A non-invasive diagnostic system for early assessment of acute renal transplant rejection.

Mohamed Nazih Mohamed Ibrahim Shehata

University of Louisville

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A NON-INVASIVE DIAGNOSTIC SYSTEM FOR EARLY ASSESSMENT OF ACUTE RENAL TRANSPLANT REJECTION

By

Mohamed Nazih Mohamed Ibrahim Shehata
B.Sc., CECS, Mansoura University, Mansoura, Egypt, 2009

A Thesis Submitted to the Faculty of
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for the Degree of

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Department of Electrical and Computer Engineering
University of Louisville
Louisville, Kentucky

August 2016
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A Thesis Approved on

July 15, 2016

by the Following Thesis Committee:

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Tamer Inanc, Ph.D.

Hermann Frieboes, Ph.D.
DEDICATION

This thesis is dedicated to my beloved parents who brought me up in obeying the orders of Allah. They said and they are still saying, “Oh Mohamed, do not despair of Allah’s mercy. Go, work hard and pay more effort and Allah will not let you down.” I wish they were here sharing with me these important moments of happiness and success in my life.
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Finally, I would like to send a brief message to my very close friends who supported and encouraged me during my M.Sc. period, “I value your friendship and I missed you very much.”
ABSTRACT

A NON-INVASIVE DIAGNOSTIC SYSTEM FOR EARLY ASSESSMENT OF ACUTE RENAL TRANSPLANT REJECTION

Mohamed Nazih Mohamed Ibrahim Shehata

July 15, 2016

Early diagnosis of acute renal transplant rejection (ARTR) is of immense importance for appropriate therapeutic treatment administration. Although the current diagnostic technique is based on renal biopsy, it is not preferred due to its invasiveness, recovery time (1-2 weeks), and potential for complications, e.g., bleeding and/or infection.

In this thesis, a computer-aided diagnostic (CAD) system for early detection of ARTR from 4D (3D + \(b\)-value) diffusion-weighted (DW) MRI data is developed. The CAD process starts from a 3D B-spline-based data alignment (to handle local deviations due to breathing and heart beat) and kidney tissue segmentation with an evolving geometric (level-set-based) deformable model. The latter is guided by a voxel-wise stochastic speed function, which follows from a joint kidney-background Markov-Gibbs random field model accounting for an adaptive kidney shape prior and for on-going visual kidney-background appearances. A cumulative empirical distribution of apparent diffusion coefficient (ADC) at different \(b\)-values of the segmented DW-MRI is considered a discriminatory transplant status feature. Finally, a classifier based on deep learning of a non-negative constrained stacked auto-encoder is employed to distinguish between rejected and non-rejected renal transplants. In the “leave-one-subject-out” experiments on 53 subjects, 98% of the subjects were correctly classified (namely, 36 out of 37 rejected transplants and 16 out of 16 non-rejected ones). Additionally, a four-fold cross-validation experiment was performed, and
an average accuracy of 96% was obtained. These experimental results hold promise of the proposed CAD system as a reliable non-invasive diagnostic tool.
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CHAPTER I
EXISTING TECHNIQUES FOR THE ASSESSMENT OF RENAL REJECTION: A SURVEY

The kidney is a very important complicated filtering organ of the body. Many complications and diseases can arise in this organ. One such disease is chronic kidney disease (CKD), which is a gradual loss of function of the nephrons. When the kidney reaches stage 5 chronic kidney disease, end stage renal failure, the preeminent therapy is renal transplantation. Although it is the best form of treatment, the dearth of kidney donors is still challenging. Therefore, all efforts should be employed to prolong the survival rate of the transplanted kidney. However, graft dysfunction (e.g., acute rejection) is one of the serious barriers to long term kidney transplant survival. Currently, graft dysfunction’s gold standard of diagnosis is renal biopsy. Although renal biopsy is helpful, it is not preferred due to its invasive nature, high morbidity rates, and expensiveness. Therefore, noninvasive imaging techniques have become the subject of extensive research and interest, giving strong promise to replace, or at least to decrease, biopsy usage in diagnosing graft dysfunction. This chapter will discuss not only the kidney anatomy, chronic kidney disease, treatment, and current diagnosis but also the state-of-the-art imaging techniques in diagnosing graft dysfunction.
A. Introduction

The kidney is a very important organ. It is the main filtration organ in the human body, keeping the nutrients that the body needs in and expelling the waste that can become toxic. Maintaining the health of this organ is critical. There are diseases that can cause the kidney to decrease in function such as diabetes, hypertension, glomerular disease, and polycystic kidney disease [3]. These can cause a gradual loss of function in the kidney leading to waste build up in the body and the patient to develop chronic kidney disease (CKD). CKD affects about 26 million people with 17,000 transplants being performed each year in the U.S. [4, 5]. Though this has greatly improved the outcome of patients diagnosed with stage 5 CKD, complications can still arise. One of the main concerns is graft dysfunction. Routine post-transplantation clinical evaluation of kidney function is of immense importance to prevent the graft loss. The diagnostic technique currently recommended by the National Kidney Foundation (NKF) to measure overall kidney function is Glomerular Filtration Rate (GFR), which is based on measuring the serum creatinine level. However, this test has low sensitivity and is a late marker for renal dysfunction (a significant change in serum creatinine level is detectable only after the loss of 60% of renal function), and it does not assess the function of individual kidneys. The current gold standard for diagnosing different types of kidney dysfunction is needle biopsy [6]. However, this can be difficult, costly, and time-consuming. Renal biopsy can also result in complications such as infections, bleeding, and at times, death. With the evolution of computer-aided diagnostic (CAD) systems, we hope to non-invasively diagnose different types of graft dysfunction, saving time and money. This chapter will give an overview on how CKD is treated and kept viable in post-transplantation when it is affected by graft dysfunction. Thus, the need for new noninvasive techniques which have the capability to provide accurate diagnosis of kidney dysfunction is of great clinical importance.

This chapter presents an overview of current clinical techniques for renal transplant function evaluation as well as an examination of new ways to improve the detection of graft dysfunction using image-based technology [7]. The rest of this chapter is organized
as follows: Section I.B provides a brief overview of the anatomy and function of the kid-
ney. Section I.C takes a closer look at who is more at risk for developing CKD and some
symptoms associated with it. In addition, an overview of the treatment options for people
that develop stage 5 CKD will be given, concentrating on transplantation as a definitive
therapy. Section I.D takes a look at follow-up post transplantation care, which includes
possible complications that could arise with a concentration on graft dysfunction. Sec-
tion I.E will concentrate on tests that are performed to detect graft dysfunctions including
the traditional methods such as blood, urine, and renal biopsy. This is followed by the
image based techniques such as ultrasound, and magnetic resonance imaging (MRI).

B. Kidney Anatomy and Function

As stated before, kidneys are the main filtration system in the body. Kidneys are
able to keep nutrients like salts, sugar, and protein in, while at the same time expelling
excess nutrients, water, and waste such as urea and ammonia out of the body. Kidneys
keep human bodies in a homeostatic state. They regulate the blood’s pH, blood pressure,
and osmolality. Osmolality is the amount of particles of solutes that are dissociated in a
solvent [8]. Each kidney is shaped like a bean and is about the size of a fist [9] and weighs
about 150 g [10]. They are located in the lower back below the rib cage.

As shown in Figure 1, the kidney is composed of an outer ”shell”, which is the renal
cortex; an inner layer, the renal medulla; and a hollow area where the urine is collected,
the renal pelvis [10] . Inside the cortex and medulla are the filtration units known as the
nephrons (see Figure 2), which are then made up of smaller subunits such as the glomerulus,
vasa recta, and loop of Henle [10]. Since the kidneys must filtrate the blood, they must be
connected to veins and arteries. The kidneys are connected to the renal artier and vein
which are connected to the iliac artery and vein, respectively. That is the general overview
of the anatomy of the kidneys, now this chapter will trace the filtration pathway of the
blood. The blood enters the kidney by way of the renal artier. Once there, the blood moves
to the nephrons of the cortex where the blood then enters the afferent arteriole which allows
the blood to enter in the glomerulus. The glomerulus is then able to filter out waste through the assistance of blood pressure. This waste is filtered into the Bowman’s capsule. From the Bowman’s capsule the waste moves to the proximal tubule then to the Loop of Henle and thin segment, which can be found in the medulla. At these places in the nephron, more filtration can be done. From here, waste moves to the distal tubule then the collecting tubule and finally into the renal pelvis. The waste that ends up in the renal pelvises will then move through the ureter to the bladder and then out of the body by way of the urethra. The clean blood exits the glomerulus by way of the efferent arteriole. Once there, more filtration can be done in the Peritubular capillaries where the blood could also move down the Vasa recta in the medulla; nutrients that were filtered out by the loop of Henle and thin segment can be resorbed there. The blood then exits through the venules and then through the arcuate veins, and finally leaves the kidney through the renal vein. The clean blood can then travel
back to the heart [10]. As one can see, this organ is very complex and with this complexity many problems can arise. In the next section this chapter will discuss what can go wrong and who is at risk of developing these complications.

C. Renal Problems/Symptoms and Treatment

The renal system is a very complex system, in which various complications and diseases can arise pertaining to it, especially with the kidney. There are multiple conditions and/or diseases that can arise in the kidney such as kidney stones, injury, infections, and cancer. The focus of this chapter will be on CKD, the 9th leading cause of death in America [11]. CKD is a gradual loss of function of the kidney, where the nephrons become compromised [4]. To date, 26 million people in the U.S. are living with CKD [4]. The risk of developing CKD could be increased due to a few different factors such as various physiological conditions, diseases, age, race, lifestyle, and family history. Conditions that
increase the likelihood of developing CKD include diabetes, high blood pressure, heart disease, and high cholesterol [12]. People with the following diseases are at increased risk of CKD; HIV, Hep C, Metabolic syndrome, cancer, and sickle cell trait [13]. Individual 65 years or older are also more at risk. African Americans tend to also have a greater risk compared to the rest of the population [12]. Lifestyles that increase risk include obesity [14] or smoking [12]. The last risk factor is family history; if a patient has a parent or family member with CKD they have a greater chance of developing the disease [12]. People with these risk factors should be aware and contact their doctor if any of the following symptoms become present. In early stage CKD the patient may be asymptomatic but as the disease progresses, kidney function will worsen and symptoms will develop. Symptoms can include change in urination, whether it changes in frequency, feeling, color, or texture. The urine can also start to contain blood [15]. Apart from the change in urination, symptoms can include limb swelling, iron build up that can cause nose bleeds and bad breath [15]. As the disease progresses infection-like symptoms can arise [16]. CKD can have an effect not only on the body but also on mental state and activity level. These symptoms can include fatigue, generalized weakness, decreased libido, change in memory [16], and a decline in mental function [15]. More rare but more serious symptoms may include rash, generalized pain, chest pain, and shortness of breath [15–18]. If a patient should have any of these last symptoms, he/she should seek immediate medical attention. Every patient is different and there is no set relationship between symptoms and the stage of the patient’s kidney disease [16]. As stated before, if one should develop any of these symptoms, he/she should talk to their doctor about checking for CKD. If left untreated, symptoms could worsen and kidney failure will advance to stage 5 CKD. The untreated patient with stage 5 CKD could die due to the build up toxins in the body. To prevent this from happening, the patient should be treated with dialysis or transplantation.

Luckily, there have been developments in treatments for patients with stage 5 renal failure. Those treatments include blood dialysis or renal transplant. Blood dialysis is when one’s blood is filtered of waste or excess water, either with use of a machine outside the body (hemodialysis) or chemically inside the body (Peritoneal Dialysis) [19]. Though
dialysis is a helpful treatment, a more long term treatment would be kidney transplantation. This is where a donor’s kidney is surgically inserted into the CKD patient. That new kidney should improve filtration for the patient. Since transplantation is the definitive therapy for End-Stage Renal Disease (ESRD), the following describes in more detail the kidney transplantation procedure and associated complications and diseases.

As previously stated, renal transplantation is a surgical procedure where a donated kidney is placed inside the patient with CKD. However, it does not mean that a nephrectomy (i.e. removal of the malfunctioned kidney) is performed on the patient with CKD. The patient with CKD usually gets to keep both of the kidneys, unless those kidneys are causing pain or other complications [20]. This means that the patient will have three kidneys after the procedure. The donated kidney also has its own ureter, renal artery, and vein intact. The donated kidney is placed below (distal of) the native kidneys with the donated ureter connecting to the bladder, and the renal artery and vein connecting to the iliac artery and vein of the patient, respectively [21]. Figure 3 demonstrates the entire anatomy of a patient renal system after transplantation.

This procedure seems fairly simple in concept. However, the process to find that donor can be fairly complicated not only medically and logistically but also due to legal hurdles. There are two different types of donors that can be used for CKD; cadaver and living donors. Only one third of the transplantations are from living donors while two thirds of the transplantations are from cadaver donors [22]. Often, the physician and the CKD patient must decide whether to use a living or a cadaver kidney. Often a more desirable choice would be to have a living donor give one of their kidneys. However, this is not without its complications. The donor must meet all criteria such as being HLA (+or−) and/or ABO compatible, physically in good health, and be in no way coerced against their will to donate [22, 23]. This means that the donors can back out at any time. This is why even if there is a willing living donor the physician may persuade the patient to get on the United Network of Organ Sharing (UNOS) [24]. Depending on where the patient is on the list determines if and when they will receive a donor. The donated kidneys from this list are from cadaver donors. In order for the cadaver’s kidney to be viable the kidney must
be functioning before death and the manner of death must not damage the organ. Also the time and location of death may play a part since the kidney must not decompose before the kidney can be donated [22]. In the United States, the living donors must be willing to give up their kidney, cadaver donors must also make it clear that they are willing to be a donor after death. With all of these criteria, it is no wonder that although there are about 17,000 kidney transplants performed annually in the United States, there are still about 100,800 people waiting for a kidney. With this in mind, it is very important that the transplanted kidney is kept viable as long as possible so that a nephrectomy and repeat transplantation do not have to be performed [5]. In the following section, an overview on what happens during post-transplantation care should be given, which in turn should improve the viability of the organ post-transplantation. This includes follow up procedures and the complications that arise for transplant patients.
D. Post-Transplant Follow-Ups and Complications

Just as with any medical procedure complications can arise such as infection and bleeding, so one must remain in constant care of one’s physician. They must continue follow ups to insure that the new kidney is functioning and that new complications do not arise. To prevent new complications from arising, the patient should follow the instructions of their physician by taking their anti-rejection medications and visiting their physician as directed [25]. The frequency at which a patient has to visit their physician will decrease as time after transplantation increases. Once the patient has reached 210 days post-operatively, they should be seeing their physician monthly or if abnormalities arise [26]. During those clinical visits various tests such as examining the patient’s weight, blood pressure, and temperature will be done to assess both the overall health of the patient and the health of the kidney. The urine and the blood tests will be discussed later in this chapter. If these tests appear abnormal, the physician may order a renal biopsy and/or scans [27]; both will be discussed in greater detail later on in the chapter. For now, this chapter will concentrate on the complications that can arise during these tests, specifically those complications that are associated with (renal) transplants. This Section will take a look at those types and concentrate specifically on graft dysfunction. First, this chapter will explain types of complications other than graft dysfunction.

1. Types of Complications

There are six categories of complications including: urological complications, vascular complications, fluid collection, neoplasms, recurrent native renal disease, and graft dysfunction. This section will take a short look at the first five complications then the next section will go a little more in depth for graft dysfunctions. The term urological complications generally refers to uncontrolled or obstructed urine flow [28, 29]. Vascular complications are complications that are associated with the vascular system of the renal system, i.e. renal or iliac artery/vein. Vascular complications can include narrowing, blockage or formations of holes in the vascular system [30]. Fluid collection is closely related
to urological complications and/or vascular complications in that if there is a urological or vascular complication, fluid such as blood or urine will collect in areas where they are not supposed to be collecting. This will create urinomas hematomas, abscesses, or lymphoceles [29–31]. Neoplasms are abnormal growth such as tumors that grow on the renal system and other areas. This is said to be caused by the prolonged exposure to immune repressor drugs [30, 32, 33]. Lastly, recurrent native renal disease is when the disease that caused the patient to develop CKD in the first place, such as diabetes, is now affecting the donated kidney [34, 35]. It is possible for the patient to develop a combination of these complications. It is important that for any of these complications, diagnosis and treatment are done as soon as possible. Most of these complications are usually easier to detect as compared to graft dysfunction. This is because these complications can be detected using various imaging techniques such as ultrasounds and MRIs [36]. These imaging techniques will be discussed later on in this chapter but the concentration will be more on graft dysfunction. The complication and cause that is more challenging to diagnose is graft dysfunction, which shall be examined next.

2. Graft Dysfunction

Graft dysfunction simply means that the newly transplanted organ is no longer functioning, toxins then build up, and the body rejects the transplanted organ [30]. It was calculated that within the first five years post transplantation, 15% of patients will experience graft dysfunction [37]. There are three classes of graft dysfunction: hyperacute, acute, and chronic. The type of graft dysfunction is differentiated by the mechanism and somewhat by the time of dysfunction onset [38]. Hyperacute rejection is relatively rare nowadays. This class of rejection is caused by antibodies attacking the donated organ due to the donor organ having the wrong HLA (+or−) and/or ABO blood antigen and will present itself within in minutes or hours after transplantation [23]. There is no cure for hyperacute rejection [39]. Chronic kidney rejection’s mechanism is not well understood but appears to present itself after five years post-transplant [23]. The main concentration in this chapter
will be on Acute Kidney Rejection (AKR). Just as there are different types of complications in renal transplant, there are different causes of graft dysfunction. This can provide somewhat of a challenge in diagnosis and treatment. This is due to the fact that there is a different treatment for each cause of graft dysfunction. There are four different causes of graft dysfunction; they include: acute tubular necrosis (ATN) antibody-mediated rejection, T-cell mediated rejection, immunosuppressive toxicity (ITox), and viral infection (VI). ATN is when the antibodies of the patient recognize the newly donated kidney as a foreign body causing the tissue to become necrotic and die. It is treated with a drug therapy regimen that may include plasmapheresis, mycophenolate mofetil, and tacrolimus [40–42]. T-cell is when killer T-cells attack the donated organ causing apoptosis in the tissue [38]. The treatment for T-cell mediated rejection includes drugs such as corticosteroids, antithymocyte globulin, and immunosuppression therapy. If the patient has antibody-mediated rejection, they may not respond to the T-cell mediated rejection treatment [43–45]. Immunosuppressive toxicity is when the immunosuppressive drugs that are supposed to be preventing the immune system from rejecting the donated kidney actually cause renal failure since these drugs can be nephrotoxic. Treatment for this would be to cease or change the medication, cease, or to reduce the dose of the nephrotoxic drug such as Cyclosporine and tacrolimus [46–48]. VI is when viruses such as Cytomegalovirus or Herpes simplex virus enters the body and damages the kidney [49]. Treatments for VI may include administering immunosuppressant and/or antiviral medications [40]. The causes of AKR can be presented singularly or in combination, which can add to the difficulty in diagnosing the cause of AKR. How the cause of these graft dysfunctions are diagnosed will be discussed in the next section of this chapter.

E. Detection/Assessment of Renal Rejection

It is important that the patient keeps regular visits with their physician in order to ensure that their newly transplanted organ is in working order. The post-transplantation follow-ups’ main concern is to keep the graft viable for as long as possible. If the patient
continues regular follow-ups and notifies the physician of any symptoms that arise, it is possible that they can catch the problem early and save the donated organ. This chapter will give an overview of the existing techniques/methods for diagnosing graft dysfunction. This includes both traditional, non-imaging, clinical methods, and the imaged-based techniques that are in use but are still being developed and/or improved for use. More details about both methods are given below.

1. Traditional Methods

Traditionally, during a routine follow-up a blood and urine analysis will be implemented. If there are any abnormal results in either of these, the physician may order a renal biopsy to get a definitive diagnosis. This diagnosis should also tell the physician what is causing the kidney malfunction. This next section will show how these diagnoses are determined. First, this chapter will discuss urine testing.

a. Urine Tests  This method is very simple in use. It can test for multiple substances and is non-invasive. Using the patients urine, the physician is able to measure a number of biomarkers to determine GFR. Most often the biomarker used to calculate GFR is serum creatinine. To calculate GFR, the concentration of creatinine found in the urine sample is placed into an equation which has constants that change based on sex, race, and age. Using the calculated GFR, one is able to determine what stage of function the kidney(s) are in, 0 being at an increased risk and 5 being end stage renal failure [50, 51]. This diagnostic technique is currently recommended by the NKF to measure overall kidney function. However, this test has low sensitivity and is a late marker of renal dysfunction (a significant change in serum creatinine level is detectable only after the loss of 60% of renal function), and it does not assess the function of individual kidneys [6]. The next test that shall be discussed will be a blood test.

b. Blood Test/Works  This method is similar to urine in that it measures estimated GFR using serum creatinine. However, since it does pierce the skin when obtaining the blood, this test is slightly more invasive. A complete blood count and differential count
CBC and differential [52] measure more substances than urine including detecting the presence of burred blood cells which can be present in patients with CKD. Burr cells are blood cells that appear almost gear like. They appear when there is an excess amount of waste in the body, which is likely to happen in patients with CKD [53, 54]. Though it has slightly more benefits than a urine test, it has the similar setback in that the test has low sensitivity and is a late marker of renal dysfunction and it does not assess the function of individual kidneys [6]. The last traditional method that shall be discussed is a renal biopsy.

c. Biopsy (Gold Standard)  Renal biopsy is a traditional method for the graft function assessment that is by far the most invasive, but to date is considered the gold standard. This procedure is performed using a renal biopsy needle that is inserted into the patient’s back and kidney while being guided by a camera, ultrasound, or x-ray, as shown in Figure 4 [52]. The tissue that is obtained is read using a microscope [55]. The patient is fully conscious and asked to hold their breath and to not move [52]; if one should breathe or move, they run the risk of piercing other organs. Along with the risk of piercing other organs the patient also runs the risk of excessive bleeding and infections. Excessive bleeding can present itself more so in a patient who is on blood thinners. Infections are likely to occur since the patient is more than likely on an immunosuppressive therapy regimen [37, 56]. These complications can lead to nephrectomy or even death; both occur in 1 in every 1,000 renal biopsies [37].

Along with the invasiveness of the procedure, there are multiple setbacks that are associated with this procedure. Renal biopsies, although a useful tool, have the tendency to give a missed diagnosis or inaccurate estimate of the extent of the problem. This is because it is only sampling a small portion of the kidney and if off target in the slightest can miss an effective portion of the kidney and give a false negative. This would mean that a repeat biopsy may have to be performed causing the patient more pain and precious time lost in order to save the graft. On the subject of time, the time it takes to obtain the results can take up to two weeks [57]. That time which could be used for treatment is wasted and can result in failure of the donated kidney. On top of these setbacks, the financial cost of the procedure can reach over ($20,000) [58]. So this test cannot only cause physical pain
but also create more of a financial burden. Though these tests have been routinely used for transplant assessment and have helped in improving graft survival, one can see that there needs to be a better way to diagnose and differentiate the cause of graft dysfunction. Additionally, existing techniques, i.e. GFR and biopsy, for diagnosis of renal rejection are late biomarkers. Moreover, renal biopsy has significant morbidity, is very expensive, takes up to two weeks to get the final report, and can result in over- or under-estimation problems by only sampling small areas of the kidney. Therefore, the development of non-invasive tests to monitor kidney transplant rejection status is of immense importance. This, in turn, will allow doctors to intervene early to prevent rejection and the damage it causes, which will improve long-term outcomes. The following section will overview existing non-invasive imaging techniques and their possible use for assessing renal function and diagnosing graft dysfunction. In this chapter, the imaging techniques that will be discussed are ultrasound and MRI.
2. Image-Based Techniques for Renal Transplant Evaluation

The development of computer-aided diagnosis (CAD) systems for renal transplant assessment using different imaging modalities is an ongoing area of increased research. Non-invasive imaging-based techniques have been clinically used to assess transplanted kidneys with the advantage of providing information on each kidney separately. For example, radionuclide imaging (also called scintigraphy), the traditional method in renal imaging, is an excellent modality for evaluating graft function, both qualitatively and quantitatively, while also screening for common complications [59]. However, this technique fails to show accurate anatomical details due to its limited spatial resolution, so functional abnormalities inside different parts of the kidney (such as cortex and medulla) cannot be discriminated precisely [60]. Furthermore, radionuclide imaging includes radiation exposure [61], thus limiting the range of its applications, especially in monitoring such diseases as ATN or cyclosporin [62]. Computed tomography (CT) is a commonly available technology that uses contrast agents that allows accurate evaluation of various diseases in renal transplantation and with lower costs than magnetic resonance imaging (MRI) [63]. However, information gathered by CT to detect renal acute rejection is unspecific and the contrast agents used still are nephrotoxic. Therefore, currently CT has a limited role in diagnosing acute renal rejection [64]. In contrast to these radionuclides and CTs, ultrasound (US) and MRI are the most popular imaging modalities used for the diagnosis of kidney diseases. In the following sections, an overview of different CAD systems for the diagnosis of acute renal rejection using these two imaging modalities is given.

3. Ultrasound (US) Imaging

Ultrasound (US) imaging is usually used for the early assessment of renal allografts functionality in the postoperative period as well as for the assessment in the long-term follow-up thanks to being a relatively easy to be performed and repeated, inexpensive, and non-nephrotoxic imaging modality. [9]. Pulsatility index (PI) and resistance index (RI) are the most common measurements to assess renal functionality using US. Some recent stud-
ies that assessed renal transplants using different forms of ultrasound (e.g., power doppler (PD), color doppler (CD), contrast enhanced (CE), etc.), please see Figure 5) are discussed.

In an investigation to characterize the factors that influence PI and RI in patients with immediate (IGF), slow (SGF), or delayed (DGF) kidney graft function, Chudek et al. [65] observed that ischemic injury which occurred mainly prior to organ harvesting played a dominant role in determining intra-renal resistance in the early post-transplant period. A study by Saracino et al. [66] investigated whether the long-term renal functionality could be predicted using RI measurements taken early after kidney transplant. On the other hand, Kramann et al. [67] concentrated in their study on evaluating the potential of RI measurements to predict renal allograft survival. They concluded that, for prediction of long-term allograft survival, RI measurements should be taken 12-18 months post-transplantation. Krejčí et al. [68] utilized a composite gray-scale, CD imaging, and PD imaging to examine the power of US for early detection of a subclinical acute rejection. A significant difference between the four different groups in their study was obtained. In another study by Damasio et al. [69], the ability of doppler US to differentiate between dual and single kidney transplantation (DKT) and (SKT), respectively, was exploited. After the measurement of RI parameters for both DKT and SKT groups, they concluded that those patients with DKT had higher RI and lower kidney volumes than those with (SKT).

A study by Shebel et al. [70] investigated the ability of PD in the differentiation between acute rejection (AR) and ATN. Their study included 67 renal transplant recipients in the early post-transplantation period. After a manual placement of regions of interest
(ROIs), cortical perfusion (CP) and RI were measured for all recipients and CP was tested with respect to serum creatinine (SCr) and RI. Upon their own CP grading scale system, they found a statistical significant correlation between their CP grading and SCr \( (P < 0.01) \) and between CP grading and RI \( (P < 0.05) \). They concluded that the PD using CP grading is more sensitive in the detection of early AR compared to RI and cross-sectional measurements.

Fischer et al. [71] proved the superiority of ultrasound contrast media (USCM) to conventional US that uses the RI indicator in the diagnosis of early allograft dysfunction. In addition, Benozzi et al. [72] found that both US and CEUS could identify grafts with early dysfunction, but only some CEUS derived parameters could differentiate between ATN and AR. Schwenger et al. [73] exploited the power of CE sonography (CES) in early prediction of long-term renal transplant functionality compared to CD ultrasonography (CDUS). In their study, 68 renal transplants were investigated using both CES and CDUS one week after transplantation. Renal blood flow (RBF) and RI were measured for all transplant recipients and were correlated with the recipients’ clinical data represented by glomerular filtration rate (GFR) in a post-transplantation period from one week to one year. They concluded from their observations that RBF measurement using CES was significantly correlated with kidney functionality in the aforementioned period after transplantation, in contrast with RI measurement using CDUS. Another study was explored by Göcze et al. [74] to differentiate between acute kidney injury (AKI) stages using CEUS based on the quantification of blood perfusion. Instead of generating time-intensity curves (TIC), they used another quantification method called arrival time parametric imaging (ATPI). Their study included 10 patients who underwent CEUS, of which four patients had no evidence of AKI, one with stage AKI, and five with stage 2 or 3 AKI. Color-maps based on inflow time (IT) of the contrast agent were generated using the CEUS-ATPI quantification method and were divided into six major categories based on their values. Then, these ITs were assessed for different poles of kidney cortex (i.e. lower, middle, and upper) and the total IT was the sum of all arrival times of these three poles for each kidney. They observed that patients with stage 2 or 3 AKI have more delayed ITs than those of the other groups. They concluded
that CEUS-ATPI technique may help in detecting different stages of AKI. Recently, Jin et al. [75] assessed renal allografts using CEUS. In their study, 57 renal transplant patients underwent CEUS. Then, they were divided into three groups: 23 patients with AR (group 1), 10 patients with ATN (group 2), and 24 patients with normal allografts (group 3). After a manual placement of ROIs, a new index to detect AR called rising time (RisT) was measured instead of arrival time (AT). In addition, time to peak (TTP) and delta time among ROIs (ΔRisT and ΔTTP) were measured, analyzed, and correlated with clinical data (e.g., GFR). They found that RisT, TTP, and (ΔRisT and ΔTTP) were significantly higher in group 1 as compared to those in group 2 and group 3.

Although several studies utilized US to evaluate and assess renal functionality pre- and post-transplantation by evaluating conventional ultrasound parameters such as the PI and RI, two contradictory studies [76, 77] concluded that RI is not an exact indicator of renal graft dysfunction, and it could only provide a prognostic marker of the graft. Moreover, doppler US may give high PI and RI values (>0.8), which is an indication similar to those of ATN [78, 79]. These contradictions led researchers and investigators to examine a different imaging modality to assess renal functionality (e.g., MRI). In the next section, the state-of-the-art studies utilizing different MR imaging modalities are discussed.

4. Magnetic Resonance Imaging (MRI)

Magnetic resonance imaging (MRI) has become the most powerful and central non-invasive tool for clinical diagnosis of diseases [80]. The main advantage of MRI is that it provides excellent morphological information and offers the best soft tissue contrast among all imaging modalities (e.g., US and CT), which allows advanced analysis of different aspects of renal function. However, structural MRI lacks functional information. On the other hand, other MRI modalities, such as dynamic MRI, BOLD MRI and diffusion MRI are frequently used for renal function evaluation. Next, the state-of-the-art studies utilizing these MRI modalities for renal transplant assessment are overviewed.
Dynamic Contrast Enhanced (DCE) MRI: Dynamic MRI of the kidney has gained considerable attention in assessing renal function due to its ability to characterize tissue-specific functional changes, and potential to measure both total and cortical volume, and other functional parameters such as RBF, GFR, and renal plasma flow (RPF). Therefore, in recent years several studies have exploited DCE-MRI to non-invasively analyze kidney function in both native and transplanted kidneys. Figure 6 shows an example of DCE-MRI before, during, and after the administration of contrast to the kidney.

![DCE-MRI sequence with pre-, post-, and late-contrast](image)

**FIGURE 6:** Illustrative example of a DCE-MRI sequence with pre-, post-, and late-contrast.

In particular, a study by de Priester et al. [81] utilized dynamic MR enhancement curves to qualitatively evaluate diseased (27 patients) and nondiseased (8 patients) renal transplants. Cortical and the medullary enhancement parameters were obtained from a physiological model that was fitted to the raw data. Cortical arterial blood volume and medullary wash-out rates were found as the main discriminatory parameters between diseased and nondiseased patients. Yuksel [82] introduced a DCE-MRI-based CAD system for the evaluation of transplant function, which employed deformable image segmentation, kidney registration, and cortical perfusion construction. After kidney segmentation, a manual cortical ROI is used to construct the perfusion curve from the co-aligned images and the kidney function is evaluated visually based on the pattern of the constructed curves. Automated CAD system for early diagnosis of acute transplant rejection by Farag et al. [83] and El-Baz et al. [84–86] included parametric deformable model segmentation, nonrigid alignment, and classification of the kidney status using empirical parameters. Their frameworks were tested on 30 data sets and classified kidney status of each patient using four
indexes: the peak signal intensity, the time-to-peak, the wash-in slope (slope between the peak and the first minimum), and the wash-out slope (peak and the signal measured from the last image in the sequence), calculated from the MRI signal for the kidney cortex. A supervised Bayesian classifier was employed and the system classified 13 out 15 and 15 out of 15 correctly for both training and testing, respectively. Similar approaches were proposed in [87–89]. The study utilized a global alignment step of the MR images exploiting a special Gibbs energy function, and the perfusion curves were estimated from the whole kidney rather than only the cortex. The latter CADs were tested on a larger cohort of 100 patients and achieved a 94% diagnostic accuracy using Bayesian supervised classifier. A semiautomated approach by Rusinek et al. [90] assessed cortical and medullary functional parameters (RPF, GFR, vascular volumes of the cortex and medulla, and rate of water absorption) using compartmental modeling for both simulated and in-vivo data. Their framework employed an initial rigid alignment (translation only) step followed by a graph-cut based segmentation approach. The system was tested on 22 clinical data sets and the study concluded that the accuracy and precision in RPF and GFR are acceptable for clinical use.

Zikic et al. [91] evaluated kidney kinetic parameters after motion correction using template-matching based registration and normalized gradient field (NGF), as the contrast-invariant similarity measure. However, the kidney was segmented manually, and the evaluation of perfusion parameters (plasma volume and tubular flow) was performed visually by trained physicians for 10 data sets of healthy volunteers. Semi-automated evaluation of renal function for both native and transplanted kidneys was explored by De Senneville [92] using rigid-body registration to handle kidney motion inside a user-defined ROI. The renal cortex was segmented manually, and the GFR was estimated with Patlak-Rutland tracer kinetic model. The study demonstrated a significant uncertainty reduction on the computed GFR for native kidneys (10 healthy volunteers), but not the transplanted ones (10 transplant patients). Aslan et al. [93] developed an automated CAD system to classify normal kidney function from kidney rejection using DCE-MRI. Following kidney segmentation, three classification methods (least square support vector machines (LS-SVMs), Ma-
halanobis distance, and the Euclidean distance) were compared to assess transplant status based on medullary perfusion curves. On a cohort of 55 clinical data sets, they achieved a diagnostic accuracy, sensitivity, and specificity of 84%, 75%, and 96%, respectively using the Mahalanobis distance-based classifier. Anderlik et al. [94] proposed a framework for quantitative assessment of kidney function using a two-step motion correction and pharmacokinetic modeling. The GFR was estimated from the time-intensity curves using Sourbron et al. [95] compartment model. Their framework has been tested on 11 data sets. Zollner et al. [96] employed a non-rigid registration using B-splines and mutual information (MI) as a similarity metric. Functional information was extracted regionally using k-means clustering [97]. This system was tested on only 4 DCE-MRI data sets and the evaluation of kidney regions was assessed qualitatively according their mean signal intensity time courses. Wentland et al. [98] utilized MRI-based intrarenal perfusion measurement to differentiate between normal-functioning kidney allografts and allografts with ATN or AR on a cohort of 24 biopsy proven patients. The study concluded that the cortical and medullary blood flow is significantly reduced in grafts experiencing AR, as compared with normal grafts. Additionally, AR patients demonstrated medullary blood flow reduction as compared with ATN patients.

Recently, a study by Abou El-Ghar et al. [99] explored the feasibility of DCE-MRI in evaluation of renal allograft dysfunction. Their CAD system employed computer based techniques for motion correction and creation of renographic curves. Functional evaluation on 55 patients using the mean medullary intensity achieved sensitivity, specificity and accuracy of 75%, 96% and 84%, respectively, to separate of normal kidneys from impaired ones. Yamamoto et al. [100] utilized dynamic MRI to prospectively assess its ability to identify the cause of acute graft dysfunction. Their study employed 60 patients, 31 of which had normal function and 29 had acute dysfunction due to AR. Their study employed a multicompartamental tracer kinetic model to estimate the GFR and mean transit time (MTT) at different compartments of the kidney. The study documented differences in the fractional MTT values between normal grafts or grafts undergoing AR or ATN; however, substantial overlaps among these groups and with normal kidneys were observed. Semi-automated
estimation of renal parameters was performed by Hodneland et al. [101]. A viscous fluid model combined with an NGF-based cost function was used for elastic kidney registration. However, the kidney was segmented interactively with the nearest neighbor approach, the framework was tested only on 4 data sets of two healthy volunteers, and the reported GFR measurements were slightly underestimated relative to the creatinine reference values. Positano et al. [102] proposed a CAD system for the estimation for renal parameters, which included a two-step rigid registration, and adaptive prediction of kidney position over the course of the respiratory cycle. The perfusion indices (peak signal intensity, MTT, initial up-slope, and time to peak) were evaluated on perfusion curves extracted from the automatically and manually registered data sets and were similar as well. An automated framework for the classification of kidney transplant status was proposed by Khalifa et al. [103, 104]. In their framework, the kidney was segmented using a stochastic geometrical deformable model approach and the local motion of the kidney was corrected for by a Laplace partial differential equation-based nonrigid alignment method [105, 106]. Their initial study [103] included only 26 data sets, and a K-nearest neighbor classifier was used. Their system achieved a 92.31% correct classification using the time-to-peak and wash-out slope empirical parameters that are estimated from the agent kidney kinetic curves. Their framework was extended in [104] by using four augmented empirical parameters (peak intensity value, time-to-peak, up-slope and average plateau) by the genetic algorithm [107]. Unlike [103], the parameters were derived from the cortex rather than from the whole kidney and the system was tested on 50 patients, and the overall diagnostic accuracy increased to 96% Another study by Khalifa et al. [108] extended the work in [103, 104] by using analytical function-based model to fit agent cortical kinetic curves. For the classification of kidney status, five features (three were derived from the gamma-variate functional model and two are from the perfusion data, namely the time-to-peak and average plateau) were chosen and the study included 50 transplant patients.

Although DCE-MRI has been employed as a widespread imaging technique to develop several CAD systems for renal transplants assessment purpose, the contrast agents may implicate nephrogenic systemic fibrosis; thus, many medical centers are reluctant in
applying the DCE-MRI to patients with renal disease [109]. In order to circumvent this major drawback, DW-MRI and BOLD-MRI have been recently exploited to assess renal transplants as they do not involve any use of contrast agents, like DCE-MRI. A brief discussion of recent renal transplant assessment studies using BOLD-MRI follows as well as a discussion on other studies that utilized DW-MRI to assess renal transplants.

b. Blood Oxygen Level Dependant (BOLD) MRI: In addition to DCE-MRI, another imaging technique, called BOLD-MRI, has been utilized to study renal rejection using the amount of oxygen diffused blood (i.e. oxygen bioavailability) in the kidney to determine whether it is functioning properly. Specifically, the amount of deoxyhemoglobin is measured by the apparent relaxation rate (R2*) parameter [110]. Figure 7 shows grey images and R2* color-maps for a normal kidney and a kidney with graft dysfunction.

FIGURE 7: A simple demonstration of grey and R2* colored images for a normal kidney and a kidney with graft dysfunction.

In particular, Djamali et al. [111] investigated the ability of BOLD-MRI to assess
renal allografts. In their study, 23 patients underwent BOLD-MRI scans, of which 5 were normal allografts and 18 had acute allograft dysfunction (5 with ATN and 13 with AR). Medullary and cortical ROIs were placed, and mean cortical (CR2*), medullary (MR2*), and medullary to cortical (MCR2*) were calculated. They found that MR2* and MCR2* values of patients with ATN and AR were significantly decreased more than those with normal allografts. However, no differences in CR2* values between the different groups were observed. In a similar study by Han et al. [112], BOLD-MRI was conducted to differentiate between patients with AR and ATN after transplantation. Their study included 110 patients, 82 with normal allografts (group 1) and 28 with kidney dysfunction, including 21 with AR (group 2) and 7 with ATN (group 3). Group 2 was divided into two subgroups: 13 patients with T-cell-mediated rejection (TMR) and 8 patients with antibody-mediated rejection (AMR). Manual ROIs were placed in the cortical and medullary regions, and CR2*, MR2*, and MCR2* were compared between different groups. They performed a statistical analysis, and they found that values of CR2*, MR2*, and MCR2* of group 2 were reduced compared to those of the other two groups. Contradictory to the Djamali et al. [111] study, they found that values of MR2* of group 3 were higher than those of group 1. However, no significant difference was observed between the TMR and AMR subgroups.

Sadowski et al. [113] employed BOLD-MRI to assess kidney transplants. Manual cortical and medullary ROIs were placed on 17 patients who underwent BOLD-MRI scans, and these patients were divided into three groups: 5 patients with normal allografts (group 1), 4 with ATN (group 2), and 8 with AR (group 3). The MR2* and CR2* were calculated in the same way as was done in their previous study [111], and compared between the different groups. Specifically, MR2* values of group 3 allografts were decreased compared to those of group 1 and group 2, while no significant difference was observed in MR2* values between group 1 and group 2. However, no difference was detected in CR2* values among the three groups. Another interesting study by Liu et al. [110] was investigated to detect renal allograft rejection using BOLD-MRI. A total number of 50 patients with renal allografts were included and divided into three groups as 35 patients with normal allografts
(group 1), 10 patients with AR (group 2), and 5 patients with ATN (group 3). After cortical and medullary ROIs placement, CR2* and MR2* were measured to assess the three groups. Group 2 had the lowest MR2*, while no significant difference was detected in CR2* values among the three groups.

Although BOLD-MRI is a valuable imaging technique that has been investigated by some researchers in detecting renal allografts dysfunction, BOLD-MRI remains challenging, not only because of the low signal-to-noise ratio (SNR) and the weakness of the electromagnetic fields [114], but also the limited applicability of renal BOLD-MRI due to kidney motions and susceptibility induced by bowel gas which may lead to impaired image quality [115].

c. Diffusion-Weighted (DW) MRI: Recently, DW-MRI has become a subject of extensive research as an emerging imaging modality for renal function assessment thanks to DW-MRI’s ability to provide both anatomical and functional information, while avoiding radiation exposures (like CT) and contrast agents administration (like DCE-MRI). For DW-MRI, its functional parameter, called apparent diffusion coefficient (ADC), is estimated from different gradient field strengths and duration (b-values), as shown in Figure 8, to describe the unique tissue characteristics of inner spatial water behavior [115]. Therefore, several studies have utilized DW-MRI to assess renal functionality by measuring the ADC values, but the results have varied [110].

FIGURE 8: A demonstration of a DW-MRI sequence at different b-values.

Eisenberger et al. [116] investigated the manually placed ROIs in the upper, middle, and lower poles of the cortex and medulla on several slices to cover large regions of the allograft. Means and standard deviations of the ADC from all b-values were measured. The
ADC combines the perfusion free ADC and microcirculation parameters, quantified with perfusion fraction, $F_p$. These parameters were significantly reduced in the cortex and the medulla for the AR and ATN cases, and their values correlated with the creatinine clearance (CrCl). Similarly, a recent study by Hueper et al. [117] included 64 patients with renal allografts, of which 33 were patients with initial graft function (IniGF) and 31 were patients with DGF. These patients underwent DW-MRI scans at two $b$-values (0 and 600 $s/mm^2$). After placement of manual ROIs and estimation of renal diffusion parameters, including ADC and $F_p$, they concluded that renal diffusion parameters were significantly reduced in patients with DGF and their values well correlated with renal function and renal allograft fibrosis in biopsy specimens. The feasibility of diagnosing the acute renal transplant rejection (ARTR) from the DW-MRI was evaluated by Xu et al. [118] on 26 biopsy-proven rejection and 43 non-rejection patients. The non-rejection patients showed higher ADCs than those of the rejection group, and also demonstrated the best sensitivity and specificity at the $b$-value of 800 $s/mm^2$, as was evinced by the ROC curve. Palmucci et al. [119] evaluated functionality of 21 transplanted kidneys by comparing the estimated ADCs and true diffusion (TD) with renal function indices. Patients were divided into three groups by their CrCl values. The cortical ADC and TD were evaluated in a user-defined ROI of the transplanted kidney for the three groups. A moderate positive correlation between the CrCl and both the ADC and TD, as well as no difference between the ADC and TD values for the adjacent groups, has been found. The subsequent extension [120] of these evaluations to 35 patients revealed a slightly smaller positive correlation than the previously reported one [119]. However, acute rejection responses after transplantation could not be detected.

Vermathen et al. [121] assessed renal functionality by determining long-term (3 years) stability and potential changes for renal allograft recipients. After selecting cortical and medullary ROIs, the ADC had been calculated from all $b$-values. For good allograft, a significant correlation between different ADC components was observed, whereas for reduced allograft, the $F_p$ values were the highest, and the medullary $F_p$ had the greatest changes. Katarzyna et al. [122] investigated possible relations between the selected laboratory results and diffusion parameters in the early period after kidney transplantation by mea-
suring additional exponential ADCs to overcome the DW-MRI T2-“shine-through” [123].

The measurements were conducted in the kidney’s cortex and medulla over multiple user-defined ROIs at the \( b \)-values of 600 and 1000 only. According to relative variability of results and SNR, the best-quality ADC measurement in the renal cortex was at the \( b \)-value of 1000 \( s/mm² \). Also, there were strong dependencies between the ADC and exponential ADC, measured in the renal cortex at \( b_{1000} \ s/mm² \), and the estimated GFR. Kaul et al. [124] assessed the renal dysfunction with cortical and medullary ADC maps. They reported a significant decrease in the ADC values of medullas compared to those of cortexes in normal donor kidneys and normally functioning transplanted kidneys. Both the medulla and cortex ADCs decreased or increased significantly for a rejection or the recovery from the rejection itself, respectively. A recent study by Abou-El-Ghar et al. [125] included 70 renal allograft patients who underwent DW-MRI scans at two \( b \)-values (0 and 800 \( s/mm² \)). In this case, 49 patients had stable renal allograft function (group 1) and 21 patients had acute graft impairment (group 2: 10 acute cellular rejection (ACR), 7 ATN, and 4 ITox rejection types). An ROI was placed at the middle of the kidney in a single cross-section and a pixel-wise ADC was calculated. They have shown that the ADC values of group 1 were significantly higher than those of group 2, and no overlap was detected between the ADCs of group 1 and the ATN patients of group 2. However, the minimal overlap was observed between the ADCs of group 1 and the patients with the ACR and IT of group 2. Recently, Liu et al. [110] detected an early renal allograft dysfunction caused by AR and ATN using the DW-MRI and BOLD-MRI with manually selected cortical and medullary ROIs. Their study revealed lower values of both the measured apparent relaxation rates and ADC for the AR group than for the control groups, and no difference in the ADC values for the AR and ATN groups. A similar earlier study was conducted by Thoeny et al. [126].
F. Chapter Summary

Though the treatment of chronic kidney disease has improved greatly with the use of transplants, there are still challenges such as graft dysfunction that provide a challenge in maintaining survival of the new organ. In the future, the use of image-based diagnosis will be improved and implemented in the diagnosis of both pre- and post-transplantation. It is hoped that by having these improved imaged based CAD systems that diagnosis of graft dysfunction, along with the cause of graft dysfunction, will be less invasive, more accurate, time saving, and inexpensive compared to renal biopsies and other traditional methods of diagnosis. By having all of these advantages it is expected that graft survival will improve in cases of graft dysfunction.

G. Thesis Organization

This thesis is presented in three chapters. The scope of each chapter is summarized as follows:

1. Chapter I

This chapter overviews the existing traditional clinical methods and the current image-based techniques which are used to evaluate and assess renal transplants functionality and to detect early signs of graft dysfunction. Additionally, a brief overview is given in this chapter including: kidney anatomy and functionality, the risk of developing chronic kidney disease (CKD) and some symptoms associated with it, an overview of the treatment options for people that develop stage 5 CKD, transplantation procedure as a definitive therapy for these people with stage 5 CKD, and post transplantation follow-ups which includes possible complications that could arise with a concentration on graft dysfunction. This is followed by the traditional tests that are performed to detect graft dysfunctions such as blood, urine, and renal biopsy. Finally, the state-of-the-art image based techniques which are used in renal transplants’ evaluation and assessment are discussed. These im-
age techniques include: ultrasounds (US) (e.g., conventional, power doppler (PD), color doppler (CD), contrast enhanced (CE), etc.) , and magnetic resonance imaging (MRI)(e.g., dynamic contrast enhanced (DCE), blood oxygen level-dependent (BOLD), and diffusion-weighted (DW)).

2. Chapter II

This chapter presents a new computer-aided diagnostic (CAD) system for the early detection of acute renal transplant rejection (ARTR) using diffusion-weighted (DW) MRI. The developed CAD system demonstrates multiple novelties including: (i) segmenting kidneys from the DW-MRI data in a fully automated mode using geometric deformable models; (ii) describing kidney functionality with the cumulative distribution function (CDF) of the ADCs; and (iii) discriminating between rejection and non-rejection renal transplants with a deep neural network learned by stacking layers of several auto-encoders with non-negativity constraint (NCAE).

3. Chapter III

This chapter concludes the thesis, highlights the main contributions and obtained results, and discusses the trend for possible future avenues to be handled.
CHAPTER II
COMPUTER-AIDED DIAGNOSTIC SYSTEM FOR EARLY DETECTION OF ACUTE RENAL TRANSPLANT REJECTION USING DIFFUSION-WEIGHTED MRI

A. Introduction

Due to the fact there are up to 17,000 renal transplants per annum in the U.S. and a limited number of donors [127], the salvage of a transplanted kidney is of serious clinical concern. The immunological response of a patient’s body to a transplanted kidney, called the acute renal transplant rejection (ARTR), is considered to be the leading cause of renal dysfunction [127] after the transplantation. An early detection of renal dysfunction increases the survival rate of the transplanted kidney [110, 125], as confirmed by clinicians. Therefore, calling for essential medical biomarkers to assess renal transplants, especially at an early stage, (i.e. before major changes in creatinine clearance (CrCl) and serum plasma creatinine (SPCR) are detected), is very necessary to distinguish the ARTR from other diagnoses, including the acute tubular necrosis (ATN) and immune drug toxicity.

Traditional blood tests and urine sampling to evaluate renal transplant dysfunction cannot assess function of individual kidneys. The glomerular filtration rate (GFR) is based on measuring the serum creatinine level and has been approved by the National Kidney Foundation (NKF) to evaluate the overall kidney function. The GFR is a relatively imprecise and late marker for renal dysfunction (a significant change in creatinine levels is only detectable after losing 60% of renal function) [6]. Biopsy, which remains the gold standard for renal transplant assessment, is an invasive procedure with a high cost and morbidity rate, and relatively small needle biopsy samples may over- or under-estimate an inflammation extent in the entire graft [127]. More favorable non-invasive imaging tests provide separate information on each kidney. However, the most frequent scintigraphy, preferred for
its good functional information, has too low spatial resolution [128] and exposes patients to a small dose of radioactivity because of reliance on gamma-cameras [129]. Computed tomography provides superior functional and anatomical information, but uses nephrotoxic contrast agents and exposes patients to radiation as well [128]. These shortcomings have been circumvented recently by evaluating kidney functions with magnetic resonance imaging (MRI). For example, the dynamic contrast-enhanced (DCE) MRI has been exploited for renal function assessment due to providing both anatomical and functional kidney information [91, 92, 96, 109]. However, because the contrast agents may cause the development of nephrogenic systemic fibrosis, many medical centers are reluctant in applying the DCE-MRI to patients with renal disease [101].

In order to circumvent this major drawback, diffusion-weighted (DW) MRI has been exploited as an emerging imaging modality for renal function assessment. The DW-MRI measures unique tissue characteristics of inner spatial water behavior, namely, its apparent diffusion coefficients (ADC). In this chapter, a CAD system for the early detection of ARTR using DW-MRI is developed, thanks to DW-MRI’s ability to provide both anatomical and functional information, exposes patients to no radiation, and needs no contrast agents, like the DCE-MRI. The proposed CAD system demonstrates multiple novelties including: (i) segmenting kidneys from the DW-MRI data in a fully automated mode (Section II.B.1); (ii) describing kidney functionality with the cumulative distribution function (CDF) of the ADCs (Section II.B.2); and (iii) discriminating between kidney rejection and non-rejection with a deep neural network learned by stacking layers of several auto-encoders with non-negativity constraint (NCAE) (Section II.B.3). Section II.A.1 below briefly outlines state-of-the-art techniques in DW-MRI segmentation, followed by Section II.A.2 which reviews the prior work on assessing the renal transplant function using the ADCs.

1. Prior Work on DW-MRI Segmentation

The process of segmenting kidneys from the DW-MRI has not been addressed yet. In literature, only known applications to other anatomical structures, such as the brain, the
prostate, and the liver, can be reviewed.

**Brain segmentation.** To segment the brain tissue in a fully automated mode, Yap et al. [130] used voxel-wise compartmental coexistence modeling of various diffusion features. After possible signal contributions from different brain tissues are assigned to each voxel using diffusion exemplars, each voxel is classified in accordance with the least fitting residual. Mujumdar et al. [131] segmented stroke lesions by using the Chan-Vese [2] deformable boundary guided with the Mumford-Shah functional. An automated window-based noise suppression at high $b$-values was conducted to enhance local contrast of the candidate lesions. Automated region growing by Saad et al. [132, 133] integrated homogeneity criteria and simple statistical regional features, such as the mean signal intensity and the number of pixels. However, both this approach and its semi-automated variant could not fully characterize the tumor lesion. Saad et al. [134] explored an automatic brain lesion segmentation, which starts from the DW-MRI normalization, background removal, and enhancement. Then both regional and boundary information, combined with statistical texture descriptors extracted from a collected gray level co-occurrence matrix, were used to guide the segmentation of brain lesions (e.g., hyperintense or hypointense). To segment near-tubular fiber bundles from the DW-MRI, Niethammer et al. [135] performed global statistical diffusion orientation modeling that utilized optimal paths or simple streamlining to obtain geometric information. Convex approximation of the probabilistic Chan-Vese energy [2] was employed using region-based directional statistics.

**Prostate segmentation.** McClure et al. [136] used geometric (level-set-based) deformable boundaries and nonnegative matrix factorization (NMF) for an automated prostate segmentation from the DW-MRI. The NMF integrated the DW-MRI intensity, a prostate shape prior, and pairwise spatial interactions between the prostate voxels. The like segmentation and maximum a posteriori decisions were used by Firjani et al. [137] for early detection of prostate cancer. An unsupervised level-set-based prostate segmentation by Liu et al. [138] applies a shape penalty term described by an elliptical deformable boundary to initialize and constrain the level-set function. The latter is then refined by connectivity and morphological analysis. Subsequently, their framework was extended to account for the 3D ADC
images derived from the original DW-MRI [139]. After an initial coarse segmentation, a shape prior weighting parameter is selected automatically, and refining morphological operations are performed to obtain the final segmentation map. Ozer et al. [140] tuned thresholds of support vector machines (SVM) and relevance vector machines (RVM) to classify “multispectral” MRI data, e.g., dynamic MRI, quantitative T2 MRI, and DW-MRI, for automated prostate segmentation in accordance with an user-defined performance criterion, such as optimal accuracy, maximal sensitivity, etc. Then, the best performance was obtained by using a first-order polynomial kernel in the SVMs and RVMs. A fuzzy Markov random field (MRF) modeling of the like multispectral MRI prostate data allowed Liu et al. [141] to simultaneously perform an unsupervised prostate segmentation and MRF parameter estimation.

Liver segmentation. Veeraraghavan et al. [142] applied simultaneous segmentation and iterative registration to the liver DW-MRI data. At each iteration, the rigid global affine registration is followed by the the fine non-rigid B-spline-based registration. A sequence of transformations is used to most accurately align the individual $b$-value images to the reference image, the modified Housdorff distance (MHD) being a similarity metric. Then, a GrowCut optimization segments the whole volume of interest after one or more 2D slices were manually segmented in the reference image. Stephen et al. [143] developed a semi-automated lesion segmentation and ADC estimation. For every given set of patient images, a large rectangular ROI is created only once manually by the analyst in one of the images, ensuring that the lesion is within the ROI in all of the images for all the $b$-values and some pixel within the lesion region is marked as a seed. For each image, an empirical probability distribution pixel intensities inside the ROI is approximated with a finite Gaussian mixture (FGM), a Markov-Gibbs random field (MGRF) model is built to quantify spatial pixel-to-pixel dependencies (interactions), and these two statistical descriptors are combined to segment the lesion within the ROI for each image. To improve the segmentation, geometric convexity constraints of a lesion are taken into account. A semi-automated joint liver segmentation and alignment framework by Veeraraghavan and Do [144] begins from a sequential least-squares alignment of the images. Then various structures in a randomly
selected reference image are manually segmented, starting from user-drawn lines on these structures. The closest image, selected with mean-shift tracking and aligned to the reference image using non-rigid B-spline-based deformations, is selected as the new reference image, and the process is repeated for all remaining images in the sequence. Jha et al. [145] developed an automated statistical clustering approach for segmenting liver lesions. The DW-MRI intensity distribution is approximated by an FGM, such that its number of Gaussian components (classes) is determined by measuring the approximation error, and an MRF model is used to quantify spatial pixel interactions, as well.

2. Prior Work on Renal Function Assessment

Prior related work on renal function assessment using DW-MRI has been covered in details in Chapter I (Section I.E.4.c).

In total, several clinical studies have been conducted for the assessment of renal function. However, they have some limitations. All the approaches employ manual delineation of the kidney using 2D ROIs, which is subjective and depends on user knowledge of anatomy. In addition, these methods did not compensate for the kidney motion since they did not account for the entire kidney. Furthermore, several of these studies performed only statistical analysis to investigate the significant difference between pairs at certain $b$-values and did not integrate all the analysis steps into a whole framework to build a fully automated CAD system. Finally, the studies mentioned above did not investigate the fusion of ADCs at multiple low and high $b$-values.

As an initial idea to overcome these limitations, a recent study to distinguish between rejection and non-rejection renal transplants was made by [146]. Their study included 36 renal transplants of which 6 were non-rejection and 30 were rejection. After DW-MRI data motion correction using a 2D B-splines approach, they segmented the largest coronal cross-section of the kidney using a fully automated level sets segmentation approach. Then, they calculated the ADCs at different $b$-values from the segmented coronal cross-section for each subject. By using a leave-one-subject-out scenario along with
a KStar classifier, they got a 87% total classification accuracy. In addition, they depicted color-maps from the calculated ADCs for the visualization purpose at different $b$-values. However, this study did not compensate for kidney motion as as it did not account for the entire kidney volume. In addition, it did not investigate the fusion of multiple ADCs at multiple low and high $b$-values for the entire kidney volume.

These initial promising diagnostic results obtained by [146] was the trigger to extend this work to overcome all of the aforementioned shortcomings. Therefore, a 4D (3D + $b$-value) fully automated CAD system has been developed [147, 148], shown in Figure 9, which is able to: (i) delineate the whole kidney and handle its motion; (ii) fuse the ADC values calculated from the segmented DW-MRI data sets acquired at multiple low and high $b$-values; (iii) describing kidney functionality with the CDFs of the ADCs; and (iv) discriminating between kidney rejection and non-rejection with a deep neural network learned by stacking layers of several auto-encoders with non-negativity constraint (NCAE). Experimental results hold promise for the developed CAD system as a reliable non-invasive diagnostic tool.

FIGURE 9: Proposed CAD system for detecting renal rejection from 4D DW-MRI.
B. Methods

Given an input 4D (3D + \( b \)-value) DW-MRI, the proposed CAD system, shown in Figure 9, performs the following steps: (i) segments kidneys from surrounding abdominal structures (Section II.B.1); (ii) estimates voxel-wise functional parameters (ADCs) to form a 3D parametric map for detecting status of the transplanted kidney (Section II.B.2); and (iii) classifies acute rejection or non-rejection transplant status in order to evaluate the system as a diagnostic test (Section II.B.3).

1. 3D Kidney Segmentation

Since the segmentation is a key step in developing any CAD system, the presented CAD starts with segmenting the kidney from the surrounding tissues using DW-MRI. However, accurate kidney segmentation is a challenging task for many reasons, including: kidney motion due to breathing and heart beating; kidney shape changes due to inter-patient anatomical differences; low contrast between the kidney and other abdominal structures, especially, at the higher gradient strengths and duration, or \( b \)-values (Figure 10); low SNR and artifacts that complicate image alignment [149, 150]; and geometric distortions due to long acquisition time [130].

![FIGURE 10: Coronal cross-sections of raw DW-MRI samples showing (a) similar intensities of kidney and surrounding tissues (e.g., at \( b_0 \)), (b) inter-patient anatomical differences (e.g., at \( b_0 \)) compared to the cross-section (a), and (c) image noise, especially, at higher gradient strengths and duration (\( b \)-values) (e.g., at \( b_{1000} \)).](image-url)
To overcome these challenges, the proposed segmentation technique relies on multiple image features to accurately delineate the kidney and thus facilitates further analysis of transplant status. Basic notations and details of the proposed segmentation approach are outlined below.

For describing these processing steps, let \( p = (x, y, z) \) denote a voxel at position with discrete Cartesian coordinates \((x, y, z)\) and let \( R = \{(x, y, z) : 0 \leq x \leq X - 1; 0 \leq y \leq Y - 1; 0 \leq z \leq Z - 1\} \) be a finite 3D arithmetic lattice of unit voxels, which has the size of \( XYZ \) and supports both grayscale images and their parametric or region (segmentation) maps. A grayscale image, \( g = \{g_p : p \in R; g_p \in \mathbb{Q}\} \), takes voxel values from a finite set, \( \mathbb{Q} = \{0, 1, \ldots, Q - 1\} \), where \( Q \) integer gray levels, i.e. \( g : R \to \mathbb{Q} \). A region map, \( m = \{m_p : p \in R; m_p \in \mathbb{L}\} \), takes voxel values from a binary set of region labels, \( \mathbb{L} = \{0, 1\} \), where 0 and 1 indicate background and object (kidney), respectively, i.e. \( m : R \to \mathbb{L} \).

A 3D geometric, or level-set-based deformable boundary, is employed in the proposed CAD system for the DW-MRI kidney segmentation. Such a tool is common and successful in a wide range of applications, including various medical imaging tasks, e.g., segmenting brain, prostate, liver, kidney, etc. [104]. Compared to alternative parametric deformable boundaries, the geometric ones are more popular due to their simplicity, flexibility, and ability to handle complex geometries and topological changes independently of surface parameterizations. Points of an object-background boundary at each time instant \( t \) are specified implicitly as a zero-level set, \( B_t = \{p : p \in R; \Phi(p, t) = 0\} \), of arguments of a specific higher-dimensional function \( \Phi(p, t) \), being supported by the lattice \( R \) and often called a signed distance map:

\[
\Phi(p, t) = \begin{cases} 
  d(p, B_t) & \text{if } p \text{ is inside the boundary } B_t; \\
  0 & \text{if } p \text{ is at the boundary } B_t, \text{ and} \\
  -d(p, B_t) & \text{if } p \text{ is outside the boundary } B_t, 
\end{cases}
\]

Here, \( d(p, B_t) = \min_{b \in B_t} d(p, b) \) denotes the distance from the point \( p \) to the boundary \( B_t \), and \( d(p, b) \) is the Euclidean distance between the lattice points \( p \) and \( b \), as illustrated in Figure 11.
The function $\Phi(p, t)$ evolves in discrete time $t = n\tau$ with a fixed step, $\tau > 0$, as [154]:

$$
\Phi(p, (n+1)\tau) = \Phi(p, n\tau) - \tau F_n(p) |\nabla \Phi(p, n\tau)|
$$

where $n = 0, 1, 2, \ldots$, is the time index; $\nabla \Phi(p, n\tau)$ is the spatial gradient of $\Phi(p, n\tau)$:

$$
\nabla \Phi(p, n\tau) = \left[ \frac{\partial \Phi(p, n\tau)}{\partial x}, \frac{\partial \Phi(p, n\tau)}{\partial y}, \frac{\partial \Phi(p, n\tau)}{\partial z} \right];
$$

$|a|$ denotes the magnitude of the vector $a$, and $F_n(p)$ is a speed function guiding the evolution of an initial boundary $B_0$, which was defined at the starting instant $t = 0$ (i.e. $n = 0$).

Most of the conventional speed functions quantify visual appearance differences between the object and its background in terms of mean values and variances of image intensities, intensity edges or gradient vector flow, and similar regional signal characteristics. Thus, their guidance may fail if images to be segmented are noisy and/or object-background contrast is low. To accurately segment the kidneys from the noisy and low-contrast DW-MRI, the developed guiding function accounts for not only regional kidney-background appearance, but also for a kidney shape prior and spatial relationships of the goal region map. To provide the voxel-wise guidance for the evolving boundary, all the employed appearance and shape descriptors are combined into a joint MGRF model of a DW-MRI image, $g$, and its binary kidney-background region map, $m$. The model is specified by a joint probability distribution $P(g, m) = P(g|m)P(m)$, where $P(g|m)$ and $P(m)$ denote a conditional probability distribution of images given a map and an unconditional distribution of region maps, respectively. The latter distribution is factored into two terms: $P(m) = P_{sp}(m)P_V(m)$, where $P_{sp}(m)$ denotes an appearance-based adaptive shape prior.
and $P_{\psi}(m)$ is a second-order Gibbs probability distribution with potentials $\psi$ of multiple nearest-neighbor pairwise dependencies, which specifies a simple spatially homogeneous MGRF model of region maps. These components of the joint image-map model are outlined in the following Sections.

\textit{a. Appearance-Based Shape Prior:} In addition to the distinct visual appearances, the well-known geometric shapes of medical structures can enhance the segmentation accuracy. To rely on this fact, an adaptive model of the expected kidney shape is used to both handle kidney motions, e.g., due to breathing and/or heart beating, and account for the kidney’s variability due to inter-patient anatomical differences. In addition, the kidney DW-MR images are very noisy, especially at high $b$-values, and have low contrast between the kidney tissue and other abdominal structures. The noise and inconsistencies due to low-frequency non-uniformity, or heterogeneity of intensities, are suppressed in part by preprocessing, namely, histogram equalization with nonparametric bias correction by Tustison [155] shown in Figure 12.

![FIGURE 12: Raw coronal DW-MRI cross-section before (a) and after (b) its preprocessing.](image)

To build the shape prior, a shape database is created from a training set of manually delineated kidney images from different subjects by co-aligning these images using a 3D B-spline-based non-rigid transformation [156]. The alignment minimizes the sum of squared voxel-wise intensity differences between the two kidney images, and kidney/background labels of the co-aligned region maps are used to learn the shape prior. Figure 13 illustrates the co-alignment of the training DW-MRI with respect to a single reference image.
FIGURE 13: 3D co-alignment of training DW-MRI to a single reference: grayscale images before (a) and after (b) their alignment and overlapped 3D binary volumes before (c) and after (d) alignment, the reference image and targets being in yellow and red, respectively.

In the performed experiments, the shape database contained 53 data sets ($b_0$-scans) manually segmented by an MRI expert and then co-aligned. Adapting the shape prior to each input DW-MR image to be segmented is guided by visual appearance of the latter image. The probabilistic shape prior is built as a spatially variant independent random field of region labels $P_{sp}(m) = \prod_{p \in \mathbb{R}} \Pr_p(m_p)$, where $\Pr_p(l)$ is the marginal empirical probability of the label $l \in L$ in the voxel $p$: $\sum_{l \in L} \Pr_p(l) = 1$. Algorithm summarizes estimating and updating the appearance-guided shape prior for each test DW-MR image to be segmented (the test images are first removed from the training set).

b. Second-Order MGRF Model of Region Maps: In order to increase the segmentation accuracy, 3D pairwise dependencies between the region labels are additionally incorporated in the proposed model by using a popular Potts MGRF [157]. Here, it is a spatially homogeneous binary field with the nearest 26-voxel neighborhoods and analytically estimated bi-valued Gibbs potentials depending only on whether or not the nearest pairs of labels are equal. Let $f_{eq}$ denote the empirical marginal probability of equal labels
Algorithm 1 Creating / Updating the Shape Prior.

1 Preprocess the training DW-MR images by bias correction and histogram equalization.

2 Construct the shape database by applying the co-alignment by Glocker et al. [156] to the preprocessed DW-MR Images.

3 Preprocess the DW-MR image for a test subject and co-align with the shape database.

4 For each voxel, \( p \in \mathbb{R} \), in the test DW-MR image, \( g_{\text{test}} \), calculate its prior shape probabilities, \( \Pr_p(l); l \in \mathbb{L} \), as follows:
   A Use the co-aligning deformation field to relate the voxel \( p \) of the test image to the shape database lattice.
   B Construct a 3D window with initial size of \( N_1 \times N_2 \times N_3 \), centred at the related voxel in the shape database lattice.
   C Find within the window all the voxels with the corresponding intensity, \( g_{\text{test};p} \), in all the training images.
   D If necessary, increase the window size and repeat Steps 4B to 4D until a non-empty set of such corresponding training intensities is found.
   E Estimate label probabilities based on relative occurrences of each label in all the training voxels found.

In the neighboring voxel pairs \((p, p + \delta) \in \mathbb{R}^2; \delta \in \mathbb{N}_{26} = \{(\pm 1, 0, 0), (0, \pm 1, 0), (0, 0, \pm 1), (\pm 1, 1, 0), (\pm 1, 0, \pm 1), (0, \pm 1, \pm 1), (\pm 1, \pm 1, 0), (\pm 1, \pm 1, \pm 1)\})

Given a region map \( m \) (e.g., after an initial rough segmentation), the maximum likelihood estimates of the potentials

\[
V(\mathbf{m}_p, \mathbf{m}_{p+\delta}) = \begin{cases} 
V_{\text{eq}} & \text{if } \mathbf{m}_p = \mathbf{m}_{p+\delta}; \mathbf{p} \in \mathbb{R}; \mathbf{\delta} \in \mathbb{N}_{26}; V_{\text{eq}} + V_{\text{ne}} = 0 \\
V_{\text{ne}} & \text{if } \mathbf{m}_p \neq \mathbf{m}_{p+\delta}
\end{cases}
\]

are analytically approximated [158] as follows: \( V_{\text{eq}} = -V_{\text{ne}} \approx 2f_{\text{eq}}(\mathbf{m}) - 1 \). This approximation is used for computing the voxel-wise probabilities \( \Pr_{V_p}(l) \) of each label; \( l \in \mathbb{L} \).

c. First-Order Kidney/Background Appearance Model: To accurately model the DW-MRI appearance, the empirical marginal probability distribution of intensities is approximated with a linear combination of discrete Gaussians (LCDG) [159]. The LCDG with two positive dominant components (one each for the kidney and background) and
multiple sign-alternate subordinate components allow for separating the mixed marginal of the DW-MRI voxel-wise intensities into the two distinct LCDGs, each associated with the kidney or background label. This LCDG model adapts the segmentation to changing appearance, such as non-linear intensity variations caused by patient weight and the data acquisition system, and it separates individual submodels of the kidney and background intensities more accurately than a conventional mixture of only positive Gaussians. This adaptation yields a better initial region map after the voxel-wise classification of only the image intensities with no account for the kidney shape.

d. Appearance- and Shape-Guided Deformable Model: Adaptation to both the kidney-background visual appearance, shape prior, and statistical spatial dependencies between kidney labels is one of the main advantages of the proposed segmentation framework. Estimated directly from the input image and a given shape database, these properties guide the evolving deformable boundary by defining, for each voxel $p$ with intensity $g_p = q$, the speed function [104] of Equation (1), $F_n(p) = \kappa \vartheta_p$, where $\kappa$ is the mean contour curvature and $\vartheta_p$ specifies the magnitude and direction of moving that voxel:

$$\vartheta_p = \begin{cases} -Pr_p(1) & \text{if } Pr_p(1) > Pr_p(0) > 0.5; \\ Pr_p(0) & \text{otherwise} \end{cases}$$

(2)

Here, $Pr_p(0)$ and $Pr_p(1)$ are the voxel-wise background and kidney probabilities, respectively:

$$Pr_p(1) = \frac{\Omega_{kd:p}}{\Omega_{kd:p} + \Omega_{bg:p}}; \quad Pr_p(0) = \frac{\Omega_{bg:p}}{\Omega_{kd:p} + \Omega_{bg:p}} = 1 - Pr_p(1)$$

where the variables $\Omega_{kd:p}$ and $\Omega_{bg:p}$ for the kidney and background, respectively, depend on the voxel-wise probabilities $Pr(q|l); l \in L$, for the LCDG submodels of the kidney ($l = 1$) or background ($l = 0$) appearance and on the kidney label probability in the MGRF spatial region map model, $Pr_V(p, 1)$, and in the adaptive shape prior, $Pr_{sp:p}(1)$, respectively:

$$\Omega_{kd:p} = Pr(q|1) Pr_V(p, 1) Pr_{sp:p}(1); \quad \Omega_{bg:p} = Pr(q|0)(1 - Pr_V(p, 1)(1 - Pr_{sp:p}(1))$$

Algorithm summarizes the basic steps of the 3D level-set-based kidney segmentation.
Algorithm 2 DW-MRI Segmentation by Geometric Deformable Boundary

1. Update the shape prior probability using Step 4 of Algorithm 1.

2. Approximate the marginal of DW-MRI intensities with the LCDG [159] with two dominant components.

3. Form an initial region map, \( m_{\text{ini}} \), using the estimated shape prior and LCDG submodes of kidney and background appearances.

4. Estimate the Gibbs potentials for the spatial MGRF map model from \( m_{\text{ini}} \).

5. Find the above speed function [104], \( F_n(p) \), using results of Steps 2 to 4.

6. Segment the input image, \( g \), by evolving the level-set function, \( \Phi(p, n\tau) \), of Equation (1) with the speed function found in Step 5.

2. Estimating and Depicting Diffusion Parameters

After segmenting the kidneys, their discriminatory functional features are estimated from the images and used to distinguish between rejection and non-rejection of kidney transplants. In this chapter, the ADC defined by Le Bihan [160] is used as a transplant status feature. This voxel-wise ADC is defined as:

\[
\text{ADC} = \frac{1}{b_0 - b} \ln \left( \frac{g_{b,p}}{g_{0,p}} \right) = \frac{\ln g_{b,p} - \ln g_{0,p}}{b_0 - b}
\]

where the segmented DW-MR images \( g_0 \) and \( g_b \) were acquired with the \( b_0 \) and a given different \( b \)-value, respectively. To reduce the noise effects on ADC estimation, the ADC at each voxel \( p \) (3D location of the DW-MRI data) is calculated using a \( 3 \times 3 \times 3 \) cube around \( p \), and \( g_{0,p} \) and \( g_{b,p} \) are represented by the average signal intensity of that cube.

It is worth noting that conventional classification methods that deal directly with the voxel-wise ADCs of the entire kidney volume as discriminative kidney features encounter two difficulties: (i) varying input data size requires either data truncation for larger kidney volumes or zero padding for smaller ones and (ii) large data volumes lead to considerable time expenditures for training and classification. In order to overcome the above challenges, the entire 3D ADC maps, collected for each subject at the 11 different \( b \)-values, is characterized by the CDFs of the ADCs, as shown in Figure 14. These descriptors are independent of the initial data size and can be quantified in accordance with the actual ac-
accuracy (signal-to-noise ratio) of the ADCs. Preliminary experiments have shown that the 1%-accuracy of measuring the ADC to within a range between the maximum and minimum ADCs for all the b-values and subjects is sufficient, as regarding the final classification accuracy. Comparing to the empirical probability distribution functions (PDFs) of the ADCs, the CDFs allow better differentiation between the PDFs across the whole range of the ADC values. The training CDFs are used for deep learning of a stacked NCAE (SNCAE) classifier detailed in Section II.B.3. Fixing the input data size to 11 for such CDFs helps to overcome the above challenges for arbitrary sizes of the original ADCs and notably accelerates the classification. Also, the estimated 3D ADCs can be displayed as voxel-wise parametric maps to be visually assessed by the radiologists.

3. Autoencoding and Deep Learning-Based Classifier

A rich variety of learnable classifiers with shallow structure [161–164] have been used in the CAD systems for organ transplantation prediction using clinical and demographic data of patients, including (but not limited to) artificial neural networks (ANN), support vector machines (SVMs), regression trees, random forests (RFs), decision trees (DTs), k-nearest neighbor (kNN), etc. However, the aforementioned popular learnable classifiers and predictors have some limitations [165, 166]: (i) some of them (e.g., RFs and DTs) cannot deal with very large scale amounts of data, which are typical for DW-MRI; (ii) some of them (e.g., SVMs) depend mainly on the selection of the kernel function and
its parameters; (iii) many classifiers (e.g., SVMs, RFs) are of high algorithmic complexity and require extensive memory.

Recent advances in deep learning of the ANNs [167] allow for overcoming these drawbacks of the classical shallow models: (i) by automated dimensionality reduction of large scale data [168]; (ii) and automatic extraction of more discriminatory features between classes through hierarchical feature extraction. In this case, the high level (global) features are derived from the low level (local) features for model training [168–171]; (iii) and flexibility compared to the classical shallow models, i.e., a classifier (e.g., a softmax-based or SVM-based) can be built on the extracted features from the deep learning ANN [168, 172].

Due to the aforementioned advantages, the proposed fully automated CAD system utilizes deep learning and auto-encoders with non-negativity constraint (NCAE) as a core ANN architecture for pre-training and classification to distinguish between the non-rejection and acute rejection of kidney transplants. In particular, the presented CAD system employs a deep neural network with a stack of auto-encoders (AE) before the output layer that computes a softmax regression, generalizing the common logistic regression to more than two classes. Each AE compresses its input data to capture the most prominent variations and is built separately by greedy unsupervised pre-training [173]. The softmax output layer facilitates the subsequent supervised back-propagation-based fine tuning of the entire classifier by minimizing the total loss (negative log-likelihood) for a given training labeled data. Using the AEs with a non-negativity constraint (NCAE) [174] yields both more reasonable data codes (features) during its unsupervised pre-training and better classification performance after the supervised refinement.

Let $W = \{W^e_j, W^d_i : j = 1, \ldots, s; i = 1, \ldots, n\}$ denote a set of column vectors of weights for encoding (e) and decoding (d) layers of a single AE in Figure 15. Let $\mathbf{T}$ denote vector transposition. The AE converts an $n$-dimensional column vector $\mathbf{u} = [u_1, \ldots, u_n]^T$ of input signals into an $s$-dimensional column vector $\mathbf{h} = [h_1, \ldots, h_s]^T$ of hidden codes (features, or activations), such that $s \ll n$, by a uniform nonlinear transformation of $s$
weighted linear combinations of signals:

\[ h_j = \sigma \left( \left( W^e_j \right)^T u \right) \equiv \sigma \left( \sum_{i=1}^{n} W^e_{ji} u_i \right) \]

FIGURE 15: Block-diagram of an NCAE (a) and an SNCAE (b) classifier.

where \( \sigma(\ldots) \) is a certain sigmoid, i.e. a differentiable monotone scalar function with values in the range \([0, 1]\). Unsupervised pre-training of the AE minimizes total deviations between each given training input vector \( u_k; k = 1, \ldots, K \), and the same-dimensional vector, \( \hat{u}_{W,k} \) reconstructed from its code, or activation vector, \( h_k \) (Figure 15(a)). The total reconstruction error of applying such AE to compress and decompress the \( K \) training input vectors integrates the \( \ell_2 \)-norms of the deviations:

\[ J_{AE}(W) = \frac{1}{2K} \sum_{k=1}^{K} \| \hat{u}_{W,k} - u_k \|^2 \]  

To reduce the number of negative weights and enforce sparsity of the NCAE, the reconstruction error of Equation (4) is appended, respectively, with quadratic negative weight penalties, \( f(w_i) = (\min\{0, w_i\})^2; i = 1, \ldots, n \), and Kullback-Leibler (KL) divergence, \( J_{KL}(h_W; \gamma) \), of activations, \( h_W \), obtained with the encoding weights \( W^e \) for the training
data, from a fixed small positive average value, $\gamma$, near 0:

$$J_{\text{NCAE}}(W) = J_{\text{AE}}(W) + \alpha \sum_{j=1}^{s} \sum_{i=1}^{n} f(w_{ji}) + \beta J_{\text{KL}}(h_{W}; \gamma) \quad (5)$$

Here, the factors $\alpha \geq 0$ and $\beta \geq 0$ specify relative contributions of the non-negativity and sparsity constraints to the overall loss, $J_{\text{NCAE}}(W)$, and

$$J_{\text{KL}}(h_{W}; \gamma) = \sum_{j=1}^{s} h_{W_{c,j}} \log \left( \frac{h_{W_{c,j}}}{\gamma} \right) + (1 - h_{W_{c,j}}) \log \left( \frac{1 - h_{W_{c,j}}}{1 - \gamma} \right) \quad (6)$$

The classifier is built by stacking the NCAE layers with an output softmax layer, as shown in Figure 15(b). Each NCAE is pre-trained separately in the unsupervised mode by using the activation vector of a lower layer as the input to the upper layer. Here, the initial input data consisted of the 11 CDFs, each of size 100, i.e. $n = 1100$. In other words, for quantizing the ADCs, the range between the minimum and maximum ADCs for all the input data sets (i.e. all the sets for 11 $b$-values and 53 subjects) was divided into 100 steps to keep the chosen 1%-accuracy of initial ADC measurements. The PDFs and then CDFs of the ADCs were built for these quantized values. The bottom NCAE compresses the input vector to $s_1$ first-level activators, compressed by the next NCAE to $s_2$ second-level activators, which are reduced in turn by the output softmax layer to $s^o$ values. The number of the NCAE layers and successive data compression ratios for each layer were chosen empirically, on the basis of comparative experiments.

Separate pre-training of the first and second layers by minimizing the loss of Equation (5) reduces the total reconstruction error, as well as increases sparsity of the extracted activations and numbers of the non-negative weights. The activations of the second NCAE layer, $h^{[2]} = \sigma(W_{c,1}^{[2]} h^{[1]})$, are inputs of the softmax classification layer, as sketched in Figure 15(b) to compute a plausibility of a decision in favor of each particular output class, $c = 1, 2$:

$$p(c; W_{o,c}) = \frac{\exp(W_{o,c}^{T} h^{[2]})}{\exp(W_{o,1}^{T} h^{[2]}) + \exp(W_{o,2}^{T} h^{[2]})} ; \quad c = 1, 2; \quad \sum_{c=1}^{2} p(c; W_{o,c}; h^{[2]}) = 1.$$ 

Its separate pre-training minimizes the total negative log-likelihood $J_o(W_o)$ of the known training classes, appended with the negative weight penalties:

$$J_o(W_o) = -\frac{1}{K} \sum_{k=1}^{K} \log p(c_k; W_{o,c_k}) + \alpha \sum_{c=1}^{2} \sum_{j=1}^{s_j} w_{o,c,j} \quad (7)$$
Finally, the entire stacked NCAE classifier (SNCAE) is fine-tuned on the labeled training data by the conventional error back-propagation through the network and penalizing only the negative weights of the softmax layer. In the performed experiments, the network was trained and tested based on a leave-one-out scenario, so that the 53 (by the number of subjects) test accuracies were averaged to estimate the overall accuracy of the classifier. These experiments were conducted for different structures and parameters of the classifier. At this point, the two-layer SNCAE classifier with the following parameters: $s_1 = 50$, $s_2 = 5$, $s^0 = 2$, $\alpha = 3 * 10^{-5}$, $\beta = 3$, and $\gamma = 0.1$, is considered to give the best accuracy and was accepted as the final choice. Algorithm summarizes classification of kidney transplant status and generation of color ADC maps.

Algorithm 3 Kidney Transplant Status Classification and ADC Color Mapping

1. Calculate, using Equation (3), the ADCs at different $b$-values for the entire transplanted kidney of each subject.

2. **Classification:**
   A. Construct the CDFs of the calculated ADCs over the entire kidney volume at different $b$-values.
   B. Use a SNCAE-based deep ANN classifier trained by unsupervised pre-training and supervised fine tuning together with a leave-one-subject-out approach to discriminate rejection from non-rejection status and get the final diagnosis.

3. **Generation of color ADC maps:** Generate voxel-wise color-coded maps of the ADCs calculated in Step 1 to demonstrate visually perceived differences between the rejection and non-rejection states of kidney transplants at different $b$-values.
C. Experimental Results

1. DW-MRI Data Collection

   The proposed CAD system has been tested on DW-MRI data that has been collected from 53 subjects (44 men and 9 women with ages between 12 and 54 years old, having a mean age of $26 \pm 10$ years). Both the rejection (37 subjects) and non-rejection (16 subjects) groups, as a part of routine medical care after transplantation, were assessed with serum creatinine laboratory values. The patient who subsequently underwent an ultrasound-guided needle biopsy was examined, based on their clinical indication, as the gold standard. The DW-MR images were acquired before any biopsy procedure using a 1.5 $T$ scanner (SIGNA Horizon, General Electric Medical Systems, Milwaukee, WI). Coronal DW-MR images have been obtained by using a body coil and a gradient multi-shot spin-echo echo-planar sequence (TR/TE, 8000/61.2; bandwidth, 142 kHz; matrix, $125 \times 125$ mm$^2$; section thickness, 4 mm; intersection gap, 0 mm; FOV, 32 cm; signals acquired, 7; water signals acquired at different $b$-values of ($b_0$, $b_{50}$, $b_{100}$, $b_{200}$, $b_{300}$, $b_{400}$, $b_{500}$, $b_{600}$, $b_{700}$, $b_{800}$, $b_{900}$, and $b_{1000}$) s/mm$^2$). Approximately 50 sections have been obtained in 60 - 120 s to cover the whole kidney.

2. Segmentation Results

   Since the segmentation is an important step in developing any CAD system to assess renal function, the performance of the proposed segmentation was tested first on the collected DW-MRI data. Figure 16 shows some segmentation results for different kidney cross-sections (coronal, axial, and sagittal) for three subjects at $b_0$. The segmentation accuracy was evaluated by two volumetric metrics, namely, the Dice similarity coefficient (DSC) [175, 176] and absolute kidney volume difference (AKVD) and one distance-based metric – the 95-percentile modified Hausdorff distance (MHD) [177], which characterize the spatial overlap and distribution of the surface to surface distances between the
segmented and ground truth kidneys, respectively. The ground truth kidney maps were manually outlined by an MRI expert. For completeness, these metrics are detailed in Appendix A.

FIGURE 16: The proposed model segmentation (red) with respect to the expert’s manual ground truth (green): coronal (left column), axial (middle column), and sagittal (right column) cross sections for three different subjects, S₁, S₂, and S₃.

As shown in Table 1, high accuracy of the developed segmentation method is confirmed by means of the DSC, MHD, and AKVD statistics for all the test data sets. Moreover, the accuracy of the proposed segmentation technique was compared against three other methods: the level-sets approach by Chan and Vese [2] (CV), the level-sets guided by image intensity only, and the level-sets guided by combined intensity and spatial features.
TABLE 1: Segmentation accuracy of the proposed segmentation method using DSC, MHD(mm), and AKVD(%). All metrics are represented as minimum (Min), maximum (Max), and mean± standard deviation (SD).

<table>
<thead>
<tr>
<th>Evaluation Metrics</th>
<th>DSC</th>
<th>MHD(mm)</th>
<th>AVKD(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Min</td>
<td>0.85</td>
<td>4.00</td>
<td>8.00</td>
</tr>
<tr>
<td>Max</td>
<td>0.96</td>
<td>20.00</td>
<td>25.00</td>
</tr>
<tr>
<td>mean±SD</td>
<td>0.92 ± 0.02</td>
<td>6.20 ± 2.00</td>
<td>15.00 ± 3.00</td>
</tr>
</tbody>
</table>

TABLE 2: Segmentation accuracy of the proposed segmentation technique against three other level-sets methods using DSC, MHD(mm), and AKVD(%). All metrics are represented as mean± standard deviation (SD).

<table>
<thead>
<tr>
<th>Evaluation Metrics</th>
<th>DSC</th>
<th>MHD</th>
<th>AVKD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean±SD</td>
<td>0.92 ± 0.02</td>
<td>6.20±2.00</td>
<td>15.00±3.00</td>
</tr>
<tr>
<td>P-value</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Method</th>
<th>DSC</th>
<th>MHD</th>
<th>AVKD</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Proposed</td>
<td>0.92 ± 0.02%</td>
<td>6.20±2.00%</td>
<td>15.00±3.00%</td>
<td>—</td>
</tr>
<tr>
<td>CV [2]</td>
<td>0.69 ± 0.11%</td>
<td>75.97±11.96%</td>
<td>46.05±13.18%</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>I Only</td>
<td>0.54 ± 0.07%</td>
<td>29.14±5.34%</td>
<td>62.33±7.12%</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>I+S</td>
<td>0.63 ± 0.05%</td>
<td>22.79±7.36%</td>
<td>53.82±5.35%</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

using the same aforementioned segmentation evaluation metrics. Table 2 shows that the advantage of the developed segmentation approach is statistically significant with respect to the other methods, evidenced by the P-values less than 0.05, which confirms the high accuracy of the proposed segmentation techniques.

Figure 17 shows 3D results of the proposed segmentation method for three subjects along with their evaluation metrics. In particular, the developed segmentation technique proved its ability to precisely segment the kidney at higher $b_i$ values.

Figure 18 shows more coronal cross-sectional segmentation results for three different subjects at $b_0$ and higher $b_i$ values ($b_{500}$ and $b_{1000}$) which in turn emphasize the high
FIGURE 17: The proposed 3D segmentation (red) with respect to the expert’s manual ground truth (green) for three subjects with the associated DSC, MHD, and AKVD accuracy scores.

accuracy and robustness to noise of the proposed kidney segmentation technique.

FIGURE 18: Four more coronal cross-sections (columns) for the proposed kidney segmentation from DW-MRI acquired at different $b$-values $s/mm^2$.
FIGURE 19: Comparative cross-sectional segmentation results for our approach, the traditional CV [2] level-set, the level-set guided by intensity alone, and intensity and spatial, respectively, in rows for three different types of cross sections (coronal cross section in the first column, axial cross section in the second column, and sagittal cross section in the third column) for one subject at $b_0$ s/mm$^2$. The model segmentation is shown in red with respect to the manual ground truth (green) from an expert.

The comparative accuracy of the proposed approach versus the CV method on representative data at different types of kidney cross sections (i.e., axial, sagittal, and coronal)
FIGURE 20: A sample coronal cross-sectional kidney segmentation for DW-MRI data acquired at $b_{300}$ s/mm$^2$ (first row), $b_{500}$ s/mm$^2$ (second row), $b_{700}$ s/mm$^2$ (third row), and $b_{1000}$ s/mm$^2$ (fourth row) for (a) the proposed approach; (b) the CV [2] approach; (c) the level-set guided by intensity only; and (d) intensity and spatial.

for one selected subject acquired at $b_0$ is shown in Figure 19.

In order to assess the renal function using DW-MRI-based CAD systems, which in turn has gained increased attention in recent years [178], the estimation of diffusion parameters (e.g., ADC) requires the accurate segmentation of the kidney on DW-MRI data acquired at both higher and lower b-values. However, the accurate segmentation of kidney volumes at higher b-values is a challenge compared with those at lower b-values because of the decreased contrast between the object and the background. In spite of the aforementioned challenge, the proposed segmentation technique extracts accurately the kidney from
diffusion data at higher b-values compared to the other three aforementioned methods as shown in Figure 20. In contrast to the existing DW-MRI approaches, the integration of the 3D appearance, shape, and spatial features increases the robustness of the proposed approach to overcome large image noise at higher b-values. Figure 20 demonstrates a sample coronal cross section segmentation for four subjects at b-values of 300, 500, 700 and 1000 $s/mm^2$ for the developed approach and the three other approaches used before. As shown in this figure, the proposed approach produces precise segmentation of the kidney at higher b-values compared with the other methods with respect to the ground truth segmentation.

3. Diagnostics Results

Following kidney segmentation, the developed CAD system classifies the transplant status with the SNCAE-based classifier and CDFs as discriminatory features. A leave-one-subject-out classification scenario applied to distinguish between the rejection and non-rejection cases from the CDFs of the ADCs for different b-values has correctly classified 98% of the cases, namely, 36 out of the 37 rejected and 16 out of the 16 non-rejected kidney transplants.

It is worth mentioning that fusing all the CDFs of the ADCs calculated at the 11 different b-values, totally improves the final diagnostic accuracy, as shown in Table 3, and helps to overcome the challenge of possible artifacts due to chemical shifts, which could occur at one or two b-values, if the artifacts exist.

TABLE 3: Diagnostic accuracy based on the input CDF of the ADC for individual b-values and the fused CDFs of all b-values.

<table>
<thead>
<tr>
<th>Classification Accuracy</th>
<th>$b_{50}$</th>
<th>$b_{100}$</th>
<th>$b_{200}$</th>
<th>$b_{300}$</th>
<th>$b_{400}$</th>
<th>$b_{500}$</th>
<th>$b_{600}$</th>
<th>$b_{700}$</th>
<th>$b_{800}$</th>
<th>$b_{900}$</th>
<th>$b_{1000}$</th>
<th>Fused b-values</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>75%</td>
<td>85%</td>
<td>75%</td>
<td>81%</td>
<td>70%</td>
<td>77%</td>
<td>62%</td>
<td>60%</td>
<td>64%</td>
<td>62%</td>
<td>74%</td>
<td>98%</td>
</tr>
</tbody>
</table>

In order to evaluate the effect of the CDFs encoding step ($\Delta$) on the overall accuracy, the CDFs of the 3D ADCs were constructed using two different $\Delta_i$ ($\Delta = 0.02$ and 0.04).
Then, the SNCAE classifier was applied on the constructed CDFs and the results are shown in Table 4. As demonstrated in Table 4, the overall accuracy, sensitivity, and specificity, have been greatly reduced. This can be explained in part by the fact that increasing the value of $\Delta$ results in losing important data information, thus making the data not well-presented, which in turn affects the classifier performance.

**TABLE 4: Diagnostic accuracy, sensitivity, and specificity for the developed CAD system with the SNCAE classifier using different CDF encoding steps ($\Delta_i$).**

<table>
<thead>
<tr>
<th>$\Delta_i$:</th>
<th>Accuracy</th>
<th>Sensitivity</th>
<th>Specificity</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\Delta_1 = 0.01$</td>
<td>98%</td>
<td>97%</td>
<td>100%</td>
</tr>
<tr>
<td>$\Delta_2 = 0.02$</td>
<td>85%</td>
<td>86%</td>
<td>81%</td>
</tr>
<tr>
<td>$\Delta_3 = 0.04$</td>
<td>83%</td>
<td>84%</td>
<td>81%</td>
</tr>
</tbody>
</table>

Furthermore, the effect of changing the SNCAE structure on the overall accuracy has been investigated by using different SNCAE layouts (different number of hidden layers ($l$) and hidden nodes at each layer ($s_l$)). From the results in Table 5, the network structure with two hidden layers $s_1 = 50$ and $s_2 = 5$, demonstrated the highest accuracy.

**TABLE 5: Diagnostic accuracy, sensitivity, and specificity for the developed CAD system with the SNCAE classifier using different structures, i.e., different number of hidden layers ($l$) and hidden nodes at each layer ($s_l$), using the same input size of 1100 (11 CDFs each of 100 region), $\alpha = 3 \times 10^{-5}$, $\beta = 3$, and $\gamma = 0.1$.**

<table>
<thead>
<tr>
<th>SNCAE Structure:</th>
<th>Accuracy</th>
<th>Sensitivity</th>
<th>Specificity</th>
</tr>
</thead>
<tbody>
<tr>
<td>$s_1 = 5$</td>
<td>85%</td>
<td>84%</td>
<td>88%</td>
</tr>
<tr>
<td>$s_1 = 25$</td>
<td>60%</td>
<td>68%</td>
<td>44%</td>
</tr>
<tr>
<td>$s_1 = 50$</td>
<td>68%</td>
<td>70%</td>
<td>63%</td>
</tr>
<tr>
<td>$s_1 = 50$ and $s_2 = 5$</td>
<td>98%</td>
<td>97%</td>
<td>100%</td>
</tr>
<tr>
<td>$s_1 = 50$, $s_2 = 25$, and $s_3 = 5$</td>
<td>77%</td>
<td>92%</td>
<td>44%</td>
</tr>
</tbody>
</table>
In addition to the leave-one-subject-out approach, a four-fold cross-validation test has been performed where 75% of the data was used for training and the other 25% for testing, to further validate and justify the performance of the SNCAE classifier. As documented in Table 6, the diagnostic accuracy of the combined SNCAE classification system is almost independent of the choice of the training and testing data sets. The four-fold cross-validation experiment demonstrated an average accuracy of 96%.

<table>
<thead>
<tr>
<th>Cross-validating SNCAE classifier</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Testing group (25%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Correct/All</td>
<td>12/13</td>
<td>12/13</td>
<td>13/13</td>
<td>14/14</td>
</tr>
<tr>
<td>Accuracy</td>
<td>92%</td>
<td>92%</td>
<td>100%</td>
<td>100%</td>
</tr>
<tr>
<td>Average Accuracy</td>
<td></td>
<td></td>
<td></td>
<td>96%</td>
</tr>
</tbody>
</table>

To evaluate capabilities of the SNCAE classifier, it has been compared with seven well-known learnable classifiers from the Weka collection [1]: K*, IBK, Naive Bayes tree (NBT), Multi-class classifier (MCC), Decorate, Random tree (RT), and Random forest (RF). Table 7 presents their and the presented diagnostic accuracy in terms of the numbers of correctly classified rejection and non-rejection cases with respect to the overall numbers of subjects, sensitivity, and specificity. The SNCAE classifier demonstrated the best total diagnostic accuracy of 98% with 100 % specificity, or 16 correctly classified non-rejected transplants out of the 16 subjects, and 97% sensitivity, or 36 correctly classified rejected transplants out of the 37 subjects.

In addition, the receiver operating characteristics (ROC) (see Appendix .A) for the developed CAD system with SNCAE and the chosen seven Weka classifiers in Figure 21 have been constructed to test the performance. As shown in Figure 21, the area under the curve (AUC) is the highest for the SNCAE classifier and approaches the top-most unit value (see Table 7). These initial diagnostic results confirm that the proposed CAD system holds promise as a reliable non-invasive diagnostic tool.

Together with the automated classification, the proposed CAD system also provides
TABLE 7: Diagnostic accuracy in terms of correctly classified vs. true non-rejection (NR) and rejection (R) cases, sensitivity, specificity, and AUC for the proposed CAD system with the SNCAE classifier and seven classifiers from the Weka collection [1].

<table>
<thead>
<tr>
<th>Classification</th>
<th>NR</th>
<th>R</th>
<th>Accuracy</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>AUC</th>
</tr>
</thead>
<tbody>
<tr>
<td>K*</td>
<td>12/16</td>
<td>30/37</td>
<td>79%</td>
<td>81%</td>
<td>75%</td>
<td>0.82</td>
</tr>
<tr>
<td>IBK</td>
<td>15/16</td>
<td>36/37</td>
<td>96%</td>
<td>97%</td>
<td>94%</td>
<td>0.96</td>
</tr>
<tr>
<td>NBT</td>
<td>11/16</td>
<td>30/37</td>
<td>77%</td>
<td>81%</td>
<td>69%</td>
<td>0.79</td>
</tr>
<tr>
<td>MCC</td>
<td>15/16</td>
<td>34/37</td>
<td>92%</td>
<td>92%</td>
<td>94%</td>
<td>0.94</td>
</tr>
<tr>
<td>Decorate</td>
<td>8/16</td>
<td>34/37</td>
<td>79%</td>
<td>92%</td>
<td>50%</td>
<td>0.80</td>
</tr>
<tr>
<td>RT</td>
<td>10/16</td>
<td>29/37</td>
<td>74%</td>
<td>78%</td>
<td>63%</td>
<td>0.74</td>
</tr>
<tr>
<td>RF</td>
<td>9/16</td>
<td>35/37</td>
<td>83%</td>
<td>95%</td>
<td>56%</td>
<td>0.82</td>
</tr>
<tr>
<td>SNCAE</td>
<td>16/16</td>
<td>36/37</td>
<td>98%</td>
<td>97%</td>
<td>100%</td>
<td>0.97</td>
</tr>
</tbody>
</table>

FIGURE 21: ROC curves and their AUC for SNCAE and Weka classifiers [1].

voxel-wise parametric ADC maps, which can help in local visual assessment of the transplanted kidney. The color-coded ADC map, which depicts the estimated voxel-wise ADCs of Equation (3), is more informative than the ADC CDFs collected over the entire kidney.
and thus hides local peculiarities of spatial behavior of the ADCs. Examples of the ADC maps for two non-rejection and two acute rejection cases for the DW-MRI at the different $b$-values in Figure 22 demonstrate some expected relations between the local ADCs, which could be helpful for assessing the transplant and detecting its non-rejection or rejection status.

FIGURE 22: Pixel-wise parametric maps for DW-MRI at different $b$-values ($b_{50}$ to $b_{1000}$) $s/mm^2$ and their average value for two non-rejection ($S_1$, $S_2$) and two rejection ($S_3$, $S_4$) subjects.

D. Chapter Summary

The developed CAD system for early detection of renal transplant rejection from 4D DW-MRI data combines existing and new techniques for non-rigid image alignment, kidney segmentation with a deformable boundary, estimation of spatial diffusion parameters (ADCs), and an SNCAE classification of the transplanted kidney status using CDFs of the ADCs as integral status descriptions. In a test on a biopsy-proven cohort of 53 participants, the developed system showed an overall accuracy of 98% in detecting rejected and non-rejected kidney transplants. These experimental results make the proposed non-invasive framework a reliable early renal diagnostic tool. In the future, the test sets of both non-
rejection and rejection kidney transplants will be increased in order to further validate the accuracy and robustness of the proposed framework in both segmentation of the DW-MRI and diagnosis. Also, new kidney transplant data sets, which are acquired at lower $b$-values, will be used to explore the ability of the proposed framework to determine the type of kidney transplant rejection, such as anti-body mediated rejection, T-cell or cellular rejection, or other causes of acute kidney dysfunction such as drug toxicity and viral infection.
CHAPTER III
CONCLUSIONS AND FUTURE WORK

This thesis has presented a novel computer-aided diagnostic (CAD) system coupled with diffusion-weighted magnetic resonance imaging (DW-MRI) for the early detection of acute renal rejection after transplantation. Fortunately, the presented work in this thesis confirms the power of DW-MRI as an emerging imaging modality, with great thanks to its ability to provide both anatomical and functional information about both original and transplanted kidneys without exposing patients to any radiation, and with no need for contrast agents administration like the DCE-MRI. Therefore, a fully automated CAD system for early detection of acute renal transplant rejection was coupled with DW-MRI utilizing the aforementioned merits of DW-MRI. The developed CAD system for early detection of renal transplant rejection from 4D DW-MRI data combines existing and new techniques for non-rigid image alignment, kidney segmentation with a deformable boundary, estimation of functional diffusion parameters, called apparent diffusion coefficients (ADCs), and a stacked nonnegativity constrained autoencoder (SNCAE) classification of the transplanted kidney status using cumulative distribution functions (CDFs) of the ADCs as integral status descriptions.

A. Summary of Contributions

The main findings and contributions of this thesis can be summarized as follows:

- A new fully-automated computer-aided diagnostic (CAD) system using 4D (3D + $b$-value) diffusion-weighted magnetic resonance imaging (DW-MRI) for early determination of the transplanted kidney status as nonrejection or rejection.

- Fusion of the apparent diffusion coefficients (ADCs) at multiple low and high gra-
dient field strengths and duration (\(b\)-values) estimated from the segmented kidneys (subjects) using the proposed segmentation approach. The segmentation is done after a non-rigid image alignment for all subjects to a single reference subject.

- Exploring new discriminatory features to distinguish between nonrejected and rejected kidney transplants. These features are based on constructing cumulative distribution functions (CDFs) from the estimated ADCs at different \(b\)-values form \(b_0\) to \(b_{1000}\) \(s/mm^2\) for all segmented kidneys. It is worth mentioning that, the use of those CDFs instead of the voxel-wise ADCs of the entire kidney volume helps in making the data well-presented and mainly solves three important challenges of different kidney volumes: (i) loss of important information by data truncation for large kidney volumes; (ii) addition of nonexisting information by zero padding for small kidney volumes; and (iii) considerable time expenditures for training and classification for large kidney volumes.

- The use of stacked autoencoders with nonnegativity constraint (SNCAE) classifier for the purpose of discriminating nonrejected from rejected transplanted kidneys. After fusing all the constructed CDFs at different \(b\)-values for each subject, the SNCAE is trained and tested with these fused CDFs for all subjects by using a leave-one-subject-out approach and a four-fold cross-validation scenario as well.

B. Future Avenues

Several future trends that can be deeply investigated include but are not limited to the following avenues:

- Exploiting a new segmentation technique using a nonnegative matrix factorization (NMF)-guided active contour model to enhance and improve the segmentation accuracy. Some initial promising segmentation results have been obtained [179]

- After differentiating rejection from nonrejection kidney transplants, the next step is to obtain new DW-MRI data volumes at lower \(b\)-values and to investigate in-depth
the ability of the proposed CAD system to differentiate between different types of rejection (e.g., T-cell mediated rejection, anti-body mediated rejection, immunosuppressive toxicity, and viral infection) as it is a very important step for therapeutic purposes and for the determination of the appropriate treatment. Preliminary results for differentiating between different types of rejection are preferably found in [180].

- A new trend to be investigated is considering the fusion of multiple discriminating features extracted from images (e.g., CDFs of ADCs) with clinical data such as creatinine clearance (CrCl) and serum plasma creatinine (SPCR) to enhance and support the classification process with the clinical data as well, which in turn can provide a meaningful coupling of information provided by imaging and clinical information. Please see [181] for some initial results.

- Constructing a new two-cascaded stages CAD system utilizing the fusion of image-based with clinical-based biomarkers with the ability to differentiate nonrejection kidney transplants from transplanted kidneys with abnormalities (graft dysfunction) in the first stage. Then, classifying these abnormal kidneys as kidneys with early rejection and kidneys with other diseases (e.g., tubular inflammation, acute tubular injury, graft amyloidosis, etc.). Preliminary results of the suggested idea can be found in [181].

- Conducting a new study to investigate the ability of the proposed segmentation approach to extract the prostate from the surrounding tissues using DCE-MRI and DW-MRI [182–195].

- Extending the developed segmentation approach to segment the heart left/right ventricle wall by using contrast enhanced cardiac magnetic resonance images [196–218].

- Exploring the ability of the proposed CAD system to detect other organ diseases like lung cancer using contrast enhanced computed tomography (CECT) images [219–267].
• Investigating the capability of the developed CAD system to detect brain disorders and abnormalities (e.g., dyslexia, autism, etc.) [159, 268–311].
REFERENCES


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APPENDIX

A. Evaluating Segmentation Accuracy

Performance of the proposed segmentation was evaluated by using three accuracy metrics: (i) the Dice similarity coefficient (DSC) [175, 176], (ii) the 95-percentile modified Hausdorff distance (MHD) [177], and (iii) the absolute kidney volume difference (AKVD), which are detailed below.

The *Dice similarity coefficient* (DSC), shown in Figure 23, measures the relative overlap between the segmented and ground truth kidney in terms of true positive ($TP$), false positive ($FP$), and false negative ($FN$) absolute volumes of the correctly segmented kidney voxels, background voxels segmented as the kidney, and kidney voxels segmented as the background, respectively:

$$DS C = \frac{2 TP}{2 TP + FP + FN}; \quad 0 \leq DS C \leq 1$$

FIGURE 23: 2D illustration of segmentation errors calculation between the segmented and ground truth objects for the DSC determination.

The higher the DSC, the better the segmentation (zero and unit DSC indicate no overlap and ideal overlap, respectively).
The *modified Hausdorff distance* (MHD), shown in Figure 24, measures the largest surface-to-surface distance between the segmented and ground truth kidney borders. The conventional asymmetric Hausdorff distance (HD) [177] from a set of points $A_1$ to a set of points $A_2$ is defined as the maximum Euclidean distance $d(p_1, p_2)$ between the points $p_1$ of $A_1$ to their nearest points $p_2$ in $A_2$:

$$\text{HD}(A_1, A_2) = \max_{p_1 \in A_1} \left\{ \min_{p_2 \in A_2} \{d(p_1, p_2)\} \right\} \quad (A-2)$$

Generally $\text{HD}(A_1, A_2) \neq \text{HD}(A_2, A_1)$. Therefore, the symmetric bidirectional HD (BHD) is defined as $\text{BHD}(A_1, A_2) = \max \{\text{HD}(A_1, A_2), \text{HD}(A_2, A_1)\}$.

![FIGURE 24: 2D schematic illustration for the HD calculation.](image)

To escape possible outliers, affecting both the HD and BHD, and measure more robustly the segmentation accuracy, the 95-percentile BHD, called the modified HD (MHD), is used in this paper. In this case, the maximum distance in Equation (A-2) is replaced with the 95-percentile of all the point-to-point distances $d(p_1, p_2)$.

The *absolute kidney volume difference* (AKVD), shown in Figure 25, measures the relative volumetric difference between the segmented and ground truth kidney (their absolute volumes are equal to $V_{\text{segm}} = TP + FN$ and $V_{\text{true}} = TP + FP$, respectively):

$$V_{\text{segm}} = TP + FN$$
$$V_{\text{true}} = TP + FP$$
The receiver operating characteristic (ROC) [312] is an alternative metric to further test the classification accuracy and robustness of a CAD system compared to other classifiers. The sensitivity and discriminability of a classifier is evaluated in a Cartesian plane of relative true positive and false positive rates by its ROC curve for different operating points, e.g., various decision thresholds. The area under the ROC curve (AUC) characterizes the classification accuracy, namely, the probability of the correct renal transplant status detection for a randomly chosen pair of patients.
B. List of Abbreviations

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Full Definition</th>
</tr>
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<tbody>
<tr>
<td>CKD</td>
<td>Chronic Kidney Disease</td>
</tr>
<tr>
<td>NKF</td>
<td>National Kidney Foundation</td>
</tr>
<tr>
<td>GFR</td>
<td>Glomerular Filtration Rate</td>
</tr>
<tr>
<td>MRI</td>
<td>Magnetic Resonance Imaging</td>
</tr>
<tr>
<td>UNOS</td>
<td>United Network of Organ Sharing</td>
</tr>
<tr>
<td>AKR</td>
<td>Acute Kidney Rejection</td>
</tr>
<tr>
<td>ATN</td>
<td>Acute Tubular Necrosis</td>
</tr>
<tr>
<td>ITox</td>
<td>Immunosuppressive Toxicity</td>
</tr>
<tr>
<td>VI</td>
<td>Viral Infection</td>
</tr>
<tr>
<td>CT</td>
<td>Computed Tomography</td>
</tr>
<tr>
<td>US</td>
<td>Ultrasound</td>
</tr>
<tr>
<td>PI</td>
<td>Pulsatility Index</td>
</tr>
<tr>
<td>RI</td>
<td>Resistance Index</td>
</tr>
<tr>
<td>PD</td>
<td>Power Doppler</td>
</tr>
<tr>
<td>CD</td>
<td>Color Doppler</td>
</tr>
<tr>
<td>CE</td>
<td>Contrast Enhanced</td>
</tr>
<tr>
<td>IGF</td>
<td>Immediate Graft Function</td>
</tr>
<tr>
<td>SGF</td>
<td>Slow Graft Function</td>
</tr>
<tr>
<td>DGF</td>
<td>Delayed Graft Function</td>
</tr>
<tr>
<td>DKT</td>
<td>Dual Kidney Transplantation</td>
</tr>
<tr>
<td>SKT</td>
<td>Single Kidney Transplantation</td>
</tr>
<tr>
<td>AR</td>
<td>Acute Rejection</td>
</tr>
<tr>
<td>ROI</td>
<td>Regions of Interest</td>
</tr>
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<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Full Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>CP</td>
<td>Cortical perfusion</td>
</tr>
<tr>
<td>SCr</td>
<td>Serum Creatinine</td>
</tr>
<tr>
<td>USCM</td>
<td>Ultrasound Contrast Media</td>
</tr>
<tr>
<td>CEUS</td>
<td>Contrast Enhanced Ultrasound</td>
</tr>
<tr>
<td>CES</td>
<td>Contrast Enhanced Sonography</td>
</tr>
<tr>
<td>CDUS</td>
<td>Contrast Enhanced UltraSonography</td>
</tr>
<tr>
<td>RBF</td>
<td>Renal Blood Flow</td>
</tr>
<tr>
<td>AKI</td>
<td>Acute Kidney Injury</td>
</tr>
<tr>
<td>TIC</td>
<td>Time-Intensity Curves</td>
</tr>
<tr>
<td>ATPI</td>
<td>Arrival Time Parametric Imaging</td>
</tr>
<tr>
<td>IT</td>
<td>Inflow Time</td>
</tr>
<tr>
<td>RisT</td>
<td>Rising Time</td>
</tr>
<tr>
<td>AT</td>
<td>Arrival Time</td>
</tr>
<tr>
<td>TTP</td>
<td>Time to Peak</td>
</tr>
<tr>
<td>DCE-MRI</td>
<td>Dynamic Contrast Enhanced MRI</td>
</tr>
<tr>
<td>RBF</td>
<td>Renal Plasma Flow</td>
</tr>
<tr>
<td>NGF</td>
<td>Normalized Gradient Field</td>
</tr>
<tr>
<td>LS-SVM</td>
<td>Least Square Support Vector Machine</td>
</tr>
<tr>
<td>MTT</td>
<td>Mean Transit Time</td>
</tr>
<tr>
<td>BOLD-MRI</td>
<td>Blood Oxygen Level Dependant MRI</td>
</tr>
<tr>
<td>R2*</td>
<td>Apparent Relaxation Rate</td>
</tr>
<tr>
<td>CR2*</td>
<td>Cortical R2*</td>
</tr>
<tr>
<td>MR2*</td>
<td>Medullary R2*</td>
</tr>
<tr>
<td>MCR2*</td>
<td>Medullary to Cortical R2*</td>
</tr>
<tr>
<td>SNR</td>
<td>Signal-to-Noise Ratio</td>
</tr>
</tbody>
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<table>
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<th>Full Definition</th>
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<tbody>
<tr>
<td>DW-MRI</td>
<td>Diffusion-Weighted MRI</td>
</tr>
<tr>
<td>ADC</td>
<td>Apparent Diffusion Coefficient</td>
</tr>
<tr>
<td>CRCL</td>
<td>Creatinine Clearance</td>
</tr>
<tr>
<td>IniGF</td>
<td>Initial Graft Function</td>
</tr>
<tr>
<td>ARTR</td>
<td>Acute Renal Transplant Rejection</td>
</tr>
<tr>
<td>TD</td>
<td>True Diffusion</td>
</tr>
<tr>
<td>ACR</td>
<td>Acute Cellular Rejection</td>
</tr>
<tr>
<td>CAD</td>
<td>Computer-Aided Diagnostic</td>
</tr>
<tr>
<td>SPCR</td>
<td>Serum Plasma Creatinine</td>
</tr>
<tr>
<td>CDF</td>
<td>Cumulative Distribution Function</td>
</tr>
<tr>
<td>NCAE</td>
<td>Non-negativity Constrained Autoencoder</td>
</tr>
<tr>
<td>NMF</td>
<td>Nonnegative Matrix Factorization</td>
</tr>
<tr>
<td>SVM</td>
<td>Support Vector Machine</td>
</tr>
<tr>
<td>RVM</td>
<td>Relevant Vector Machine</td>
</tr>
<tr>
<td>MRF</td>
<td>Markov Random Field</td>
</tr>
<tr>
<td>MHD</td>
<td>Modified Hausdorff Distance</td>
</tr>
<tr>
<td>MGRF</td>
<td>Markov-Gibbs random field</td>
</tr>
<tr>
<td>FGM</td>
<td>Finite Gaussian Mixture</td>
</tr>
<tr>
<td>LCDG</td>
<td>Linear Combination of Discrete Gaussians</td>
</tr>
<tr>
<td>PDF</td>
<td>Probability Distribution Function</td>
</tr>
<tr>
<td>SNCAE</td>
<td>Stacked Non-negativity Constrained Autoencoder</td>
</tr>
<tr>
<td>ANN</td>
<td>Artificial Neural Network</td>
</tr>
<tr>
<td>RF</td>
<td>Random Forest</td>
</tr>
<tr>
<td>DT</td>
<td>Decision Tree</td>
</tr>
<tr>
<td>kNN</td>
<td>K-nearest Neighbor</td>
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Table 8 – continued from the previous page

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<tbody>
<tr>
<td>AE</td>
<td>Auto-encoder</td>
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<tr>
<td>KL</td>
<td>Kullback-Leibler</td>
</tr>
<tr>
<td>DSC</td>
<td>Dice Similarity Coefficient</td>
</tr>
<tr>
<td>AKVD</td>
<td>Absolute Kidney Volume Difference</td>
</tr>
<tr>
<td>BHD</td>
<td>Bidirectional Hausdorff Distance</td>
</tr>
<tr>
<td>NBT</td>
<td>Naive Bayes Tree</td>
</tr>
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<td>MCC</td>
<td>Multi-class Classifier</td>
</tr>
<tr>
<td>RT</td>
<td>Random Tree</td>
</tr>
<tr>
<td>ROC</td>
<td>Receiver Operating Characteristic</td>
</tr>
<tr>
<td>AUC</td>
<td>Area Under the Curve</td>
</tr>
</tbody>
</table>
CURRICULUM VITAE

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University of Louisville, Louisville, KY, USA
University E-mail: mnsheh01@louisville.edu
Personal E-mail: eng.mohamednazeem@hotmail.com
Office: (502) 852–4032
Cell: (502) 797–1805

Current Research Interest

I am a graduate student at Electrical and Computer Engineering Department at the University of Louisville, Louisville, KY, USA. My general research interests are in digital image processing, medical imaging, and computer vision. In August 2014, I have joined the BioImaging Lab and have actively been working on developing a novel computer-aided diagnostic (CAD) systems for early detection of acute renal transplant rejection from 4D diffusion-weighted MRIs. This research includes developing new computer aided diagnostic systems, image modeling, and image segmentation and registration.
**Education**

2016  M.Sc., Electrical and Computer Engineering Department, Speed School of Engineering, University of Louisville, Louisville, KY 40292

**M.Sc. Thesis:** *A Non-Invasive Diagnostic System for Early Assessment of Acute Renal Transplant Rejection* - Prof. Ayman S. El-Baz, Thesis Advisor.

GPA = 4.0 *(Graduation date: Summer 2016)*

2009  B.Sc., Computers Engineering and Control Systems, Mansoura University, Mansoura 35516, Egypt.

**Sr. Project:** *Remote Controlling of Moving Objects Via Mobile or Fixed Phones* - Prof. Aly El-Desouky, Project Supervisor.

GPA = 4.0 *(Graduated with excellence with honor and ranked the first in a class of 200)*

**Experience**

Fall 2014–Present  Graduate Research Assistant, BioImaging laboratory, Department of Bioengineering, University of Louisville, Louisville, KY 40292, USA.

2010–2014  Graduate Research Assistant, Department of Computers Engineering and Control Systems, Mansoura University, Mansoura 35516, Egypt.
Teaching/Administration Experience

Assist in teaching, laboratory demonstration, conducting tutorials, grading, and senior graduation projects of the following Electrical Engineering undergraduate courses:

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<td>Computer Maintenance</td>
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<tr>
<td>Mansoura University</td>
<td>Supervision support for an undergraduate final project (Smart Modern Automatic Control Restaurant trend) using PAC</td>
<td>2012-2013</td>
</tr>
<tr>
<td>Mansoura University</td>
<td>Summer trainer for undergraduate student on Matlab basic tools and PAC controller</td>
<td>2011-2012</td>
</tr>
<tr>
<td>Mansoura University</td>
<td>Supervision support for an undergraduate final project (Smart Gate) using image processing techniques; specifically, face recognition</td>
<td>2010-2011</td>
</tr>
</tbody>
</table>
Professional Affiliations and Memberships

- Young Professional Member, Institute of Electrical and Electronics Engineers (IEEE).
- Member, IEEE Signal Processing (SPS) Society.
- Member, IEEE Engineering in Medicine and Biology (EMBS) Society.
- Member, IEEE Biometrics Council.
- Member, IEEE Sensors Council.
- Member, IEEE Nanotechnology Council.
- Member, IEEE Electronic Design Automation Council.
- Member, IEEE Cloud Computing Community.
- Member, IEEE Systems Council.
- Member, Egyptian Engineers Syndicate, Electrical Engineering.

Awards and Recognition

- Merit of Excellence Certificate from Graduate Student Council (GSC) at the University of Louisville, in the area of community engagement in Graduate Student Regional Research Conference (GSRRC), April 2016.
- One of The Best 39 papers in the International Symposium on Biomedical Imaging: From Nano to Macro, Prague, Czech Republic, April 16–20, 2016 (ISBI’16).
- IEEE Signal Processing Society (SPS) Travel Award 2015 to attend the IEEE International Conference on Image processing, Quebec City, Canada, September 27–30, 2015 (ICIP’15).
- GSC at the University of Louisville Travel Award 2015.
- Theobald Scholarship Award in the Electrical and Computer Engineering Department, Speed school of Engineering, University of Louisville, 2015.
Higher Education Enhancement Project Fund Discretionary Certificate for distinct students in the Department of Communications and Electronics Engineering for Excellent grade ranked third in the second undergraduate year of education, Mansoura University, 2005–2006.

Class Work Grade

A total of 30 credit hours in Electrical Engineering and Bioengineering subjects with a cumulative GPA of 4. Particular course concentration has been in medical image analysis and machine learning.

Research Activities

- Image modeling, 2D and 3D image segmentation and registration.
- Development of computer-aided diagnostic (CAD) system using diffusion-weighted magnetic resonance imaging (DW-MRI) for the early assessment of acute renal transplant rejection.
- Development of a CAD system for the early detection of different types of kidney rejection using DW-MRIs.
- Development of a cascaded two-stage CAD system for differentiating nonrejection renal transplants from transplanted kidneys with abnormalities using DW-MRIs and then, classifying abnormal kidney transplants into early rejection and other kidney diseases including: tubular inflammation, acute tubular injury, graft amyloidosis, and acute tubular necrosis.
- Assisted in grants writing and preparing primary results for the BioImaging Lab, University of Louisville.
Out of Reach Connectivity

- I have trained one of the high school students during Fall and Spring of 2015 and we had two published conference papers.
- I have trained one of the middle school students during Fall and Spring of 2016 and we had one published conference paper.

Publications

During Fall 2014–Summer 2016, I have authored or co-authored 2 journal articles, 1 book chapter, 6 peer-reviewed conference papers, 1 abstract in proceedings. The first article have been submitted to Medical Image Analysis Journal (5-year impact factor 4.950); current status (under-review) and the second one to be submitted to The Egyptian Journal of Radiology and Nuclear Medicine. The conference papers were reported as top-rank international conferences in medical imaging, image processing, and pattern recognition e.g., MICCAI, ISBI, and ICIP with acceptance rate less than 30%.

- Journal Articles (Total = 2)

- Book Chapters (Total = 1)

- **Peer-Reviewed Conference Proceedings (Total = 6)**


5. **M. Shehata**, F. Khalifa, E. Hollis, A. Soliman, E. Hosseini-Asl, M. Abou El-Ghar,


- Abstracts Published in Proceedings (Total = 1)


- Patents and Disclosures (Total = 1)


Graduate Advisor

Dr. Ayman S. El-Baz, Department of Bioengineering, University of Louisville, KY.

Personal Skills, Hobbies, and Activities

- Active, self-motivated, ability to work alone and in a team, ability to work under pressure, Internet browsing, reading, swimming, and football.