suburban	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.33	3333	110

Appendix Table 9 (cont.)

B11	Suburban	<i>Lonicera maackii</i> (Rupr.) Herder	0.33	2222	90
B12	Suburban	<i>Amelanchier arborea</i> (Michx. f.) Fern.	0.33	833	18
B12	Suburban	<i>Ligustrum sinense</i> Lour.	1.00	10167	107
B12	Suburban	<i>Lindera benzoin</i> (L.) Blume	1.00	3167	59
B12	Suburban	<i>Lonicera maackii</i> (Rupr.) Herder	0.33	500	16
G1	Suburban	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.67	833	44
G1	Suburban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	20167	156
G10	Suburban	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.33	167	200
G11	Suburban	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.67	667	41
G11	Suburban	<i>Hibiscus syriacus</i> L.	0.08	1667	26
G11	Suburban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	3833	98
G12	Suburban	<i>Ligustrum sinense</i> Lour.	1.00	3167	72
G12	Suburban	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	9167	87
G14	Suburban	<i>Lonicera maackii</i> (Rupr.) Herder	0.33	3333	100
G14	Suburban	<i>Rubus</i> spp.	0.33	3333	100
G3	Suburban	<i>Cornus racemosa</i> Lam.	0.33	1333	86
G3	Suburban	<i>Ligustrum sinense</i> Lour.	0.33	2333	114
G6	Suburban	<i>Ligustrum sinense</i> Lour.	1.00	20833	174
G6	Suburban	<i>Lindera benzoin</i> (L.) Blume	0.33	167	26
H10	Suburban	<i>Ligustrum sinense</i> Lour.	0.33	175	40
H10	Suburban	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	2281	160
H2	Suburban	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.08	833	19
H2	Suburban	<i>Ligustrum sinense</i> Lour.	0.33	167	26
H2	Suburban	<i>Lindera benzoin</i> (L.) Blume	1.00	5500	155
H4	Suburban	<i>Lindera benzoin</i> (L.) Blume	0.67	2083	200
B1	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	0.33	333	200
B13	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	6111	200
B14	Urban	<i>Lindera benzoin</i> (L.) Blume	1.00	3667	121
B14	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	2333	79
B2	Urban	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.33	333	14
B2	Urban	<i>Hypericum prolificum</i> L.	0.33	167	13
B2	Urban	<i>Ligustrum sinense</i> Lour.	0.67	4667	65
B2	Urban	<i>Lindera benzoin</i> (L.) Blume	0.67	833	30
B2	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	4833	78
B3	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	12333	200
B4	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	13000	200
B5	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	6833	200
B6	Urban	<i>Cornus</i> sp.	0.33	9583	34
B6	Urban	<i>Hibiscus syriacus</i> L.	0.67	7917	45

Appendix Table 9 (cont.)

B6	Urban	<i>Lagerstroemia indica</i> L.	0.33	5000	25
B6	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	25417	96
B7	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	12778	156
B7	Urban	<i>Rosa palustris</i> Marsh.	0.56	1167	44
B8	Urban	<i>Euonymus alatus</i> (Thunb.) Sieb.	0.33	500	118
B8	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	0.08	833	83
B9	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	1.00	6500	200
G8	Urban	<i>Lindera benzoin</i> (L.) Blume	0.67	1000	67
G8	Urban	<i>Lonicera maackii</i> (Rupr.) Herder	0.67	4750	133

Appendix Table 10: Tree seedling species within 41 riparian sites in Jefferson and Oldham counties in Kentucky. Site names refer to the watershed in which the site was located, where B = Middle Fork Beargrass, G = Goose, and H = South Fork Harrods Creek watersheds. Relative importance values (IV) were calculated from relative density and basal area by land cover category. Land use categories for each site were determined by the proportion of impervious surface (IS) within the subcatchment in which the site was located. Urban $\geq 30\%$ IS, Rural $\leq 10\%$ IS, and Suburban is between 10 and 30% IS.

Site	Land use category	Tree seedling species names	Freq.	Density (stems/ha)	IV
G2	Rural	<i>Acer negundo</i> L.	0.0011	2778	89
G2	Rural	<i>Acer saccharum</i> Marsh.	0.0006	556	28
G2	Rural	<i>Celtis occidentalis</i> L.	0.0017	1667	83
G4	Rural	<i>Acer negundo</i> L.	0.4444	6667	67
G4	Rural	<i>Acer saccharum</i> Marsh.	0.4444	5556	61
G4	Rural	<i>Celtis occidentalis</i> L.	0.3333	6667	58
G4	Rural	<i>Platanus occidentalis</i> L.	0.1111	1111	14
H1	Rural	<i>Asimina triloba</i> (L.) Dunal	0.0833	833	45
H1	Rural	<i>Carya cordiformis</i> (Wangenh.) K. Koch	0.0833	833	45
H1	Rural	<i>Fraxinus americana</i> L.	0.0833	833	45
H1	Rural	<i>Ulmus rubra</i> Muhl.	0.0833	1667	65
H11	Rural	<i>Acer negundo</i> L.	0.2500	5000	161
H11	Rural	<i>Asimina triloba</i> (L.) Dunal	0.0833	833	39
H12	Rural	<i>Celtis occidentalis</i> L.	0.1667	833	200
H13	Rural	<i>Acer negundo</i> L.	0.1667	1667	200
H14	Rural	<i>Acer negundo</i> L.	0.0833	417	100
H14	Rural	<i>Aesculus glabra</i> Willd.	0.0833	417	100
H5	Rural	<i>Acer</i> sp.	0.0004	417	25
H5	Rural	<i>Acer negundo</i> L.	0.0004	417	25
H5	Rural	<i>Asimina triloba</i> (L.) Dunal	0.0017	1667	100
H5	Rural	<i>Robinia pseudoacacia</i> L.	0.0004	417	25
H5	Rural	<i>Ulmus rubra</i> Muhl.	0.0004	417	25
H6	Rural	<i>Acer saccharum</i> Marsh.	0.4444	7778	57
H6	Rural	<i>Celtis occidentalis</i> L.	0.2222	2222	22
H6	Rural	<i>Fraxinus americana</i> L.	0.7778	13333	99
H6	Rural	<i>Fraxinus quadrangulata</i> Michx.	0.2222	2222	22
H7	Rural	<i>Asimina triloba</i> (L.) Dunal	0.1250	1250	100
H7	Rural	<i>Juglans cinerea</i> L.	0.1250	1250	100
H8	Rural	<i>Acer</i> sp.	0.1667	833	100
H8	Rural	<i>Aesculus glabra</i> Willd.	0.0833	417	50

Appendix Table 10 (cont.)

H8	Rural	<i>Celtis occidentalis</i> L.	0.0833	417	50
H9	Rural	<i>Fraxinus americana</i> L.	0.5000	8889	140
H9	Rural	<i>Quercus rubra</i> L.	0.3333	2222	60
B10	Suburban	<i>Acer negundo</i> L.	0.0833	833	200
B11	Suburban	<i>Acer negundo</i> L.	0.3333	3333	67
B11	Suburban	<i>Celtis occidentalis</i> L.	0.3333	3333	67
B11	Suburban	<i>Cornus drummondii</i> C.A. Mey.	0.3333	3333	67
B12	Suburban	<i>Acer negundo</i> L.	0.0833	1667	20
B12	Suburban	<i>Acer saccharum</i> Marsh.	0.1667	1667	28
B12	Suburban	<i>Cercis canadensis</i> L.	0.1667	1667	28
B12	Suburban	<i>Celtis occidentalis</i> L.	0.1667	1667	28
B12	Suburban	<i>Fraxinus americana</i> L.	0.1667	1667	28
B12	Suburban	<i>Liriodendron tulipifera</i> L.	0.1667	1667	28
B12	Suburban	<i>Platanus occidentalis</i> L.	0.1667	1667	28
B12	Suburban	<i>Tilia americana</i> L.	0.0833	833	14
G1	Suburban	<i>Acer negundo</i> L.	0.0833	833	67
G1	Suburban	<i>Aesculus glabra</i> Willd.	0.0833	833	67
G1	Suburban	<i>Celtis occidentalis</i> L.	0.0833	833	67
G10	Suburban	<i>Celtis occidentalis</i> L.	0.1667	1667	200
G11	Suburban	<i>Acer negundo</i> L.	0.3333	4167	138
G11	Suburban	<i>Celtis occidentalis</i> L.	0.1667	1667	62
G12	Suburban	<i>Acer negundo</i> L.	0.0833	833	19
G12	Suburban	<i>Acer saccharum</i> Marsh.	0.1667	1667	38
G12	Suburban	<i>Celtis occidentalis</i> L.	0.3333	4167	85
G12	Suburban	<i>Fraxinus americana</i> L.	0.1667	1667	38
G12	Suburban	<i>Liriodendron tulipifera</i> L.	0.0833	833	19
G14	Suburban	<i>Acer negundo</i> L.	0.3333	3333	31
G14	Suburban	<i>Carya cordiformis</i> (Wangenh.) K. Koch	0.3333	3333	31
G14	Suburban	<i>Celtis occidentalis</i> L.	1.0000	23333	138
G3	Suburban	<i>Acer negundo</i> L.	0.0833	833	100
G3	Suburban	<i>Celtis occidentalis</i> L.	0.0833	833	100
G6	Suburban	<i>Acer negundo</i> L.	0.0833	1667	16
G6	Suburban	<i>Acer saccharum</i> Marsh.	0.2500	2500	36
G6	Suburban	<i>Aesculus glabra</i> Willd.	0.0833	833	12
G6	Suburban	<i>Asimina triloba</i> (L.) Dunal	0.6667	14167	135
H10	Suburban	<i>Fraxinus americana</i> L.	0.0909	909	13
H10	Suburban	<i>Fraxinus quadrangulata</i> Michx.	0.2727	3636	43
H10	Suburban	<i>Prunus serotina</i> Ehrh.	0.4545	6364	73
H10	Suburban	<i>Quercus shumardii</i> Buckl.	0.0909	909	13
H10	Suburban	<i>Ulmus rubra</i> Muhl.	0.2727	6364	58

Appendix Table 10 (cont.)

H2	Suburban	<i>Asimina triloba</i> (L.) Dunal	0.1667	2500	142
H2	Suburban	<i>Celtis occidentalis</i> L.	0.0833	833	58
H4	Suburban	<i>Acer saccharum</i> Marsh.	0.1667	6667	35
H4	Suburban	<i>Aesculus glabra</i> Willd.	0.1667	1667	20
H4	Suburban	<i>Asimina triloba</i> (L.) Dunal	0.1667	5000	30
H4	Suburban	<i>Fraxinus pennsylvanica</i> Marsh.	0.6667	18333	115
B1	Urban	<i>Acer negundo</i> L.	0.5833	37083	200
B13	Urban	<i>Fraxinus americana</i> L.	0.1667	1667	200
B14	Urban	<i>Acer negundo</i> L.	0.2500	2500	100
B14	Urban	<i>Aesculus glabra</i> Willd.	0.1667	1667	67
B14	Urban	<i>Platanus occidentalis</i> L.	0.0833	833	33
B2	Urban	<i>Celtis occidentalis</i> L.	0.4167	6667	200
B3	Urban	<i>Celtis occidentalis</i> L.	0.2500	4167	158
B3	Urban	<i>Fraxinus</i> sp.	0.0833	833	42
B5	Urban	<i>Acer negundo</i> L.	0.0833	833	200
B6	Urban	<i>Gleditsia tricanthos</i> L.	0.1667	1667	200
B7	Urban	<i>Acer negundo</i> L.	0.0006	556	25
B7	Urban	<i>Betula alleghaniensis</i> Britt.	0.0003	278	13
B7	Urban	<i>Celtis occidentalis</i> L.	0.0022	3333	124
B7	Urban	<i>Fraxinus americana</i> L.	0.0006	556	25
B7	Urban	<i>Morus rubra</i> L.	0.0003	278	13
B8	Urban	<i>Fraxinus americana</i> L.	0.0833	833	100
B8	Urban	<i>Morus rubra</i> L.	0.0833	833	100
G8	Urban	<i>Acer negundo</i> L.	0.3750	6250	146
G8	Urban	<i>Celtis occidentalis</i> L.	0.1250	2500	54

Appendix Table 11: Herbaceous and vine species within 41 riparian sites in Jefferson and Oldham counties in Kentucky. Sites were located in Middle Fork Beargrass, Goose, and South Fork Harrods Creek watersheds. Species codes and wetland indicator status (WIS) were obtained from USDA PLANTS Database (2010).

Species name	Code	WIS
<i>Ageratina altissima</i> (L.) King & H.E. Robins.	AGAL5	FACU
<i>Ageratina</i> spp. L.	AGERA2	
<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	ALPE4	FACU
<i>Allium vineale</i> L.*	ALVI	FACU
<i>Ampelamus albidus</i> (Nutt.) Britt.	AMAL3	FAC
<i>Ambrosia artemisiifolia</i> L.	AMAR2	FACU
<i>Ambrosia trifida</i> L.	AMTR	FAC
<i>Arundinaria gigantea</i> (Walt.) Muhl.	ARGI	FACW
<i>Aristolochia tomentosa</i> Sims	ARTO3	FAC
<i>Arisaema triphyllum</i> L.	ARTR	FACW
<i>Asarum canadense</i> L.	ASCA	NI
<i>Aureolaria</i> spp. L.	AUREO	
<i>Bidens</i> spp. L.	BIDEN	FACW
<i>Boehmeria cylindrica</i> (L.) Sw.	BOCY	FACW
<i>Carex blanda</i> Dewey	CABL	FAC
<i>Cardamine pensylvanica</i> Muhl. ex Willd.	CAPE3	OBL
<i>Campsis radicans</i> (L.) Seem. ex Bureau	CARA2	FAC
<i>Cirsium arvense</i> (L.) Scop.*	CIAR4	FACU
<i>Commelina communis</i> L.*	COCO3	FAC
<i>Cystopteris tenuis</i> (Michx.) Desv.	CYTE7	NI
<i>Desmodium canescens</i> (L.) DC	DECA8	NI
<i>Desmodium</i> spp. L.	DESMO	
<i>Dichanthelium clandestinum</i> (L.) Gould	DICL	FAC
<i>Duchesnea indica</i> (Andr.) Focke*	DUIN	FACU
<i>Elephantopus carolinianus</i> Raeusch.	ELCA3	FACU
<i>Elymus riparius</i> Wieg.	ELRI	FACW
<i>Elymus</i> spp. L.	ELYMU	
<i>Erigeron</i> spp. L.	ERIGE2	FACU
<i>Erigeron philadelphicus</i> L.	ERPH	FACU
<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	EUFO5	NI
<i>Galium aparine</i> L.	GAAP2	FACU

**Appendix
Table 11
(cont.)**

<i>Geum vernum</i> (Raf.) Torr. & Gray	GEVE	FACU
<i>Glechoma hederacea</i> L.*	GLHE2	FACU
<i>Hedera helix</i> L.*	HEHE	NI
<i>Helianthus</i> spp. L.	HELIA3	
<i>Hydrophyllum canadense</i> L.	HYCA3	FACU
<i>Impatiens capensis</i> Meerb.	IMCA	FACW
<i>Iodanthus pinnatifidus</i> (Michx.) Steud.	IOPI	FACW
<i>Juncus</i> spp. L.	JUNCU	FACW
<i>Lamium amplexicaule</i> L.	LAAM	NI
<i>Lactuca biennis</i> (Moench) Fern.	LABI	FACU
<i>Lactuca canadensis</i> L.	LACA	FACU
<i>Lonicera japonica</i> Thunb.*	LOJA	FAC
<i>Lysimachia nummularia</i> L.*	LYNU	OBL
<i>Maianthemum racemosum</i> (L.) Link	MARA7	FACU
<i>Microstegium vimineum</i> (Trin.) A. Camus *	MIVI	FAC
<i>Ornithogalum umbellatum</i> L.*	ORUM	FACU
<i>Osmorhiza claytonii</i> (Michx.) C.B. Clarke	OSCL	FACU
<i>Oxalis stricta</i> L.	OXST	UPL
<i>Packera</i> spp. L.	PACKE	
<i>Packera obovata</i> (Muhl. ex Willd.) W.A. Weber & A. Löve	PAOB6	FACU
<i>Parthenocissus quinquefolia</i> (L.) Planch.	PAQU2	FACU
<i>Phytolacca americana</i> L.	PHAM4	FACU
<i>Pilea pumila</i> (L.) Gray	PIPU2	FACW
<i>Plantago rugelii</i> Dcne.	PLRU	FACU
<i>Polygonatum biflorum</i> (Walt.) Ell.	POBI2	FACU
<i>Polygonum</i> spp. L.	POLYG4	
<i>Podophyllum peltatum</i> L.	POPE	FACU
<i>Polygonum virginianum</i> L.	POVI2	FAC
<i>Ranunculus hispidus</i> Michx.	RAHI	FAC
<i>Ruellia caroliniensis</i> (J.F. Gmel.) Steud.	RUCA4	NI
<i>Rudbeckia laciniata</i> L.	RULA3	FACW
<i>Rumex verticillatus</i> L.	RUVE3	OBL
<i>Sanicula canadensis</i> L.	SACA15	UPL
<i>Sanicula</i> spp. L.	SANIC	
<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	SAOD	FACU
<i>Sedum ternatum</i> Michx.	SETE3	NI
<i>Securigera varia</i> (L.) Lassen*	SEVA4	NI
<i>Sicyos angulatus</i> L.	SIAN	FACU

**Appendix
Table 11
(cont.)**

<i>Smilax rotundifolia</i> L.	SMRO	FAC
<i>Solidago canadensis</i> L.	SOCA6	FACU
<i>Sorghum halepense</i> (L.) Pers.*	SOHA	FACU
<i>Solidago</i> spp. L.	SOLID	
<i>Spiraea</i> spp. L.	SPIRA	
<i>Stachys</i> spp. L.	STACH	
<i>Stellaria media</i> (L.) Vill.*	STME2	UPL
<i>Symphyotrichum cordifolium</i> (L.) Nesom	SYCO4	NI
<i>Symphyotrichum lowrieianum</i> (Porter) Nesom	SYLO2	NI
<i>Symphyotrichum</i> spp. L.	SYMPH4	
<i>Symphyotrichum pilosum</i> (Willd.) Nesom	SYPI2	UPL
<i>Symphyotrichum racemosum</i> (Ell.) Nesom	SYRA5	FACW
<i>Taraxacum officinale</i> G.H. Weber ex Wiggers*	TAOF	FACU
<i>Thalictrum thalictroides</i> (L.) Eames & Boivin	THTH2	NI
<i>Toxicodendron radicans</i> (L.) Kuntze	TORA2	FAC
<i>Trifolium</i> spp. L.	TRIFO	
<i>Trillium</i> spp. L.	TRILL	
<i>Urtica dioica</i> L.	URDI	FACU
<i>Valerianella umbilicata</i> (Sullivant) Wood	VAUM	FAC
<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	VEAL	FAC
<i>Veronica arvensis</i> L.*	VEAR	NI
<i>Vernonia</i> spp. L.	VERNO	
<i>Vinca minor</i> L.*	VIMI2	NI
<i>Viola</i> spp. L.	VIOLA	
<i>Viola striata</i> Ait.	VIST3	FACW
<i>Vitis vulpina</i> L.	VIVU	FAC
<i>Woodsia obtusa</i> (Spreng.) Torr.	WOOB2	NI

Non-native species indicated with an (*)

Note: Plants were identified only to genus group if species could not be determined

Appendix Table 12: Herbaceous and vine species within 41 riparian sites in Jefferson and Oldham counties in Kentucky. Site names refer to the watershed in which the site was located, where B = Middle Fork Beargrass, G = Goose, and H = South Fork Harrods Creek watersheds. Importance values (IV) for each species were calculated by adding average absolute % cover and frequency for each site. Data within multi-plot sites were combined. Land use categories for each site were determined by the proportion of impervious surface (IS) within the subcatchment in which the site was located. Urban $\geq 30\%$ IS, Rural $\leq 10\%$ IS, and Suburban is between 10 and 30% IS.

Site	Land use categ.	Species name	Avg. % cover	Freq	IV
G2	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.96	17	18
G2	Rural	<i>Duchesnea indica</i> (Andr.) Focke*	0.58	6	6
G2	Rural	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	12.83	61	73
G2	Rural	<i>Galium aparine</i> L.	0.42	11	11
G2	Rural	<i>Glechoma hederacea</i> L.*	0.67	44	45
G2	Rural	<i>Impatiens capensis</i> Meerb.	0.04	6	6
G2	Rural	<i>Oxalis stricta</i> L.	0.50	6	6
G2	Rural	<i>Plantago rugelii</i> Dcne.	0.83	17	18
G2	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.58	17	18
G2	Rural	<i>Taraxacum officinale</i> G.H. Weber ex Wiggers*	0.17	6	6
G2	Rural	<i>Toxicodendron radicans</i> (L.) Kuntze	0.67	17	18
G2	Rural	<i>Veronica arvensis</i> L.*	1.50	22	23
G2	Rural	<i>Vinca minor</i> L.*	12.96	44	56
G2	Rural	<i>Viola</i> spp. L.	0.25	28	28
G4	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	32.89	100	133
G4	Rural	<i>Duchesnea indica</i> (Andr.) Focke*	0.78	22	23
G4	Rural	<i>Juncus</i> spp. L.	2.78	56	58
G4	Rural	<i>Lonicera japonica</i> Thunb.*	1.33	22	24
G4	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	0.11	11	11
G4	Rural	<i>Parthenocissus quinquefolia</i> (L.) Planch.	3.94	78	82
G4	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	1.61	56	57
G4	Rural	<i>Solidago</i> spp. L.	1.28	44	46
G4	Rural	<i>Symphotrichum cordifolium</i> (L.) Nesom	0.22	11	11
G4	Rural	<i>Toxicodendron radicans</i> (L.) Kuntze	0.06	11	11

Appendix Table 12 (cont.)

G4	Rural	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	4.44	11	16
G7	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	12.75	100	113
G7	Rural	<i>Asarum canadense</i> L.	0.08	17	17
G7	Rural	<i>Glechoma hederacea</i> L.*	2.75	100	103
G7	Rural	Grass	0.83	17	18
G7	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	1.83	33	35
G7	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.33	17	17
G7	Rural	<i>Symphotrichum</i> spp. L.	3.33	67	70
G7	Rural	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	0.50	17	17
H1	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.83	25	26
H1	Rural	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	4.33	33	38
H1	Rural	<i>Allium vineale</i> L.*	0.17	33	34
H1	Rural	<i>Ambrosia artemisiifolia</i> L.	0.04	8	8
H1	Rural	<i>Arisaema triphyllum</i> L.	0.04	8	8
H1	Rural	<i>Apiaceae</i>	0.04	8	8
H1	Rural	<i>Desmodium</i> spp. L.	0.13	17	17
H1	Rural	<i>Duchesnea indica</i> (Andr.) Focke*	1.29	67	68
H1	Rural	<i>Elymus riparius</i> Wieg.	0.46	33	34
H1	Rural	<i>Erigeron philadelphicus</i> L.	0.04	8	8
H1	Rural	<i>Galium aparine</i> L.	1.71	50	52
H1	Rural	<i>Glechoma hederacea</i> L.*	8.42	100	108
H1	Rural	<i>Impatiens capensis</i> Meerb.	0.17	17	17
H1	Rural	<i>Lonicera japonica</i> Thunb.*	0.58	67	67
H1	Rural	<i>Lamiaceae</i>	0.04	8	8
H1	Rural	<i>Ornithogalum umbellatum</i> L.*	0.04	8	8
H1	Rural	<i>Osmorhiza claytonii</i> (Michx.) C.B. Clarke	1.38	50	51
H1	Rural	<i>Packera</i> spp. L.	0.04	8	8
H1	Rural	<i>Parthenocissus quinquefolia</i> (L.) Planch.	0.42	25	25
H1	Rural	<i>Pilea pumila</i> (L.) Gray	0.33	42	42
H1	Rural	<i>Podophyllum peltatum</i> L.	1.08	8	9
H1	Rural	<i>Sanicula</i> spp. L.	1.50	58	60
H1	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	7.00	100	107
H1	Rural	<i>Smilax rotundifolia</i> L.	0.42	25	25
H1	Rural	<i>Solidago</i> spp. L.	0.58	25	26
H1	Rural	<i>Stachys</i> spp. L.	0.08	8	8
H1	Rural	<i>Thalictrum thalictroides</i> (L.) Eames & Boivin	0.13	17	17

Appendix Table 12 (cont.)

H1	Rural	<i>Toxicodendron radicans</i> (L.) Kuntze	0.33	17	17
H1	Rural	<i>Trillium</i> spp. L.	0.13	17	17
H1	Rural	<i>Urtica dioica</i> L.	0.17	8	9
H1	Rural	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	0.17	8	9
H1	Rural	<i>Viola</i> spp. L.	0.38	42	42
H1	Rural	<i>Viola striata</i> Ait.	0.75	42	42
H1	Rural	<i>Vitis vulpina</i> L.	0.08	8	8
H11	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.21	8	9
H11	Rural	<i>Glechoma hederacea</i> L.*	8.00	67	75
H11	Rural	<i>Lonicera japonica</i> Thunb.*	0.67	42	42
H11	Rural	<i>Lamiaceae</i>	0.29	17	17
H11	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	10.00	33	43
H11	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.75	17	17
H11	Rural	<i>Solidago</i> spp. L.	0.50	25	26
H11	Rural	<i>Symphyotrichum lowrieianum</i> (Porter) Nesom	0.25	8	9
H11	Rural	<i>Toxicodendron radicans</i> (L.) Kuntze	0.58	17	17
H11	Rural	<i>Viola striata</i> Ait.	0.04	8	8
H12	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	4.43	63	67
H12	Rural	<i>Ambrosia trifida</i> L.	0.19	8	9
H12	Rural	<i>Cirsium arvense</i> (L.) Scop.*	0.04	4	4
H12	Rural	<i>Desmodium</i> spp. L.	0.06	4	4
H12	Rural	<i>Elephantopus carolinianus</i> Raeusch.	0.13	13	13
H12	Rural	<i>Glechoma hederacea</i> L.*	0.13	4	4
H12	Rural	<i>Lactuca biennis</i> (Moench) Fern.	0.42	21	21
H12	Rural	<i>Lonicera japonica</i> Thunb.*	2.32	42	44
H12	Rural	<i>Lysimachia nummularia</i> L.*	1.69	33	35
H12	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	8.63	29	38
H12	Rural	<i>Packera</i> spp. L.	0.08	8	8
H12	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	2.69	42	44
H12	Rural	<i>Securigera varia</i> (L.) Lassen*	2.42	21	23
H12	Rural	<i>Solidago canadensis</i> L.	0.92	17	17
H12	Rural	<i>Solidago</i> spp. L.	0.31	12	13
H12	Rural	<i>Symphyotrichum pilosum</i> (Willd.) Nesom	0.83	8	9
H12	Rural	<i>Toxicodendron radicans</i> (L.) Kuntze	0.29	12	13
H12	Rural	<i>Trifolium</i> spp. L.	0.21	4	4
H12	Rural	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	1.56	42	43
H12	Rural	<i>Viola</i> spp. L.	0.13	8	8

Appendix Table 12 (cont.)

H13	Rural	<i>Securigera varia</i> (L.) Lassen*	4.75	67	71
H13	Rural	<i>Vernonia</i> spp. L.	0.08	17	17
H14	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.21	8	9
H14	Rural	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	0.33	21	21
H14	Rural	<i>Asarum canadense</i> L.	0.15	13	13
H14	Rural	<i>Desmodium canescens</i> (L.) DC	9.50	54	64
H14	Rural	<i>Elymus</i> spp. L.	0.08	4	4
H14	Rural	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.08	8	8
H14	Rural	<i>Glechoma hederacea</i> L.*	20.94	87	108
H14	Rural	Grass	2.46	17	19
H14	Rural	<i>Hydrophyllum canadense</i> L.	5.20	88	93
H14	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	0.04	4	4
H14	Rural	<i>Phytolacca americana</i> L.	0.29	4	4
H14	Rural	<i>Polygonum</i> spp. L.	1.54	13	14
H14	Rural	<i>Polygonum virginianum</i> L.	0.46	4	5
H14	Rural	<i>Rudbeckia laciniata</i> L.	0.54	8	9
H14	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.23	21	21
H14	Rural	<i>Symphotrichum</i> spp. L.	0.92	29	30
H14	Rural	<i>Urtica dioica</i> L.	20.28	58	78
H14	Rural	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	1.44	21	22
H14	Rural	<i>Viola striata</i> Ait.	0.46	21	21
H14	Rural	<i>Woodsia obtusa</i> (Spreng.) Torr.	0.02	4	4
H3	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.83	17	18
H3	Rural	<i>Boehmeria cylindrica</i> (L.) Sw.	0.92	8	9
H3	Rural	<i>Apiaceae</i>	0.17	25	25
H3	Rural	<i>Duchesnea indica</i> (Andr.) Focke*	0.96	25	26
H3	Rural	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.17	25	25
H3	Rural	<i>Glechoma hederacea</i> L.*	1.00	33	34
H3	Rural	Grass	13.75	33	47
H3	Rural	<i>Lysimachia nummularia</i> L.*	0.04	8	8
H3	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	7.50	42	49
H3	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.25	17	17
H3	Rural	<i>Spiraea</i> spp. L.	0.33	8	9
H3	Rural	<i>Symphotrichum</i> spp. L.	0.96	33	34
H3	Rural	<i>Symphotrichum racemosum</i> (Ell.) Nesom	0.17	8	9
H3	Rural	<i>Viola</i> spp. L.	1.21	42	43

Appendix Table 12 (cont.)

H5	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	2.10	71	73
H5	Rural	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	0.06	13	13
H5	Rural	<i>Allium vineale</i> L.*	0.27	54	55
H5	Rural	<i>Arisaema triphyllum</i> L.	0.08	4	4
H5	Rural	<i>Asarum canadense</i> L.	0.10	13	13
H5	Rural	<i>Carex blanda</i> Dewey	0.27	25	25
H5	Rural	<i>Cystopteris tenuis</i> (Michx.) Desv.	0.02	4	4
H5	Rural	<i>Desmodium</i> spp. L.	0.38	33	34
H5	Rural	<i>Dichanthelium clandestinum</i> (L.) Gould	0.17	8	8
H5	Rural	<i>Elephantopus carolinianus</i> Raeusch.	0.29	25	25
H5	Rural	<i>Erigeron philadelphicus</i> L.	0.08	13	13
H5	Rural	<i>Galium aparine</i> L.	0.15	17	17
H5	Rural	<i>Glechoma hederacea</i> L.*	5.71	79	85
H5	Rural	Grass	1.77	54	56
H5	Rural	<i>Impatiens capensis</i> Meerb.	0.04	8	8
H5	Rural	<i>Iodanthus pinnatifidus</i> (Michx.) Steud.	0.06	8	8
H5	Rural	<i>Lonicera japonica</i> Thunb.*	1.10	29	30
H5	Rural	<i>Lysimachia nummularia</i> L.*	0.06	8	8
H5	Rural	<i>Lamiaceae</i>	0.08	4	4
H5	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	5.52	50	56
H5	Rural	<i>Packera</i> spp. L.	0.02	4	4
H5	Rural	<i>Pilea pumila</i> (L.) Gray	0.23	37	37
H5	Rural	<i>Plantago rugelii</i> Dcne.	0.02	4	4
H5	Rural	<i>Polygonum</i> spp. L.	0.42	4	5
H5	Rural	<i>Sanicula canadensis</i> L.	2.38	71	73
H5	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.56	50	51
H5	Rural	<i>Solidago</i> spp. L.	0.31	29	29
H5	Rural	<i>Symphyotrichum cordifolium</i> (L.) Nesom	0.06	13	13
H5	Rural	<i>Thalictrum thalictroides</i> (L.) Eames & Boivin	0.04	8	8
H5	Rural	<i>Toxicodendron radicans</i> (L.) Kuntze	0.02	4	4
H5	Rural	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	1.00	37	38
H5	Rural	<i>Viola</i> spp. L.	0.79	59	59
H5	Rural	<i>Viola striata</i> Ait.	0.33	38	38
H6	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	1.11	89	90
H6	Rural	<i>Asarum canadense</i> L.	0.72	67	67
H6	Rural	Grass	0.28	44	45
H6	Rural	<i>Lonicera japonica</i> Thunb.*	0.39	67	67

Appendix Table 12 (cont.)

H6	Rural	<i>Lysimachia nummularia</i> L.*	0.28	44	45
H6	Rural	<i>Packera obovata</i> (Muhl. ex Willd.) W.A. Weber & A. Löve	0.11	22	22
H6	Rural	<i>Ruellia caroliniensis</i> (J.F. Gmel.) Steud.	0.11	11	11
H6	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.17	33	34
H6	Rural	<i>Symphotrichum cordifolium</i> (L.) Nesom	0.22	11	11
H6	Rural	<i>Viola</i> spp. L.	0.61	67	67
H6	Rural	<i>Viola striata</i> Ait.	0.11	17	17
H7	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.94	50	51
H7	Rural	<i>Asarum canadense</i> L.	4.63	100	105
H7	Rural	<i>Asteraceae</i>	0.69	75	76
H7	Rural	<i>Aureolaria</i> spp. L.	0.25	38	38
H7	Rural	<i>Lysimachia nummularia</i> L.*	0.31	50	50
H7	Rural	<i>Ranunculus hispidus</i> Michx.	0.19	13	13
H7	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.19	38	38
H7	Rural	<i>Symphotrichum lowrieianum</i> (Porter) Nesom	0.13	13	13
H7	Rural	<i>Viola</i> spp. L.	2.81	100	103
H8	Rural	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.25	25	25
H8	Rural	<i>Allium vineale</i> L.*	0.31	63	63
H8	Rural	<i>Arisaema triphyllum</i> L.	0.63	4	5
H8	Rural	<i>Asarum canadense</i> L.	0.21	17	17
H8	Rural	<i>Aureolaria</i> spp. L.	0.06	8	8
H8	Rural	<i>Carex blanda</i> Dewey	0.02	4	4
H8	Rural	<i>Cystopteris tenuis</i> (Michx.) Desv.	0.06	17	17
H8	Rural	<i>Desmodium</i> spp. L.	0.21	29	29
H8	Rural	<i>Duchesnea indica</i> (Andr.) Focke*	0.02	4	4
H8	Rural	<i>Elymus riparius</i> Wieg.	0.04	4	4
H8	Rural	<i>Erigeron philadelphicus</i> L.	0.02	4	4
H8	Rural	<i>Galium aparine</i> L.	0.13	25	25
H8	Rural	<i>Geum vernum</i> (Raf.) Torr. & Gray	0.08	13	13
H8	Rural	Grass	0.81	29	30
H8	Rural	<i>Impatiens capensis</i> Meerb.	0.04	8	8
H8	Rural	<i>Iodanthus pinnatifidus</i> (Michx.) Steud.	0.02	4	4
H8	Rural	<i>Juncus</i> spp. L.	0.21	21	21
H8	Rural	<i>Lonicera japonica</i> Thunb.*	0.02	4	4
H8	Rural	<i>Lysimachia nummularia</i> L.*	2.60	59	61
H8	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	1.50	13	14

Appendix Table 12 (cont.)

H8	Rural	<i>Packera</i> spp. L.	0.21	29	30
H8	Rural	<i>Packera obovata</i> (Muhl. ex Willd.) W.A. Weber & A. Löve	0.02	4	4
H8	Rural	<i>Pilea pumila</i> (L.) Gray	0.70	62	63
H8	Rural	<i>Plantago rugelii</i> Dcne.	0.04	8	8
H8	Rural	<i>Ranunculus hispidus</i> Michx.	0.04	8	8
H8	Rural	<i>Sanicula canadensis</i> L.	2.83	58	61
H8	Rural	<i>Sanicula</i> spp. L.	0.17	25	25
H8	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	2.42	71	73
H8	Rural	<i>Sedum ternatum</i> Michx.	0.15	8	8
H8	Rural	<i>Smilax rotundifolia</i> L.	0.02	4	4
H8	Rural	<i>Solidago</i> spp. L.	0.50	38	38
H8	Rural	<i>Symphyotrichum cordifolium</i> (L.) Nesom	0.10	17	17
H8	Rural	<i>Thalictrum thalictroides</i> (L.) Eames & Boivin	0.06	8	8
H8	Rural	<i>Valerianella umbilicata</i> (Sullivant) Wood	0.06	8	8
H8	Rural	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	1.08	42	43
H8	Rural	<i>Viola</i> spp. L.	1.48	83	85
H8	Rural	<i>Viola striata</i> Ait.	0.48	54	55
H9	Rural	<i>Campsis radicans</i> (L.) Seem. ex Bureau	0.83	17	18
H9	Rural	<i>Lonicera japonica</i> Thunb.*	0.67	33	34
H9	Rural	<i>Lysimachia nummularia</i> L.*	0.33	17	17
H9	Rural	<i>Microstegium vimineum</i> (Trin.) A. Camus *	0.08	17	17
H9	Rural	<i>Packera</i> spp. L.	0.25	33	34
H9	Rural	<i>Plantago rugelii</i> Dcne.	0.17	17	17
H9	Rural	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	5.33	83	89
H9	Rural	<i>Solidago</i> spp. L.	0.42	33	34
H9	Rural	<i>Toxicodendron radicans</i> (L.) Kuntze	0.33	33	34
H9	Rural	<i>Viola</i> spp. L.	0.50	17	17
H9	Rural	<i>Vitis vulpina</i> L.	0.33	33	34
B10	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.08	8	8
B10	Suburban	<i>Ambrosia trifida</i> L.	6.92	17	24
B10	Suburban	<i>Bidens</i> spp. L.	0.58	8	9
B10	Suburban	<i>Commelina communis</i> L.*	0.17	8	9
B10	Suburban	<i>Duchesnea indica</i> (Andr.) Focke*	0.08	8	8
B10	Suburban	<i>Elephantopus carolinianus</i> Raeusch.	0.67	17	17
B10	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	4.17	8	13
B10	Suburban	<i>Glechoma hederacea</i> L.*	0.42	8	9

Appendix Table 12 (cont.)

B10	Suburban	<i>Helianthus</i> spp. L.	2.50	8	11
B10	Suburban	<i>Lonicera japonica</i> Thunb.*	0.08	8	8
B10	Suburban	<i>Lysimachia nummularia</i> L.*	0.04	8	8
B10	Suburban	<i>Oxalis stricta</i> L.	0.17	8	9
B10	Suburban	<i>Plantago rugelii</i> Dcne.	0.17	8	9
B10	Suburban	<i>Solidago</i> spp. L.	1.33	17	18
B10	Suburban	<i>Taraxacum officinale</i> G.H. Weber ex Wiggers*	0.29	17	17
B10	Suburban	<i>Toxicodendron radicans</i> (L.) Kuntze	1.92	17	19
B10	Suburban	<i>Viola</i> spp. L.	2.33	17	19
B11	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	4.00	67	71
B11	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	6.67	33	40
B11	Suburban	<i>Parthenocissus quinquefolia</i> (L.) Planch.	2.83	67	70
B11	Suburban	<i>Phytolacca americana</i> L.	3.00	33	36
B11	Suburban	<i>Rosaceae</i>	1.67	33	35
B11	Suburban	<i>Toxicodendron radicans</i> (L.) Kuntze	1.67	33	35
B11	Suburban	<i>Viola</i> spp. L.	0.17	8	9
B12	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	22.25	100	122
B12	Suburban	<i>Asarum canadense</i> L.	1.50	17	18
B12	Suburban	<i>Cystopteris tenuis</i> (Michx.) Desv.	0.08	8	8
B12	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	1.63	58	60
B12	Suburban	<i>Glechoma hederacea</i> L.*	11.08	92	103
B12	Suburban	<i>Hedera helix</i> L.*	0.17	8	9
B12	Suburban	<i>Impatiens capensis</i> Meerb.	1.21	33	35
B12	Suburban	<i>Lonicera japonica</i> Thunb.*	1.17	50	51
B12	Suburban	<i>Microstegium vimineum</i> (Trin.) A. Camus *	38.42	75	113
B12	Suburban	<i>Parthenocissus quinquefolia</i> (L.) Planch.	0.25	8	9
B12	Suburban	<i>Phytolacca americana</i> L.	0.13	8	8
B12	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.46	25	25
B12	Suburban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.17	8	9
B12	Suburban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	1.17	17	18
B12	Suburban	<i>Viola</i> spp. L.	0.08	8	8
B12	Suburban	<i>Vitis vulpina</i> L.	1.54	67	68
G1	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	3.13	42	45
G1	Suburban	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	10.75	33	44
G1	Suburban	<i>Glechoma hederacea</i> L.*	2.75	17	19
G1	Suburban	<i>Microstegium vimineum</i> (Trin.) A. Camus *	2.92	17	20

Appendix Table 12 (cont.)

G1	Suburban	<i>Stellaria media</i> (L.) Vill.*	0.08	8	8
G10	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	2.42	42	44
G10	Suburban	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	0.67	8	9
G10	Suburban	<i>Desmodium</i> spp. L.	1.00	50	51
G10	Suburban	<i>Elymus riparius</i> Wieg.	3.83	83	87
G10	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.92	8	9
G10	Suburban	<i>Galium aparine</i> L.	0.04	8	8
G10	Suburban	<i>Glechoma hederacea</i> L.*	18.25	100	118
G10	Suburban	<i>Impatiens capensis</i> Meerb.	0.17	25	25
G10	Suburban	<i>Iodanthus pinnatifidus</i> (Michx.) Steud.	0.17	8	9
G10	Suburban	<i>Parthenocissus quinquefolia</i> (L.) Planch.	0.25	17	17
G10	Suburban	<i>Rudbeckia laciniata</i> L.	2.58	33	36
G10	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.83	25	26
G10	Suburban	<i>Smilax rotundifolia</i> L.	1.25	8	10
G10	Suburban	<i>Solidago</i> spp. L.	8.75	92	100
G10	Suburban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	10.08	67	77
G10	Suburban	<i>Viola</i> spp. L.	0.08	8	8
G11	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	2.33	42	44
G11	Suburban	<i>Allium vineale</i> L.*	0.04	8	8
G11	Suburban	<i>Desmodium</i> spp. L.	0.58	67	67
G11	Suburban	<i>Duchesnea indica</i> (Andr.) Focke*	0.71	33	34
G11	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	28.42	67	95
G11	Suburban	<i>Glechoma hederacea</i> L.*	9.42	50	59
G11	Suburban	Grass	10.83	25	36
G11	Suburban	<i>Impatiens capensis</i> Meerb.	1.00	8	9
G11	Suburban	<i>Lonicera japonica</i> Thunb.*	1.83	50	52
G11	Suburban	<i>Lysimachia nummularia</i> L.*	0.42	25	25
G11	Suburban	<i>Microstegium vimineum</i> (Trin.) A. Camus *	7.83	17	25
G11	Suburban	<i>Plantago rugelii</i> Dcne.	0.08	8	8
G11	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.54	25	26
G11	Suburban	<i>Smilax rotundifolia</i> L.	0.04	8	8
G11	Suburban	<i>Sorghum halepense</i> (L.) Pers.*	0.50	8	9
G11	Suburban	<i>Solidago</i> spp. L.	0.67	42	42
G11	Suburban	<i>Toxicodendron radicans</i> (L.) Kuntze	1.96	25	27
G11	Suburban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	2.50	58	61
G12	Suburban	<i>Boehmeria cylindrica</i> (L.) Sw.	0.92	25	26
G12	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	19.17	50	69

Appendix Table 12 (cont.)

G12	Suburban	Grass	0.17	8	9
G12	Suburban	<i>Impatiens capensis</i> Meerb.	0.21	17	17
G12	Suburban	<i>Lonicera japonica</i> Thunb.*	0.96	58	59
G12	Suburban	<i>Maianthemum racemosum</i> (L.) Link	0.58	17	17
G12	Suburban	<i>Parthenocissus quinquefolia</i> (L.) Planch.	13.38	92	105
G12	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.33	25	25
G12	Suburban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.63	17	17
G12	Suburban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	5.33	75	80
G12	Suburban	<i>Viola</i> spp. L.	0.08	8	8
G12	Suburban	<i>Woodsia obtusa</i> (Spreng.) Torr.	0.08	8	8
G13	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	3.63	83	87
G13	Suburban	<i>Arundinaria gigantea</i> (Walt.) Muhl.	17.67	92	109
G13	Suburban	<i>Desmodium</i> spp. L.	0.29	25	25
G13	Suburban	<i>Duchesnea indica</i> (Andr.) Focke*	1.17	42	43
G13	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.13	17	17
G13	Suburban	<i>Glechoma hederacea</i> L.*	1.13	100	101
G13	Suburban	Grass	0.33	17	17
G13	Suburban	<i>Impatiens capensis</i> Meerb.	0.04	8	8
G13	Suburban	<i>Microstegium vimineum</i> (Trin.) A. Camus *	1.67	33	35
G13	Suburban	<i>Pilea pumila</i> (L.) Gray	0.63	42	42
G13	Suburban	<i>Symphotrichum</i> spp. L.	0.13	17	17
G13	Suburban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	0.42	17	17
G13	Suburban	<i>Viola</i> spp. L.	0.21	42	42
G13	Suburban	<i>Viola striata</i> Ait.	0.21	17	17
G14	Suburban	<i>Erigeron philadelphicus</i> L.	3.67	67	70
G14	Suburban	<i>Glechoma hederacea</i> L.*	0.67	33	34
G14	Suburban	Grass	95.00	100	195
G14	Suburban	<i>Pilea pumila</i> (L.) Gray	2.33	33	36
G14	Suburban	<i>Plantago rugelii</i> Dcne.	0.50	33	34
G14	Suburban	<i>Ruellia caroliniensis</i> (J.F. Gmel.) Steud.	2.00	67	69
G14	Suburban	<i>Rumex verticillatus</i> L.	0.33	33	34
G14	Suburban	<i>Symphotrichum lowrieanum</i> (Porter) Nesom	0.67	33	34
G14	Suburban	<i>Viola</i> spp. L.	0.33	33	34
G3	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.54	42	42
G3	Suburban	<i>Aristolochia tomentosa</i> Sims	0.17	8	9
G3	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	2.50	42	44
G3	Suburban	<i>Impatiens capensis</i> Meerb.	0.04	8	8

Appendix Table 12 (cont.)

G3	Suburban	<i>Juncus</i> spp. L.	0.21	25	25
G3	Suburban	<i>Lysimachia nummularia</i> L.*	1.75	25	27
G3	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.17	17	17
G3	Suburban	<i>Solidago</i> spp. L.	2.46	67	69
G3	Suburban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.67	25	26
G3	Suburban	<i>Viola</i> spp. L.	11.54	67	78
G6	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	2.38	42	44
G6	Suburban	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	0.04	8	8
G6	Suburban	<i>Arundinaria gigantea</i> (Walt.) Muhl.	8.75	100	109
G6	Suburban	<i>Asarum canadense</i> L.	0.04	8	8
G6	Suburban	<i>Cystopteris tenuis</i> (Michx.) Desv.	0.13	17	17
G6	Suburban	<i>Duchesnea indica</i> (Andr.) Focke*	0.50	17	17
G6	Suburban	<i>Glechoma hederacea</i> L.*	34.92	83	118
G6	Suburban	Grass	0.08	8	8
G6	Suburban	<i>Impatiens capensis</i> Meerb.	0.21	25	25
G6	Suburban	<i>Lamiaceae</i>	0.33	42	42
G6	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.21	25	25
G6	Suburban	<i>Solidago</i> spp. L.	0.13	8	8
G6	Suburban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	0.58	17	17
G6	Suburban	<i>Viola</i> spp. L.	0.04	8	8
G9	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	4.58	75	80
G9	Suburban	<i>Duchesnea indica</i> (Andr.) Focke*	0.38	25	25
G9	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.08	8	8
G9	Suburban	<i>Galium aparine</i> L.	0.04	8	8
G9	Suburban	<i>Glechoma hederacea</i> L.*	41.38	42	83
G9	Suburban	Grass	0.83	25	26
G9	Suburban	<i>Impatiens capensis</i> Meerb.	2.21	42	44
G9	Suburban	<i>Lysimachia nummularia</i> L.*	0.25	8	9
G9	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.83	75	76
G9	Suburban	<i>Smilax rotundifolia</i> L.	0.08	8	8
G9	Suburban	<i>Solidago</i> spp. L.	7.17	75	82
G9	Suburban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.04	8	8
G9	Suburban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	1.08	25	26
G9	Suburban	<i>Viola</i> spp. L.	2.54	75	78
G9	Suburban	<i>Viola striata</i> Ait.	2.38	58	61
H10	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	1.50	73	74

Appendix Table 12 (cont.)

H10	Suburban	<i>Lonicera japonica</i> Thunb.*	2.91	45	48
H10	Suburban	<i>Packera obovata</i> (Muhl. ex Willd.) W.A. Weber & A. Löve	0.05	9	9
H10	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	5.41	91	96
H10	Suburban	<i>Symphyotrichum cordifolium</i> (L.) Nesom	0.14	18	18
H2	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	2.00	67	69
H2	Suburban	<i>Asarum canadense</i> L.	1.42	67	68
H2	Suburban	<i>Apiaceae</i>	0.04	8	8
H2	Suburban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.58	58	59
H2	Suburban	<i>Glechoma hederacea</i> L.*	1.63	83	85
H2	Suburban	Grass	0.92	58	59
H2	Suburban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.58	67	67
H2	Suburban	<i>Smilax rotundifolia</i> L.	0.17	8	9
H2	Suburban	<i>Symphyotrichum</i> spp. L.	0.42	25	25
H2	Suburban	<i>Thalictrum thalictroides</i> (L.) Eames & Boivin	0.04	8	8
H2	Suburban	<i>Viola</i> spp. L.	0.92	58	59
H4	Suburban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	0.25	33	34
H4	Suburban	<i>Ampelamus albidus</i> (Nutt.) Britt.	0.08	17	17
H4	Suburban	<i>Asarum canadense</i> L.	1.75	100	102
H4	Suburban	<i>Lactuca canadensis</i> L.	0.08	17	17
H4	Suburban	<i>Microstegium vimineum</i> (Trin.) A. Camus *	0.67	17	17
H4	Suburban	<i>Viola</i> spp. L.	0.08	17	17
H4	Suburban	<i>Viola striata</i> Ait.	0.08	17	17
B1	Urban	<i>Ageratina</i> spp. L.	0.13	2	2
B1	Urban	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	7.13	75	82
B1	Urban	<i>Asarum canadense</i> L.	0.02	4	4
B1	Urban	<i>Duchesnea indica</i> (Andr.) Focke*	0.83	25	26
B1	Urban	<i>Erigeron</i> spp. L.	0.04	4	4
B1	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.17	8	9
B1	Urban	<i>Glechoma hederacea</i> L.*	18.10	79	97
B1	Urban	<i>Impatiens capensis</i> Meerb.	3.00	63	66
B1	Urban	<i>Lamium amplexicaule</i> L.	0.21	4	4
B1	Urban	<i>Lonicera japonica</i> Thunb.*	0.04	4	4
B1	Urban	<i>Sanicula</i> spp. L.	0.31	8	9
B1	Urban	<i>Sicyos angulatus</i> L.	0.42	25	25
B1	Urban	<i>Solidago</i> spp. L.	1.37	42	43

Appendix Table 12 (cont.)

B1	Urban	<i>Urtica dioica</i> L.	1.88	29	31
B1	Urban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	1.81	21	23
B1	Urban	<i>Viola</i> spp. L.	0.06	8	8
B13	Urban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	1.83	33	35
B13	Urban	<i>Helianthus</i> spp. L.	1.17	33	35
B13	Urban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	0.25	17	17
B13	Urban	<i>Vitis vulpina</i> L.	0.08	17	17
B14	Urban	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	0.17	17	17
B14	Urban	<i>Asarum canadense</i> L.	1.25	17	18
B14	Urban	<i>Cystopteris tenuis</i> (Michx.) Desv.	0.08	8	8
B14	Urban	<i>Duchesnea indica</i> (Andr.) Focke*	0.08	8	8
B14	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	53.08	100	153
B14	Urban	<i>Smilax rotundifolia</i> L.	0.04	8	8
B14	Urban	<i>Urtica dioica</i> L.	1.92	42	44
B2	Urban	<i>Ageratina</i> spp. L.	1.46	50	51
B2	Urban	<i>Asarum canadense</i> L.	1.83	25	27
B2	Urban	<i>Apiaceae</i>	0.17	8	9
B2	Urban	<i>Duchesnea indica</i> (Andr.) Focke*	0.04	8	8
B2	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	27.00	83	110
B2	Urban	<i>Glechoma hederacea</i> L.*	2.29	50	52
B2	Urban	Grass	11.50	67	78
B2	Urban	<i>Microstegium vimineum</i> (Trin.) A. Camus *	0.17	8	9
B2	Urban	<i>Packera obovata</i> (Muhl. ex Willd.) W.A. Weber & A. Löve	0.04	8	8
B2	Urban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.92	42	43
B2	Urban	<i>Smilax rotundifolia</i> L.	1.08	8	9
B2	Urban	<i>Toxicodendron radicans</i> (L.) Kuntze	2.21	50	52
B2	Urban	<i>Urtica dioica</i> L.	1.92	8	10
B2	Urban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	0.54	25	26
B2	Urban	<i>Viola</i> spp. L.	0.63	33	34
B2	Urban	<i>Vitis vulpina</i> L.	0.17	17	17
B3	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	14.67	50	65
B3	Urban	<i>Parthenocissus quinquefolia</i> (L.) Planch.	0.04	8	8
B3	Urban	<i>Polygonatum biflorum</i> (Walt.) Ell.	0.75	17	17
B3	Urban	<i>Smilax rotundifolia</i> L.	0.04	8	8
B3	Urban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.04	8	8
B4	Urban	<i>Ageratina</i> spp. L.	0.13	8	8
B4	Urban	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	9.67	50	60

Appendix Table 12 (cont.)

B4	Urban	<i>Cardamine pensylvanica</i> Muhl. ex Willd.	0.04	8	8
B4	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	68.08	100	168
B4	Urban	<i>Impatiens capensis</i> Meerb.	0.42	25	25
B5	Urban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	7.00	42	49
B5	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	36.33	67	103
B5	Urban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.08	8	8
B5	Urban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	2.21	25	27
B6	Urban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	17.25	83	101
B6	Urban	<i>Ampelamus albidus</i> (Nutt.) Britt.	0.58	33	34
B6	Urban	<i>Duchesnea indica</i> (Andr.) Focke*	0.42	33	34
B6	Urban	<i>Solidago</i> spp. L.	3.92	83	87
B6	Urban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	0.50	17	17
B6	Urban	<i>Viola</i> spp. L.	1.00	33	34
B7	Urban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	10.57	63	73
B7	Urban	<i>Alliaria petiolata</i> (Bieb.) Cavara & Grande *	0.27	8	9
B7	Urban	<i>Aureolaria</i> spp. L.	0.02	4	4
B7	Urban	<i>Cystopteris tenuis</i> (Michx.) Desv.	0.04	8	8
B7	Urban	<i>Duchesnea indica</i> (Andr.) Focke*	0.33	17	17
B7	Urban	<i>Elymus riparius</i> Wieg.	0.17	4	4
B7	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.71	13	13
B7	Urban	<i>Glechoma hederacea</i> L.*	1.90	25	27
B7	Urban	Grass	6.92	54	61
B7	Urban	<i>Lonicera japonica</i> Thunb.*	0.29	13	13
B7	Urban	<i>Lysimachia nummularia</i> L.*	0.13	4	4
B7	Urban	<i>Oxalis stricta</i> L.	0.02	4	4
B7	Urban	<i>Plantago rugelii</i> Dcne.	0.11	13	13
B7	Urban	<i>Ranunculus hispidus</i> Michx.	0.02	4	4
B7	Urban	<i>Sanicula</i> spp. L.	0.25	4	4
B7	Urban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	1.24	38	39
B7	Urban	<i>Smilax rotundifolia</i> L.	0.15	8	8
B7	Urban	<i>Solidago</i> spp. L.	1.31	25	26
B7	Urban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.12	8	8
B7	Urban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	13.32	84	97
B7	Urban	<i>Viola</i> spp. L.	0.32	17	17
B8	Urban	<i>Aristolochia tomentosa</i> Sims	0.13	8	8
B8	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.42	25	25
B8	Urban	<i>Juncus</i> spp. L.	0.21	8	9

Appendix Table 12 (cont.)

B8	Urban	<i>Oxalis stricta</i> L.	0.17	17	17
B8	Urban	<i>Phytolacca americana</i> L.	0.17	8	9
B8	Urban	<i>Solidago</i> spp. L.	2.21	17	19
B8	Urban	<i>Viola</i> spp. L.	0.08	8	8
B9	Urban	<i>Sorghum halepense</i> (L.) Pers.*	0.08	8	8
B9	Urban	<i>Toxicodendron radicans</i> (L.) Kuntze	0.13	8	8
G8	Urban	<i>Ageratina altissima</i> (L.) King & H.E. Robins.	3.13	50	53
G8	Urban	<i>Carex blanda</i> Dewey	12.56	25	38
G8	Urban	<i>Euonymus fortunei</i> (Turcz.) Hand.-Maz. *	0.06	13	13
G8	Urban	<i>Glechoma hederacea</i> L.*	3.19	63	66
G8	Urban	<i>Sanicula odorata</i> (Raf.) K.M. Pryer & L.R. Phillippe	0.13	25	25
G8	Urban	<i>Verbesina alternifolia</i> (L.) Britt. ex Kearney	3.00	25	28
G8	Urban	<i>Viola</i> spp. L.	0.06	13	13

Non-native species indicated with an (*).

Note: Plants were identified to the most distinct taxonomic group possible.

Land use categories based upon % impervious surface at the subcatchment scale.

Appendix Table 13: Soil properties and trace nutrients from soil samples removed from static chamber centers located in riparian sites in urban, suburban, and rural subcatchments in October 2008. Urban subcatchments contain $\geq 30\%$ impervious surface cover, rural subcatchments contain $\leq 10\%$ impervious surface cover, and suburban subcatchments contain values in between 10 and 30% impervious surface cover. Site names beginning with R, S, and U are located in rural, suburban, and urban subcatchments, respectively. The second letter in the site name indicates whether the site is located in Harrods (H), Goose (G), or Beargrass (B) creek watershed.

Site	Sand (%)	Silt (%)	Clay (%)	TEC (meq/100 g)	pH (1:1 soil to water)	Organic Matter (%)	S (mg/kg)	Bulk dens.
RG4	19.9	65.1	15.0	12.9	7.1	4.48	21	0.59
RG7	17.7	67.2	15.1	13.8	7.1	4.06	36	0.53
RH13	27.3	54.8	18.0	21.8	6.6	9.98	44	0.73
RH5	43.2	43.2	13.6	21.1	7.2	3.4	29	0.52
SB12	7.8	80.8	11.5	22.3	7.6	5.06	38	0.56
SG13	21.1	64.1	14.9	14.4	7.3	4.95	36	0.58
SG9	10.9	72.5	16.6	13.3	7.5	3.5	23	0.56
UB1	11.3	67.9	20.8	17.1	7.2	4.74	36	0.63
UB14	22.6	67.5	10.0	28.4	6.9	6.49	81	0.49
UB2	28.9	54.9	16.2	20.0	7.7	3.98	26	0.66
UB3	15.8	66.3	17.9	15.5	7.0	4.78	37	0.5
UB5	8.3	70.6	21.1	13.1	7.4	6.36	30	0.55
UG8	28.3	59.1	12.6	11.6	6.1	4.15	31	0.58

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Appendix Table 13 (cont.)

Site	Bray II P (mg/kg)	Ca (mg/ kg)	Mg (mg/ kg)	K (mg/ kg)	Na (mg/ kg)	Ca (%)	Mg (%)	K (%)	Na (%)	H (%)
RG4	113	1992	251	81	22	77.2	16.2	1.61	0.74	0
RG7	75	2143	269	51	26	77.7	16.3	0.95	0.82	0
RH13	158	3410	232	128	22	78.4	8.9	1.51	0.44	6
RH5	114	3694	168	54	40	87.7	6.6	0.66	0.83	0
SB12	55	3843	238	66	23	86.1	8.9	0.76	0.45	0
SG13	63	2002	411	94	22	69.7	23.9	1.68	0.67	0
SG9	121	2033	271	75	32	76.6	17.0	1.45	1.05	0
UB1	151	2559	404	58	26	74.7	19.6	0.87	0.66	0
UB14	122	4787	283	102	27	84.4	8.3	0.92	0.41	1.5
UB2	11	3395	237	69	22	85.1	9.9	0.89	0.48	0
UB3	86	2568	208	61	22	82.8	11.2	1.01	0.62	0
UB5	53	1901	340	79	18	72.3	21.6	1.54	0.60	0
UG8	122	1428	213	157	22	61.7	15.3	3.48	0.83	14

Appendix Table 13 (cont.)

Site	B (mg/kg)	Fe (mg/kg)	Mn (mg/kg)	Cu (mg/kg)	Zn (mg/kg)	Al (mg/kg)	P (mg/kg)
RG4	0.97	186	317	5.16	8.74	461	26
RG7	0.85	226	472	4.45	11.42	403	45
RH13	1.09	324	284	4.78	11.47	404	97
RH5	0.60	208	334	6.83	15.22	371	61
SB12	0.98	218	411	3.59	5.99	290	41
SG13	0.93	152	409	3.97	11.26	439	24
SG9	0.77	187	419	6.01	12.87	478	41
UB1	0.86	289	464	7.67	16.59	467	92
UB14	0.99	398	476	5.15	9.69	339	97
UB2	0.70	173	356	2.04	3.27	254	13
UB3	0.64	195	361	4.89	12.98	363	34
UB5	0.96	153	525	2.02	4.57	359	22
UG8	0.64	262	311	5.52	19.55	608	102

Methods for the data collected are referenced as follows: Total exchange capacity (Ross 1995); pH (McLean 1982); organic matter (Schulte and Hopkins 1996); S, Ca, Mg, K, Na, B, Fe, Mn, Cu, Zn, Al, and P (Mehlich 1984); soil texture (ASTM Standard D422-63 2002); Bray-II Phosphorus (Bray and Kurtz 1945); Zn, Mn, Fe, and Cu (Linsay and Norvell 1978).

Appendix Table 14: Monthly mean carbon dioxide flux rates (mg CO₂-C/m²*d) from riparian soils within urban, suburban, and rural subcatchments from January to October 2008.

Site	Jan	Feb	March	April	May	June	July	Aug	Sept	Oct
RG4	0	454	422	1817	2377	3666	3056	2555	1927	1103
RG7	994	276	1233	2940	1460	1512	1544	1950	1146	773
RH13	174	-47	245	2581	2377	2084	4816	1711	1699	1559
RH5	-255	-359	225	1485	1127	1029	1836	1056	1174	735
SB12	-	741	190	1794	2262	3087	2838	1260	2493	1260
SG13	511	715	104	1536	1007	1387	1953	2140	2388	1366
SG9	513	488	793	2189	1895	1666	3250	1593	1782	1160
UB1	181	-78	241	2171	1253	1642	2806	687	988	874
UB14	-	-	-	-	-	-	5166	3527	4336	2840
UB2	73	301	690	2800	3376	3970	3761	2591	2959	2775
UB3	254	180	586	1750	1635	2524	2596	2920	2355	3532
UB5	257	466	616	2116	1997	3778	3148	2612	3025	1561
UG8	488	323	446	2567	2015	2665	4743	2605	2326	1523

Appendix Table 15: Monthly mean methane flux rates (mg CH₄-C/m²*d) from riparian soils within urban, suburban, and rural subcatchments from January to October 2008.

Site	Jan	Feb	March	April	May	June	July	Aug	Sept	Oct
RG4	-0.10	-0.26	-0.15	-0.41	-0.26	-0.24	-0.29	-0.47	-0.55	-0.59
RG7	0.25	1.16	1.92	6.61	0.72	0.64	1.11	0.06	0.13	-0.29
RH13	-0.13	-	-0.11	0.01	0.09	0.39	-0.09	-0.31	-0.19	-0.16
RH5	-0.48	-0.28	-0.30	-0.62	-0.48	0.04	-0.33	-0.98	-1.24	-0.94
SB12	0.03	-0.19	0.08	-0.33	-0.44	-0.30	-0.82	-1.37	-1.30	-1.02
SG13	-0.15	-0.01	-0.04	-0.10	-0.29	-0.12	0.13	0.05	-0.25	-0.29
SG9	-0.03	-0.22	0.30	-0.25	-0.50	-0.43	-1.69	-0.79	-1.03	-1.14
UB1	-0.11	0.12	-0.16	1.91	-0.17	-0.24	-0.46	-1.06	-0.70	-1.00
UB14	-	-	-	-	-	-	-0.49	-0.69	-0.62	-0.66
UB2	0.64	-0.18	-0.24	-0.35	-0.51	-0.54	-0.66	-0.65	-0.91	-0.73
UB3	-0.35	-0.21	-0.23	-0.51	0.01	-0.11	-0.32	-0.88	-0.82	-0.79
UB5	-0.72	-0.34	-0.23	-0.51	-0.46	-0.75	-0.62	-1.06	-0.88	-0.88
UG8	-0.41	0.04	0.01	-0.28	-0.65	-0.16	0.07	-0.54	-0.97	-0.62

Appendix Table 16: Monthly mean nitrous oxide flux rates (mg N₂O-N/m²*d) from riparian soils within urban, suburban, and rural subcatchments in August and October 2008.

Site	Aug	Oct
RG4	0.15	0.20
RG7	0.08	0.04
RH13	0.34	0.05
RH5	0.09	0.09
SB12	0.05	0.13
SG13	0.30	0.13
SG9	0.17	0.10
UB1	0.11	0.14
UB14	0.23	0.23
UB2	0.16	0.22
UB3	0.14	-0.02
UB5	0.17	0.14
UG8	0.17	0.13

CURRICULUM VITAE

Education:

- 2003-present PhD. Candidate, Biology, University of Louisville, Louisville, KY
Dissertation Title: Riparian corridor vegetation structure and soil function along urban, suburban, and rural streams in Louisville, KY, USA.
Advisor: Dr. Margaret Carreiro
GPA: 3.67
- 1999-2001 Master of Science, Biology, Western Carolina University, Cullowhee, NC
Thesis: Role of the alternatively spliced *Arabidopsis thaliana* AtB'g5' untranslated region on translation during heat shock and development.
Advisor: Dr. Sabine Rundle
GPA: 3.84
- 1998-1999 Post-baccalaureate, Biology, University of North Carolina
GPA: 3.76
- 1991-1996 Bachelor of Science, Psychology, Western Carolina University
GPA: 3.06

Research Experience:

- 2004-2008 Dissertation research, University of Louisville
Studied nitrate removal and methane uptake functions from riparian zones as a means to protect our urban waterways. Determined herbaceous and woody plant community structure along streambanks in urban environments.
- 2007 Research Assistant, University of Louisville
Conducted field and lab work studying leaf decomposition in a channelized stream and riparian area.
- 2001-2002 Research Assistant I, UT Southwestern Medical Center
Used analytical techniques such as amphotropic transfections of mammalian T cells, radioactive isotope analysis, DNA amplification, cloning, and analysis of plasmid DNA.
- 2000-2001 Research Assistant, Western Carolina University
Studied translation control mechanisms within *Arabidopsis thaliana* using techniques for RNA amplification and RNA and protein quantification.

Teaching Experience:

- 2003-2009 University of Louisville, Department of Biology, Teaching Assistant
Human Anatomy and Physiology Lab, 8 semesters
General Biology Lab, 1 semester
Biology for Non-majors lab, 2 semesters
General Ecology lab, 1 semester
- 2004 University of Louisville, Department of Minority Affairs, Adjunct
Instructor. Biology for Pre-med students, Summer term
- 2003 Southwestern Community College, Biology Department, Adjunct
Instructor. Human Anatomy and Physiology, Summer term
- 2001 Western Carolina University, Department of Biology, Teaching Assistant
Human Anatomy and Physiology Lab, 1 semester
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Grants:

- 2008 Kentucky Society of Natural History Research Grant (\$400)
University of Louisville Student Government Association Travel Grant
(\$200)
- 2005 Woody Boebinger Memorial Scholarship, Kentucky Society of Natural
History (\$800)
Center for Environmental Science Matching Funds, University of
Louisville (\$500)

Publications and Posters:

- 2010 Voices of the next generation: Perspectives from the participants in the
2010 conservation policy Park Break session. Heath Garris, Jessica
Goodrich, Annamarie Leon Guerrero, Matthew J. Heard, Meghan
Lindsey, Archi Rastogi, Rodney J. White.
<http://www.georgewright.org/perspectives>
- 2008 Characterization of riparian plant communities along an urban-to-rural
gradient in Louisville, KY. White, R. Jonathan and Carreiro, Margaret M.
Ecological Society of America Annual Meeting.
- 2007 Variation in near-stream groundwater depth in urban and rural
subcatchments, White, R. J. and Carreiro, M. M. Kentucky Academy of
Science Annual Meeting.
- 2002 Functional analysis of the B' subunit of *Arabidopsis* protein phosphatase
2A. White, R. J., Davis, M., Esmon, C. A., Myrick, T. L., Vallee, N.,
Cochran, D. S., Stewart, C. N. Jr., and Rundle, S. J. *Plant Science*, 162:
201-209.

Professional Affiliations and Meetings:

- George Wright Society
- 2010-2011, Student member
 - 2010, Park Break Fellow

Ecological Society of America

- 2006-2008, webmaster, Student Section
- 2005-2008, Student member

Kentucky Native Plant Society

- 2007-2009, interpretive native plant hike

Kentucky Society of Natural History

- 2008, Student member

Golden Key International Honour Society, University of Louisville

- 2006 inductee

Pi Gamma Mu International Honour Society, University of North Carolina

- 1996 inductee

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Journal of Environmental Quality

Urban Ecosystems

Statistical/Analytical Packages

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Graduate Coursework (grade in parentheses):

Conservation Biology (B+)

Evolution (B)

Biostatistics (A)

Ecosystem Ecology (A-)

Special Topics: Ecology of Urban and Suburban Landscapes (A-)

Aquatic Ecology (A-)

Population and Community Ecology (A)

Geographic Information Systems (A)